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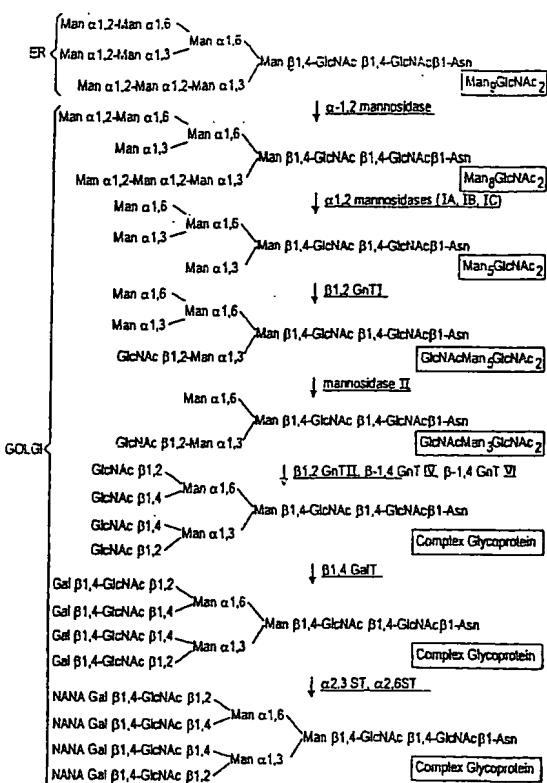
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(54) Title: PRODUCTION OF MODIFIED GLYCOPROTEINS HAVING MULTIPLE ANTENNAARY STRUCTURES



(57) Abstract: The present invention relates to eukaryotic host cells, especially lower eukaryotic host cells, having modified oligosaccharides which may be modified further by heterologous expression of a set of glycosyltransferases, sugar and sugar nucleotide transporters to become host-strains for the production of mammalian, e.g., human therapeutic glycoproteins. The process provides an engineered host cell which can be used to express and target any desirable gene(s) involved in glycosylation. Host cells with modified lipid-linked oligosaccharides are created or selected. N-glycans made in the engineered host cells exhibit GnTIII, GnTIV, GnTV, GnT VI or GnTIX activity, which produce bisected and/or multiantennary N-glycan structures and may be modified further by heterologous expression of one or more enzymes, e.g., glycosyltransferases, sugar, sugar nucleotide transporters, to yield human-like glycoproteins. For the production of therapeutic proteins, this method may be adapted to engineer cell lines in which any desired glycosylation structure may be obtained.

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**PRODUCTION OF MODIFIED GLYCOPROTEINS HAVING
MULTIPLE ANTENNARY STRUCTURES**

CROSS-REFERENCE TO RELATED APPLICATIONS

[0001] This application is a continuation-in-part of U. S. Application No. 5 10/680,963, filed on Oct. 7, 2003, which is a continuation-in-part of U. S. Application No. 10/371,877, filed on Feb. 20, 2003, which is a continuation-in-part of U. S. Application No. 09/892,591, filed June 27, 2001, which claims the benefit under 35 U.S.C. § 119(e) of U.S. Provisional Application No. 60/214,358, filed 10 June 28, 2000, U.S. Provisional Application No. 60/215,638, filed June 30, 2000, and U.S. Provisional Application No. 60/279,997, filed March 30, 2001, each of 15 which is incorporated herein by reference in its entirety. This application is also a continuation-in-part of PCT/US02/41510, filed on December 24, 2002, which claims the benefit of U. S. Provisional Application No. 60/344,169, filed on Dec. 27, 2001, each of which is incorporated herein by reference in its entirety.

15

FIELD OF THE INVENTION

[0002] The present invention is directed to methods and compositions by which non-human eukaryotic host cells, such as fungi or other eukaryotic cells, can be genetically modified to produce glycosylated proteins (glycoproteins) having 20 patterns of glycosylation similar to those of glycoproteins produced by animal cells, especially human cells, which are useful as human or animal therapeutic agents.

BACKGROUND OF THE INVENTION

Glycosylation Pathways in Humans and Lower Eukaryotes

[0003] After DNA is transcribed and translated into a protein, further post-translational processing involves the attachment of sugar residues, a process known as glycosylation. Different organisms produce different glycosylation enzymes (glycosyltransferases and glycosidases), and have different substrates (nucleotide sugars) available, so that the glycosylation patterns as well as composition of the individual oligosaccharides, even of the same protein, will be different depending on the host system in which the particular protein is being expressed. Bacteria typically do not glycosylate proteins, and if so only in a very unspecific manner (Moens and Vanderleyden (1997) *Arch Microbiol.* 168(3):169-175). Lower eukaryotes such as filamentous fungi and yeast add primarily mannose and mannosylphosphate sugars. The resulting glycan is known as a "high-mannose" type glycan or a mannan. Plant cells and insect cells (such as Sf9 cells) glycosylate proteins in yet another way. By contrast, in higher eukaryotes such as humans, the nascent oligosaccharide side chain may be trimmed to remove several mannose residues and elongated with additional sugar residues that typically are not found in the N-glycans of lower eukaryotes. See, e.g., Bretthauer, *et al.* (1999) *Biotechnology and Applied Biochemistry* 30:193-200; Martinet, *et al.* (1998) *Biotechnology Letters* 20:1171-1177; Weikert, *et al.* (1999) *Nature Biotechnology*, 17:1116-1121; M. Malissard, *et al.* (2000) *Biochemical and Biophysical Research Communications* 267:169-173; Jarvis, *et al.*, (1998) *Current Opinion in Biotechnology* 9:528-533; and Takeuchi (1997) *Trends in Glycoscience and Glycotechnology* 9:S29-S35.

[0004] Synthesis of a mammalian-type oligosaccharide structure begins with a set of sequential reactions in the course of which sugar residues are added and removed while the protein moves along the secretory pathway in the host organism. The enzymes which reside along the glycosylation pathway of the host organism or cell determine the resulting glycosylation patterns of secreted proteins. Thus, the resulting glycosylation pattern of proteins expressed in lower eukaryotic host cells differs substantially from the glycosylation pattern of proteins expressed

in higher eukaryotes such as humans and other mammals (Bretthauer, 1999). The structure of a typical fungal *N*-glycan is shown in **Figure 1A**.

[0005] The early steps of human glycosylation can be divided into at least two different phases: (i) lipid-linked Glc₃Man₉GlcNAc₂ oligosaccharides are assembled
5 by a sequential set of reactions at the membrane of the endoplasmic reticulum (ER)
(**Figure 13**) and (ii) the transfer of this oligosaccharide from the lipid anchor
dolichyl pyrophosphate onto *de novo* synthesized protein. The site of the specific
transfer is defined by an asparagine (Asn) residue in the sequence Asn-Xaa-
Ser/Thr (SEQ ID NOs:1 and 2) where Xaa can be any amino acid except proline
10 (Gavel and von Heijne (1990) *Protein Eng.* 3:433-42). Further processing by
glucosidases and mannosidases occurs in the ER before the nascent glycoprotein is
transferred to the early Golgi apparatus, where additional mannose residues are
removed by Golgi specific alpha (α)-1,2-mannosidases. Processing continues as
the protein proceeds through the Golgi. In the medial Golgi, a number of
15 modifying enzymes, including *N*-acetylglucosaminyl transferases (GnT_I, GnT_{II},
GnT_{III}, GnT_{IV} and GnT_V), mannosidase II and fucosyltransferases, add and
remove specific sugar residues. Finally, in the trans-Golgi, galactosyltransferases
(GalT) and sialyltransferases (ST) produce a glycoprotein structure that is released
from the Golgi. It is this structure, characterized by bi-, tri- and tetra-antennary
20 structures, containing galactose, fucose, N-acetylglucosamine and a high degree of
terminal sialic acid, that gives glycoproteins their human characteristics. The
structure of a typical human *N*-glycan is shown in **Figure 1B**. See also **Figures 14**
and **15** for steps involved in mammalian-type *N*-glycan processing.

[0006] In all eukaryotes studied to date, glycoproteins are derived from a
25 common lipid-linked oligosaccharide precursor Glc₃Man₉GlcNAc₂-dolichol-
pyrophosphate. Within the endoplasmic reticulum, synthesis and processing of
dolichol pyrophosphate bound oligosaccharides are identical between all known
eukaryotes. However, further processing of the core oligosaccharide by fungal
cells, *e.g.*, yeast, differs significantly from humans as it moves along the secretory
30 pathway.

[0007] In yeast, these steps are catalyzed by Golgi residing mannosyl-
transferases, like Och1p, Mnt1p and Mnn1p, which sequentially add mannose

sugars to the core oligosaccharide. The resulting structure is undesirable for the production of human-like proteins and it is thus desirable to reduce or eliminate mannosyltransferase activity. Mutants of *S. cerevisiae*, deficient in mannosyl-transferase activity (for example *och1* or *mnn9* mutants) have been shown to be
5 non-lethal and display reduced mannose content in the oligosaccharide of yeast glycoproteins. Other oligosaccharide processing enzymes, such as mannosylphosphate transferases, may also have to be eliminated depending on the host's particular glycosylation pathways.

10 Sugar Nucleotide Precursors

[0008] The *N*-glycans of animal glycoproteins typically include galactose, fucose, and terminal sialic acid. These sugars are not found on glycoproteins produced in yeast and filamentous fungi. In humans, the full range of nucleotide sugar precursors (e.g., UDP-*N*-acetylglucosamine, UDP-*N*-acetylgalactosamine, 15 CMP-*N*-acetylneuraminic acid, UDP-galactose, GDP-fucose, etc.) are synthesized in the cytosol and transported into the Golgi, where they are attached to the core oligosaccharide by glycosyltransferases. (Sommers and Hirschberg (1981) *J. Cell Biol.* 91(2):A406-A406; Sommers and Hirschberg (1982) *J. Biol. Chem.* 257(18):811-817; Perez and Hirschberg (1987) *Methods in Enzymology* 138:709-20 715).

[0009] Glycosyl transfer reactions typically yield a side product which is a nucleoside diphosphate or monophosphate. While monophosphates can be directly exported in exchange for nucleoside triphosphate sugars by an antiport mechanism, diphosphonucleosides (e.g., GDP) have to be cleaved by phosphatases (e.g. 25 GDPase) to yield nucleoside monophosphates and inorganic phosphate prior to being exported. This reaction is important for efficient glycosylation; for example, GDPase from *Saccharomyces cerevisiae* (*S. cerevisiae*) has been found to be necessary for mannosylation. However that GDPase has 90% reduced activity toward UDP (Berninsone *et al.* (1994) *J. Biol. Chem.* 269(1):207-211). Lower 30 eukaryotes typically lack UDP-specific diphosphatase activity in the Golgi since they do not utilize UDP-sugar precursors for Golgi-based glycoprotein synthesis. *Schizosaccharomyces pombe*, a yeast found to add galactose residues to cell wall

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10 **Sequential Processing of N-glycans by Compartmentalized Enzyme Activities**

[0010] Sugar transferases and glycosidases (*e.g.*, mannosidases) line the inner (luminal) surface of the ER and Golgi apparatus and thereby provide a “catalytic” surface that allows for the sequential processing of glycoproteins as they proceed through the ER and Golgi network. The multiple compartments of the cis, medial, 15 and trans Golgi and the trans-Golgi Network (TGN), provide the different localities in which the ordered sequence of glycosylation reactions can take place. As a glycoprotein proceeds from synthesis in the ER to full maturation in the late Golgi or TGN, it is sequentially exposed to different glucosidases, mannosidases and glycosyltransferases such that a specific carbohydrate structure may be synthesized. Much work has been dedicated to revealing the exact mechanism by 20 which these enzymes are retained and anchored to their respective organelle. The evolving picture is complex but evidence suggests that stem region, membrane spanning region and cytoplasmic tail, individually or in concert, direct enzymes to the membrane of individual organelles and thereby localize the associated catalytic 25 domain to that locus (*see, e.g.*, Gleeson (1998) *Histochem. Cell Biol.* 109:517-532).

[0011] In some cases, these specific interactions were found to function across species. For example, the membrane spanning domain of α 2,6-ST from rats, an enzyme known to localize in the trans-Golgi of the animal, was shown to also 30 localize a reporter gene (invertase) in the yeast Golgi (Schwientek *et al.* (1995) *J. Biol. Chem.* 270(10):5483-9). However, the very same membrane spanning domain as part of a full-length α 2,6-ST was retained in the ER and not further



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[0011] In some cases, these specific interactions were found to function across species. For example, the membrane spanning domain of α 2,6-ST from rats, an enzyme known to localize in the trans-Golgi of the animal, was shown to also 30 localize a reporter gene (invertase) in the yeast Golgi (Schwientek *et al.* (1995) *J. Biol. Chem.* 270(10):5483-9). However, the very same membrane spanning domain as part of a full-length α 2,6-ST was retained in the ER and not further

transported to the Golgi of yeast (Krezdorn *et al.* (1994) *Eur. J. Biochem.* 220(3):809-17). Full length GalT from humans was not even synthesized in yeast, despite demonstrably high transcription levels. In contrast, the transmembrane region of the same human GalT fused to an invertase reporter was able to direct 5 localization to the yeast Golgi, albeit it at low levels. Schwientek and co-workers have shown that fusing 28 amino acids of a yeast mannosyltransferase (*MNT1*), a region containing a cytoplasmic tail, a transmembrane region and eight amino acids of the stem region, to the catalytic domain of human GalT are sufficient for Golgi localization of an active GalT. Other galactosyltransferases appear to rely 10 on interactions with enzymes resident in particular organelles because, after removal of their transmembrane region, they are still able to localize properly.

[0012] Improper localization of a glycosylation enzyme may prevent proper functioning of the enzyme in the pathway. For example, *Aspergillus nidulans*, which has numerous α -1,2-mannosidases (Eades and Hintz (2000) *Gene* 255(1):25-34), does not add GlcNAc to Man₅GlcNAc₂ when transformed with the rabbit GnTI gene, despite a high overall level of GnTI activity (Kalsner *et al.* (1995) *Glycoconj. J.* 12(3):360-370). GnTI, although actively expressed, may be incorrectly localized such that the enzyme is not in contact with both of its substrates: UDP-GlcNAc and a productive Man₅GlcNAc₂ substrate (not all 15 20 25 30 35 40 45 50 55 60 65 70 75 80 85 90 95 100 105 110 115 120 125 130 135 140 145 150 155 160 165 170 175 180 185 190 195 200 205 210 215 220 225 230 235 240 245 250 255 260 265 270 275 280 285 290 295 300 305 310 315 320 325 330 335 340 345 350 355 360 365 370 375 380 385 390 395 400 405 410 415 420 425 430 435 440 445 450 455 460 465 470 475 480 485 490 495 500 505 510 515 520 525 530 535 540 545 550 555 560 565 570 575 580 585 590 595 600 605 610 615 620 625 630 635 640 645 650 655 660 665 670 675 680 685 690 695 700 705 710 715 720 725 730 735 740 745 750 755 760 765 770 775 780 785 790 795 800 805 810 815 820 825 830 835 840 845 850 855 860 865 870 875 880 885 890 895 900 905 910 915 920 925 930 935 940 945 950 955 960 965 970 975 980 985 990 995 1000 1005 1010 1015 1020 1025 1030 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fraction could be converted to $\text{Man}_5\text{GlcNAc}_2$ (less than about 5%, *see Chiba et al. WO 01/14522*).

[0013] To date, there is no reliable way of predicting whether a particular heterologously expressed glycosyltransferase or mannosidase in a lower eukaryote 5 will be (1), sufficiently translated (2), catalytically active or (3) located to the proper organelle within the secretory pathway. Because all three of these are necessary to affect glycosylation patterns in lower eukaryotes, a systematic scheme to achieve the desired catalytic function and proper retention of enzymes in the absence of predictive tools, which are currently not available, would be desirable.

10

Production of Therapeutic Glycoproteins

[0014] A significant number of proteins isolated from humans or animals are post-translationally modified, with glycosylation being one of the most significant modifications. An estimated 70% of all therapeutic proteins are glycosylated and 15 thus currently rely on a production system (*i.e.*, host cell) that is able to glycosylate in a manner similar to humans. Several studies have shown that glycosylation plays an important role in determining the (1) immunogenicity, (2) pharmacokinetic properties, (3) trafficking, and (4) efficacy of therapeutic proteins. It is thus not surprising that substantial efforts by the pharmaceutical industry have 20 been directed at developing processes to obtain glycoproteins that are as “humanoid” or “human-like” as possible. To date, most glycoproteins are made in a mammalian host system. This may involve the genetic engineering of such mammalian cells to enhance the degree of sialylation (*i.e.*, terminal addition of sialic acid) of proteins expressed by the cells, which is known to improve 25 pharmacokinetic properties of such proteins. Alternatively, one may improve the degree of sialylation by *in vitro* addition of such sugars using known glycosyltransferases and their respective nucleotide sugars (*e.g.*, 2,3-sialyltransferase and CMP-sialic acid).

[0015] While most higher eukaryotes carry out glycosylation reactions that are 30 similar to those found in humans, recombinant human proteins expressed in the above mentioned host systems invariably differ from their “natural” human counterpart (Raju *et al.* (2000) *Glycobiology* 10(5): 477-486). Extensive

development work has thus been directed at finding ways to improve the "human character" of proteins made in these expression systems. This includes the optimization of fermentation conditions and the genetic modification of protein expression hosts by introducing genes encoding enzymes involved in the formation 5 of human-like glycoforms. Goochee *et al.* (1999) *Biotechnology* 9(12):1347-55; Andersen and Goochee (1994) *Curr Opin Biotechnol.* 5(5):546-49; Werner *et al.* (1998) *Arzneimittelforschung.* 48(8):870-80; Weikert *et al.* (1999) *Nat Biotechnol.* 17(11):1116-21; Yang and Butler (2000) *Biotech. Bioeng.* 68:370-80. Inherent problems associated with all mammalian expression systems have not been solved.

10

Glycoprotein Production Using Eukaryotic Microorganisms

[0016] Although the core oligosaccharide structure transferred to a protein in the endoplasmic reticulum is basically identical in mammals and lower eukaryotes, substantial differences have been found in the subsequent processing reactions 15 which occur in the Golgi apparatus of fungi and mammals. In fact, even amongst different lower eukaryotes there exist a great variety of glycosylation structures. This has historically prevented the use of lower eukaryotes as hosts for the production of recombinant human glycoproteins despite otherwise notable advantages over mammalian expression systems.

20 [0017] Therapeutic glycoproteins produced in a microorganism host such as yeast utilizing the endogenous host glycosylation pathway differ structurally from those produced in mammalian cells and typically show greatly reduced therapeutic efficacy. Such glycoproteins are typically immunogenic in humans and show a reduced half-life (and thus bioactivity) *in vivo* after administration (Takeuchi 25 (1997) *Trends in Glycoscience and Glycotechnology* 9:S29-S35). Specific receptors in humans and animals (*i.e.*, macrophage mannose receptors) can recognize terminal mannose residues and promote the rapid clearance of the foreign glycoprotein from the bloodstream. Additional adverse effects may include changes in protein folding, solubility, susceptibility to proteases, 30 trafficking, transport, compartmentalization, secretion, recognition by other proteins or factors, antigenicity, or allergenicity.

[0018] Yeast and filamentous fungi have both been successfully used for the production of recombinant proteins, both intracellular and secreted (Cereghino and Cregg (2000) *FEMS Microbiology Reviews* 24(1):45-66; Harkki *et al.* (1989) *Bio-Technology* 7(6):596; Berka *et al.* (1992) *Abstr. Papers Amer. Chem. Soc.* 203:121-5 BIOT; Svetina *et al.* (2000) *J. Biotechnol.* 76(2-3):245-251). Various yeasts, such as *K. lactis*, *Pichia pastoris*, *Pichia methanolica*, and *Hansenula polymorpha*, have played particularly important roles as eukaryotic expression systems because they are able to grow to high cell densities and secrete large quantities of recombinant protein. Likewise, filamentous fungi, such as *Aspergillus niger*, *Fusarium sp*, *Neurospora crassa* and others, have been used to efficiently produce glycoproteins at the industrial scale. However, as noted above, glycoproteins expressed in any of these eukaryotic microorganisms differ substantially in *N*-glycan structure from those in animals. This has prevented the use of yeast or filamentous fungi as hosts for the production of many therapeutic glycoproteins.

[0019] Although glycosylation in yeast and fungi is very different than in humans, some common elements are shared. The first step, the transfer of the core oligosaccharide structure to the nascent protein, is highly conserved in all eukaryotes including yeast, fungi, plants and humans (compare Figures 1A and 1B). Subsequent processing of the core oligosaccharide, however, differs significantly in yeast and involves the addition of several mannose sugars. This step is catalyzed by mannosyltransferases residing in the Golgi (*e.g.*, *OCH1*, *MNT1*, *MNN1*, etc.), which sequentially add mannose sugars to the core oligosaccharide. The resulting structure is undesirable for the production of humanoid proteins and it is thus desirable to reduce or eliminate mannosyltransferase activity. Mutants of *S. cerevisiae* deficient in mannosyltransferase activity (*e.g.*, *och1* or *mnn9* mutants) have shown to be non-lethal and display a reduced mannose content in the oligosaccharide of yeast glycoproteins. Other oligosaccharide processing enzymes, such as mannosylphosphate transferase, may also have to be eliminated depending on the host's particular endogenous glycosylation pattern. After reducing undesired endogenous glycosylation reactions, the formation of complex *N*-glycans has to be engineered into the host system. This requires the stable expression of several

enzymes and sugar-nucleotide transporters. Moreover, one has to localize these enzymes so that a sequential processing of the maturing glycosylation structure is ensured.

[0020] Several efforts have been made to modify the glycosylation pathways of eukaryotic microorganisms to provide glycoproteins more suitable for use as mammalian therapeutic agents. For example, several glycosyltransferases have been separately cloned and expressed in *S. cerevisiae* (GalT, GnTI), *Aspergillus nidulans* (GnTI) and other fungi (Yoshida *et al.* (1999) *Glycobiology* 9(1):53-8, Kalsner *et al.* (1995) *Glycoconj. J.* 12(3):360-370). However, *N*-glycans resembling those made in human cells were not obtained.

[0021] Yeasts produce a variety of mannosyltransferases (*e.g.*, 1,3-mannosyltransferases such as *MNN1* in *S. cerevisiae*; Graham and Emr (1991) *J. Cell. Biol.* 114(2):207-218), 1,2-mannosyltransferases (*e.g.*, *KTR/KRE* family from *S. cerevisiae*), 1,6-mannosyltransferases (*e.g.*, *OCH1* from *S. cerevisiae*), mannosylphosphate transferases and their regulators (*e.g.*, *MNN4* and *MNN6* from *S. cerevisiae*) and additional enzymes that are involved in endogenous glycosylation reactions. Many of these genes have been deleted individually giving rise to viable organisms having altered glycosylation profiles. Examples are shown in Table 1.

20 **Table 1. Examples of yeast strains having altered mannosylation**

Strain	<i>N</i> -glycan (wild type)	Mutation	<i>N</i> -glycan (mutant)	Reference
<i>S. pombe</i>	Man ₂₋₉ GlcNAc ₂	OCH1	Man ₈ GlcNAc ₂	Yoko-o <i>et al.</i> (2001) <i>FEBS Lett.</i> 489(1):75-80
<i>S. cerevisiae</i>	Man ₂₋₉ GlcNAc ₂	OCH1/MNN1	Man ₈ GlcNAc ₂	Nakanishi-Shindo <i>et al.</i> (1993) <i>J. Biol. Chem.</i> 268(35):26338-26345
<i>S. cerevisiae</i>	Man ₂₋₉ GlcNAc ₂	OCH1/MNN1/MNN4	Man ₈ GlcNAc ₂	Chiba <i>et al.</i> (1998) <i>J. Biol. Chem.</i> 273, 26298-26304
<i>P. pastoris</i>	Hyperglycosylated	OCH1 (complete deletion)	Not hyperglycosylated	Welfide, Japanese Application Publication No. 8-336387
<i>P. pastoris</i>	Man ₂₋₈ GlcNAc ₂	OCH1 (disruption)	Man ₈ GlcNAc ₂	Contreras <i>et al.</i> WO 02/00856 A2

[0022] Japanese Patent Application Publication No. 8-336387 discloses the deletion of an *OCH1* homolog in *Pichia pastoris*. In *S. cerevisiae*, *OCH1* encodes a 1,6-mannosyltransferase, which adds a mannose to the glycan structure $\text{Man}_8\text{GlcNAc}_2$ to yield $\text{Man}_9\text{GlcNAc}_2$. The $\text{Man}_9\text{GlcNAc}_2$ structure, which 5 contains three 1,6 mannose residues, is then a substrate for further 1,2-, 1,6-, and 1,3- mannosyltransferases *in vivo*, leading to the hypermannosylated glycoproteins that are characteristic for *S. cerevisiae* and which typically may have 30-40 mannose residues per *N*-glycan. Because the Och1p initiates the transfer of 1,6 mannose to the $\text{Man}_8\text{GlcNAc}_2$ core, it is often referred to as the “initiating 1,6 10 mannosyltransferase” to distinguish it from other 1,6 mannosyltransferases acting later in the Golgi. In an *och1 mnn1 mnn4* mutant strain of *S. cerevisiae*, proteins glycosylated with $\text{Man}_8\text{GlcNAc}_2$ accumulate and hypermannosylation does not occur. However, $\text{Man}_8\text{GlcNAc}_2$ is not a substrate for mammalian 15 glycosyltransferases, such as human UDP-GlcNAc transferase I, and accordingly, the use of that mutant strain, in itself, is not useful for producing mammalian-like proteins, *i.e.*, those with complex or hybrid glycosylation patterns.

[0023] One can trim $\text{Man}_8\text{GlcNAc}_2$ structures to a $\text{Man}_5\text{GlcNAc}_2$ isomer in *S. cerevisiae* (although high efficiency trimming greater than 50% *in vivo* has yet to be demonstrated) by engineering a fungal mannosidase from *A. saitoi* into the 20 endoplasmic reticulum (ER). The shortcomings of this approach are two-fold: (1) it is not clear whether the $\text{Man}_5\text{GlcNAc}_2$ structures formed are in fact formed *in vivo* (rather than having been secreted and further modified by mannosidases outside the cell); and (2) it is not clear whether any $\text{Man}_5\text{GlcNAc}_2$ structures formed, if in fact formed *in vivo*, are the correct isoform to be a productive 25 substrate for subsequent *N*-glycan modification by GlcNAc transferase I (Maras *et al.* (1997) *Eur. J. Biochem.* 249:701-707).

[0024] With the objective of providing a more human-like glycoprotein derived from a fungal host, U.S. Patent No. 5,834,251 discloses a method for producing a hybrid glycoprotein derived from *Trichoderma reseei*. A hybrid *N*-glycan has only 30 mannose residues on the $\text{Man}\alpha 1\text{-}6$ arm of the core mannose structure and one or two complex antennae on the $\text{Man}\alpha 1\text{-}3$ arm. While this structure has utility, the method has the disadvantage that numerous enzymatic steps must be performed *in*



KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NA, NI, NO, NZ, OM, PG, PH, PL, PT, RO, RU, SC, SD, SE, SG, SK, SL, SY, TJ, TM, TN, TR, TT, TZ, UA, UG, US, UZ, VC, VN, YU, ZA, ZM, ZW.

(84) **Designated States** (*unless otherwise indicated, for every kind of regional protection available*): ARIPO (BW, GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZM, ZW), Eurasian (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European (AT, BE, BG, CH, CY, CZ, DE, DK, EE, ES, FI, FR,

GB, GR, HU, IE, IT, LU, MC, NL, PT, RO, SE, SI, SK, TR), OAPI (BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, ML, MR, NE, SN, TD, TG).

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PRODUCTION OF MODIFIED GLYCOPROTEINS HAVING
MULTIPLE ANTENNAARY STRUCTURES

CROSS-REFERENCE TO RELATED APPLICATIONS

[0001] This application is a continuation-in-part of U. S. Application No. 5 10/680,963, filed on Oct. 7, 2003, which is a continuation-in-part of U. S. Application No. 10/371,877, filed on Feb. 20, 2003, which is a continuation-in-part of U. S. Application No. 09/892,591, filed June 27, 2001, which claims the benefit under 35 U.S.C. § 119(e) of U.S. Provisional Application No. 60/214,358, filed 10 June 28, 2000, U.S. Provisional Application No. 60/215,638, filed June 30, 2000, and U.S. Provisional Application No. 60/279,997, filed March 30, 2001, each of 15 which is incorporated herein by reference in its entirety. This application is also a continuation-in-part of PCT/US02/41510, filed on December 24, 2002, which claims the benefit of U. S. Provisional Application No. 60/344,169, filed on Dec. 27, 2001, each of which is incorporated herein by reference in its entirety.

15

FIELD OF THE INVENTION

[0002] The present invention is directed to methods and compositions by which non-human eukaryotic host cells, such as fungi or other eukaryotic cells, can be genetically modified to produce glycosylated proteins (glycoproteins) having 20 patterns of glycosylation similar to those of glycoproteins produced by animal cells, especially human cells, which are useful as human or animal therapeutic agents.

BACKGROUND OF THE INVENTION

Glycosylation Pathways in Humans and Lower Eukaryotes

[0003] After DNA is transcribed and translated into a protein, further post-translational processing involves the attachment of sugar residues, a process known as glycosylation. Different organisms produce different glycosylation enzymes (glycosyltransferases and glycosidases), and have different substrates (nucleotide sugars) available, so that the glycosylation patterns as well as composition of the individual oligosaccharides, even of the same protein, will be different depending on the host system in which the particular protein is being expressed. Bacteria typically do not glycosylate proteins, and if so only in a very unspecific manner (Moens and Vanderleyden (1997) *Arch Microbiol.* 168(3):169-175). Lower eukaryotes such as filamentous fungi and yeast add primarily mannose and mannosylphosphate sugars. The resulting glycan is known as a "high-mannose" type glycan or a mannan. Plant cells and insect cells (such as SF9 cells) glycosylate proteins in yet another way. By contrast, in higher eukaryotes such as humans, the nascent oligosaccharide side chain may be trimmed to remove several mannose residues and elongated with additional sugar residues that typically are not found in the N-glycans of lower eukaryotes. See, e.g., Brethauer, *et al.* (1999) *Biotechnology and Applied Biochemistry* 30:193-200; Martinet, *et al.* (1998) *Biotechnology Letters* 20:1171-1177; Weikert, *et al.* (1999) *Nature Biotechnology*, 17:1116-1121; M. Malissard, *et al.* (2000) *Biochemical and Biophysical Research Communications* 267:169-173; Jarvis, *et al.*, (1998) *Current Opinion in Biotechnology* 9:528-533; and Takeuchi (1997) *Trends in Glycoscience and Glycotechnology* 9:S29-S35.

[0004] Synthesis of a mammalian-type oligosaccharide structure begins with a set of sequential reactions in the course of which sugar residues are added and removed while the protein moves along the secretory pathway in the host organism. The enzymes which reside along the glycosylation pathway of the host organism or cell determine the resulting glycosylation patterns of secreted proteins. Thus, the resulting glycosylation pattern of proteins expressed in lower eukaryotic host cells differs substantially from the glycosylation pattern of proteins expressed

in higher eukaryotes such as humans and other mammals (Bretthauer, 1999). The structure of a typical fungal *N*-glycan is shown in **Figure 1A**.

[0005] The early steps of human glycosylation can be divided into at least two different phases: (i) lipid-linked Glc₃Man₉GlcNAc₂ oligosaccharides are assembled 5 by a sequential set of reactions at the membrane of the endoplasmic reticulum (ER) (**Figure 13**) and (ii) the transfer of this oligosaccharide from the lipid anchor dolichyl pyrophosphate onto *de novo* synthesized protein. The site of the specific transfer is defined by an asparagine (Asn) residue in the sequence Asn-Xaa-Ser/Thr (SEQ ID NOs:1 and 2) where Xaa can be any amino acid except proline 10 (Gavel and von Heijne (1990) *Protein Eng.* 3:433-42). Further processing by glucosidases and mannosidases occurs in the ER before the nascent glycoprotein is transferred to the early Golgi apparatus, where additional mannose residues are removed by Golgi specific alpha (α)-1,2-mannosidases. Processing continues as the protein proceeds through the Golgi. In the medial Golgi, a number of 15 modifying enzymes, including *N*-acetylglucosaminyl transferases (GnTⅠ, GnTⅡ, GnTⅢ, GnTⅣ and GnTⅤ), mannosidase II and fucosyltransferases, add and remove specific sugar residues. Finally, in the trans-Golgi, galactosyltransferases (GalT) and sialyltransferases (ST) produce a glycoprotein structure that is released from the Golgi. It is this structure, characterized by bi-, tri- and tetra-antennary 20 structures, containing galactose, fucose, N-acetylglucosamine and a high degree of terminal sialic acid, that gives glycoproteins their human characteristics. The structure of a typical human *N*-glycan is shown in **Figure 1B**. See also **Figures 14** and **15** for steps involved in mammalian-type *N*-glycan processing.

[0006] In all eukaryotes studied to date, glycoproteins are derived from a 25 common lipid-linked oligosaccharide precursor Glc₃Man₉GlcNAc₂-dolichol-pyrophosphate. Within the endoplasmic reticulum, synthesis and processing of dolichol pyrophosphate bound oligosaccharides are identical between all known eukaryotes. However, further processing of the core oligosaccharide by fungal cells, *e.g.*, yeast, differs significantly from humans as it moves along the secretory 30 pathway.

[0007] In yeast, these steps are catalyzed by Golgi residing mannosyl-transferases, like Och1p, Mnt1p and Mnn1p, which sequentially add mannose

sugars to the core oligosaccharide. The resulting structure is undesirable for the production of human-like proteins and it is thus desirable to reduce or eliminate mannosyltransferase activity. Mutants of *S. cerevisiae*, deficient in mannosyltransferase activity (for example *och1* or *mnn9* mutants) have been shown to be 5 non-lethal and display reduced mannose content in the oligosaccharide of yeast glycoproteins. Other oligosaccharide processing enzymes, such as mannosylphosphate transferases, may also have to be eliminated depending on the host's particular glycosylation pathways.

10 Sugar Nucleotide Precursors

[0008] The *N*-glycans of animal glycoproteins typically include galactose, fucose, and terminal sialic acid. These sugars are not found on glycoproteins produced in yeast and filamentous fungi. In humans, the full range of nucleotide sugar precursors (e.g., UDP-*N*-acetylglucosamine, UDP-*N*-acetylgalactosamine, 15 CMP-*N*-acetylneuraminic acid, UDP-galactose, GDP-fucose, etc.) are synthesized in the cytosol and transported into the Golgi, where they are attached to the core oligosaccharide by glycosyltransferases. (Sommers and Hirschberg (1981) *J. Cell Biol.* 91(2):A406-A406; Sommers and Hirschberg (1982) *J. Biol. Chem.* 257(18):811-817; Perez and Hirschberg (1987) *Methods in Enzymology* 138:709-20 715).

[0009] Glycosyl transfer reactions typically yield a side product which is a nucleoside diphosphate or monophosphate. While monophosphates can be directly exported in exchange for nucleoside triphosphate sugars by an antiport mechanism, diphosphonucleosides (e.g., GDP) have to be cleaved by phosphatases (e.g. 25 GDPase) to yield nucleoside monophosphates and inorganic phosphate prior to being exported. This reaction is important for efficient glycosylation; for example, GDPase from *Saccharomyces cerevisiae* (*S. cerevisiae*) has been found to be necessary for mannosylation. However that GDPase has 90% reduced activity toward UDP (Berninsone *et al.* (1994) *J. Biol. Chem.* 269(1):207-211). Lower 30 eukaryotes typically lack UDP-specific diphosphatase activity in the Golgi since they do not utilize UDP-sugar precursors for Golgi-based glycoprotein synthesis. *Schizosaccharomyces pombe*, a yeast found to add galactose residues to cell wall

polysaccharides (from UDP-galactose) has been found to have specific UDPase activity, indicating the potential requirement for such an enzyme (Berninsone *et al.* (1994) *J. Biol. Chem.* 269(1):207-211). UDP is known to be a potent inhibitor of glycosyltransferases and the removal of this glycosylation side product may be

5 important to prevent glycosyl-transferase inhibition in the lumen of the Golgi (Khatara *et al.* (1974) *Eur. J. Biochem.* 44:537-560). See Berninsone *et al.* (1995) *J. Biol. Chem.* 270(24):14564-14567; Beaudet *et al.* (1998) *Abc Transporters: Biochemical, Cellular, and Molecular Aspects* 292: 397-413.

10 **Sequential Processing of N-glycans by Compartmentalized Enzyme Activities**

[0010] Sugar transferases and glycosidases (*e.g.*, mannosidases) line the inner (luminal) surface of the ER and Golgi apparatus and thereby provide a “catalytic” surface that allows for the sequential processing of glycoproteins as they proceed through the ER and Golgi network. The multiple compartments of the cis, medial, 15 and trans Golgi and the trans-Golgi Network (TGN), provide the different localities in which the ordered sequence of glycosylation reactions can take place. As a glycoprotein proceeds from synthesis in the ER to full maturation in the late Golgi or TGN, it is sequentially exposed to different glucosidases, mannosidases and glycosyltransferases such that a specific carbohydrate structure may be synthesized. Much work has been dedicated to revealing the exact mechanism by which these enzymes are retained and anchored to their respective organelle. The evolving picture is complex but evidence suggests that stem region, membrane spanning region and cytoplasmic tail, individually or in concert, direct enzymes to the membrane of individual organelles and thereby localize the associated catalytic 20 domain to that locus (*see, e.g.*, Gleeson (1998) *Histochem. Cell Biol.* 109:517-532).

[0011] In some cases, these specific interactions were found to function across species. For example, the membrane spanning domain of α 2,6-ST from rats, an enzyme known to localize in the trans-Golgi of the animal, was shown to also 30 localize a reporter gene (invertase) in the yeast Golgi (Schwientek *et al.* (1995) *J. Biol. Chem.* 270(10):5483-9). However, the very same membrane spanning domain as part of a full-length α 2,6-ST was retained in the ER and not further

transported to the Golgi of yeast (Krezdorn *et al.* (1994) *Eur. J. Biochem.* 220(3):809-17). Full length GalT from humans was not even synthesized in yeast, despite demonstrably high transcription levels. In contrast, the transmembrane region of the same human GalT fused to an invertase reporter was able to direct 5 localization to the yeast Golgi, albeit at low levels. Schwientek and co-workers have shown that fusing 28 amino acids of a yeast mannosyltransferase (*MNT1*), a region containing a cytoplasmic tail, a transmembrane region and eight amino acids of the stem region, to the catalytic domain of human GalT are sufficient for Golgi localization of an active GalT. Other galactosyltransferases appear to rely 10 on interactions with enzymes resident in particular organelles because, after removal of their transmembrane region, they are still able to localize properly. [0012] Improper localization of a glycosylation enzyme may prevent proper functioning of the enzyme in the pathway. For example, *Aspergillus nidulans*, which has numerous α -1,2-mannosidases (Eades and Hintz (2000) *Gene* 15 255(1):25-34), does not add GlcNAc to Man₅GlcNAc₂ when transformed with the rabbit GnTI gene, despite a high overall level of GnTI activity (Kalsner *et al.* (1995) *Glycoconj. J.* 12(3):360-370). GnTI, although actively expressed, may be incorrectly localized such that the enzyme is not in contact with both of its substrates: UDP-GlcNAc and a productive Man₅GlcNAc₂ substrate (not all 20 25 Man₅GlcNAc₂ structures are productive; see below). Alternatively, the host organism may not provide an adequate level of UDP-GlcNAc in the Golgi or the enzyme may be properly localized but nevertheless inactive in its new environment. In addition, Man₅GlcNAc₂ structures present in the host cell may differ in structure from Man₅GlcNAc₂ found in mammals. Maras and coworkers found that about one third of the *N*-glycans from cellobiohydrolase I (CBHI) 30 obtained from *T. reesei* can be trimmed to Man₅GlcNAc₂ by *A. saitoi* 1,2 mannosidase *in vitro*. Fewer than 1% of those *N*-glycans, however, could serve as a productive substrate for GnTI. Maras *et al.* (1997) *Eur. J. Biochem.* 249:701-707. The mere presence of Man₅GlcNAc₂, therefore, does not assure that further *in vivo* processing of Man₅GlcNAc₂ can be achieved. It is formation of a productive, GnTI-reactive Man₅GlcNAc₂ structure that is required. Although Man₅GlcNAc₂ could be produced in the cell (about 27 mol %), only a small

fraction could be converted to $\text{Man}_5\text{GlcNAc}_2$ (less than about 5%, *see Chiba et al. WO 01/14522*).

[0013] To date, there is no reliable way of predicting whether a particular heterologously expressed glycosyltransferase or mannosidase in a lower eukaryote will be (1), sufficiently translated (2), catalytically active or (3) located to the proper organelle within the secretory pathway. Because all three of these are necessary to affect glycosylation patterns in lower eukaryotes, a systematic scheme to achieve the desired catalytic function and proper retention of enzymes in the absence of predictive tools, which are currently not available, would be desirable.

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Production of Therapeutic Glycoproteins

[0014] A significant number of proteins isolated from humans or animals are post-translationally modified, with glycosylation being one of the most significant modifications. An estimated 70% of all therapeutic proteins are glycosylated and thus currently rely on a production system (*i.e.*, host cell) that is able to glycosylate in a manner similar to humans. Several studies have shown that glycosylation plays an important role in determining the (1) immunogenicity, (2) pharmacokinetic properties, (3) trafficking, and (4) efficacy of therapeutic proteins. It is thus not surprising that substantial efforts by the pharmaceutical industry have been directed at developing processes to obtain glycoproteins that are as "humanoid" or "human-like" as possible. To date, most glycoproteins are made in a mammalian host system. This may involve the genetic engineering of such mammalian cells to enhance the degree of sialylation (*i.e.*, terminal addition of sialic acid) of proteins expressed by the cells, which is known to improve pharmacokinetic properties of such proteins. Alternatively, one may improve the degree of sialylation by *in vitro* addition of such sugars using known glycosyltransferases and their respective nucleotide sugars (*e.g.*, 2,3-sialyltransferase and CMP-sialic acid).

[0015] While most higher eukaryotes carry out glycosylation reactions that are similar to those found in humans, recombinant human proteins expressed in the above mentioned host systems invariably differ from their "natural" human counterpart (Raju *et al.* (2000) *Glycobiology* 10(5): 477-486). Extensive

development work has thus been directed at finding ways to improve the "human character" of proteins made in these expression systems. This includes the optimization of fermentation conditions and the genetic modification of protein expression hosts by introducing genes encoding enzymes involved in the formation 5 of human-like glycoforms. Goochee *et al.* (1999) *Biotechnology* 9(12):1347-55; Andersen and Goochee (1994) *Curr Opin Biotechnol.* 5(5):546-49; Werner *et al.* (1998) *Arzneimittelforschung*. 48(8):870-80; Weikert *et al.* (1999) *Nat Biotechnol.* 17(11):1116-21; Yang and Butler (2000) *Biotech. Bioeng.* 68:370-80. Inherent problems associated with all mammalian expression systems have not been solved.

10

Glycoprotein Production Using Eukaryotic Microorganisms

[0016] Although the core oligosaccharide structure transferred to a protein in the endoplasmic reticulum is basically identical in mammals and lower eukaryotes, substantial differences have been found in the subsequent processing reactions 15 which occur in the Golgi apparatus of fungi and mammals. In fact, even amongst different lower eukaryotes there exist a great variety of glycosylation structures. This has historically prevented the use of lower eukaryotes as hosts for the production of recombinant human glycoproteins despite otherwise notable advantages over mammalian expression systems.

20 [0017] Therapeutic glycoproteins produced in a microorganism host such as yeast utilizing the endogenous host glycosylation pathway differ structurally from those produced in mammalian cells and typically show greatly reduced therapeutic efficacy. Such glycoproteins are typically immunogenic in humans and show a reduced half-life (and thus bioactivity) *in vivo* after administration (Takeuchi 25 (1997) *Trends in Glycoscience and Glycotechnology* 9:S29-S35). Specific receptors in humans and animals (*i.e.*, macrophage mannose receptors) can recognize terminal mannose residues and promote the rapid clearance of the foreign glycoprotein from the bloodstream. Additional adverse effects may include changes in protein folding, solubility, susceptibility to proteases, 30 trafficking, transport, compartmentalization, secretion, recognition by other proteins or factors, antigenicity, or allergenicity.

[0018] Yeast and filamentous fungi have both been successfully used for the production of recombinant proteins, both intracellular and secreted (Cereghino and Cregg (2000) *FEMS Microbiology Reviews* 24(1):45-66; Harkki *et al.* (1989) *Bio-Technology* 7(6):596; Berka *et al.* (1992) *Abstr. Papers Amer. Chem. Soc.* 203:121-5 BIOT; Svetina *et al.* (2000) *J. Biotechnol.* 76(2-3):245-251). Various yeasts, such as *K. lactis*, *Pichia pastoris*, *Pichia methanolica*, and *Hansenula polymorpha*, have played particularly important roles as eukaryotic expression systems because they are able to grow to high cell densities and secrete large quantities of recombinant protein. Likewise, filamentous fungi, such as *Aspergillus niger*, *Fusarium sp*,

10 *Neurospora crassa* and others, have been used to efficiently produce glycoproteins at the industrial scale. However, as noted above, glycoproteins expressed in any of these eukaryotic microorganisms differ substantially in *N*-glycan structure from those in animals. This has prevented the use of yeast or filamentous fungi as hosts for the production of many therapeutic glycoproteins.

15 [0019] Although glycosylation in yeast and fungi is very different than in humans, some common elements are shared. The first step, the transfer of the core oligosaccharide structure to the nascent protein, is highly conserved in all eukaryotes including yeast, fungi, plants and humans (compare Figures 1A and 1B). Subsequent processing of the core oligosaccharide, however, differs

20 significantly in yeast and involves the addition of several mannose sugars. This step is catalyzed by mannosyltransferases residing in the Golgi (*e.g.*, *OCH1*, *MNT1*, *MNN1*, etc.), which sequentially add mannose sugars to the core oligosaccharide. The resulting structure is undesirable for the production of humanoid proteins and it is thus desirable to reduce or eliminate

25 mannosyltransferase activity. Mutants of *S. cerevisiae* deficient in mannosyltransferase activity (*e.g.*, *och1* or *mnn9* mutants) have shown to be non-lethal and display a reduced mannose content in the oligosaccharide of yeast glycoproteins. Other oligosaccharide processing enzymes, such as mannosylphosphate transferase, may also have to be eliminated depending on the

30 host's particular endogenous glycosylation pattern. After reducing undesired endogenous glycosylation reactions, the formation of complex *N*-glycans has to be engineered into the host system. This requires the stable expression of several

enzymes and sugar-nucleotide transporters. Moreover, one has to localize these enzymes so that a sequential processing of the maturing glycosylation structure is ensured.

[0020] Several efforts have been made to modify the glycosylation pathways of 5 eukaryotic microorganisms to provide glycoproteins more suitable for use as mammalian therapeutic agents. For example, several glycosyltransferases have been separately cloned and expressed in *S. cerevisiae* (GalT, GnTI), *Aspergillus nidulans* (GnTI) and other fungi (Yoshida *et al.* (1999) *Glycobiology* 9(1):53-8, Kalsner *et al.* (1995) *Glycoconj. J.* 12(3):360-370). However, *N*-glycans 10 resembling those made in human cells were not obtained.

[0021] Yeasts produce a variety of mannosyltransferases (*e.g.*, 1,3-mannosyltransferases such as *MNN1* in *S. cerevisiae*; Graham and Emr (1991) *J. Cell. Biol.* 114(2):207-218), 1,2-mannosyltransferases (*e.g.*, *KTR/KRE* family from *S. cerevisiae*), 1,6-mannosyltransferases (*e.g.*, *OCH1* from *S. cerevisiae*), 15 mannosylphosphate transferases and their regulators (*e.g.*, *MNN4* and *MNN6* from *S. cerevisiae*) and additional enzymes that are involved in endogenous glycosylation reactions. Many of these genes have been deleted individually giving rise to viable organisms having altered glycosylation profiles. Examples are shown in Table 1.

20 **Table 1. Examples of yeast strains having altered mannosylation**

Strain	<i>N</i> -glycan (wild type)	Mutation	<i>N</i> -glycan (mutant)	Reference
<i>S. pombe</i>	Man ₉ GlcNAc ₂	OCH1	Man ₈ GlcNAc ₂	Yoko-o <i>et al.</i> (2001) <i>FEBS Lett.</i> 489(1):75-80
<i>S. cerevisiae</i>	Man ₉ GlcNAc ₂	OCH1/MNN1	Man ₈ GlcNAc ₂	Nakanishi-Shindo <i>et al.</i> (1993) <i>J. Biol. Chem.</i> 268(35):26338-26345
<i>S. cerevisiae</i>	Man ₉ GlcNAc ₂	OCH1/MNN1/MNN4	Man ₈ GlcNAc ₂	Chiba <i>et al.</i> (1998) <i>J. Biol. Chem.</i> 273, 26298-26304
<i>P. pastoris</i>	Hyperglycosylated	OCH1 (complete deletion)	Not hyperglycosylated	Welfide, Japanese Application Publication No. 8-336387
<i>P. pastoris</i>	Man ₈ GlcNAc ₂	OCH1 (disruption)	Man ₈ GlcNAc ₂	Contreras <i>et al.</i> WO 02/00856 A2

[0022] Japanese Patent Application Publication No. 8-336387 discloses the deletion of an *OCH1* homolog in *Pichia pastoris*. In *S. cerevisiae*, *OCH1* encodes a 1,6-mannosyltransferase, which adds a mannose to the glycan structure $\text{Man}_8\text{GlcNAc}_2$ to yield $\text{Man}_9\text{GlcNAc}_2$. The $\text{Man}_9\text{GlcNAc}_2$ structure, which contains three 1,6 mannose residues, is then a substrate for further 1,2-, 1,6-, and 1,3- mannosyltransferases *in vivo*, leading to the hypermannosylated glycoproteins that are characteristic for *S. cerevisiae* and which typically may have 30-40 mannose residues per *N*-glycan. Because the Och1p initiates the transfer of 1,6 mannose to the $\text{Man}_8\text{GlcNAc}_2$ core, it is often referred to as the “initiating 1,6 mannosyltransferase” to distinguish it from other 1,6 mannosyltransferases acting later in the Golgi. In an *och1 mnn1 mnn4* mutant strain of *S. cerevisiae*, proteins glycosylated with $\text{Man}_8\text{GlcNAc}_2$ accumulate and hypermannosylation does not occur. However, $\text{Man}_8\text{GlcNAc}_2$ is not a substrate for mammalian glycosyltransferases, such as human UDP-GlcNAc transferase I, and accordingly, the use of that mutant strain, in itself, is not useful for producing mammalian-like proteins, *i.e.*, those with complex or hybrid glycosylation patterns.

[0023] One can trim $\text{Man}_8\text{GlcNAc}_2$ structures to a $\text{Man}_5\text{GlcNAc}_2$ isomer in *S. cerevisiae* (although high efficiency trimming greater than 50% *in vivo* has yet to be demonstrated) by engineering a fungal mannosidase from *A. saitoi* into the endoplasmic reticulum (ER). The shortcomings of this approach are two-fold: (1) it is not clear whether the $\text{Man}_5\text{GlcNAc}_2$ structures formed are in fact formed *in vivo* (rather than having been secreted and further modified by mannosidases outside the cell); and (2) it is not clear whether any $\text{Man}_5\text{GlcNAc}_2$ structures formed, if in fact formed *in vivo*, are the correct isoform to be a productive substrate for subsequent *N*-glycan modification by GlcNAc transferase I (Maras *et al.* (1997) *Eur. J. Biochem.* 249:701-707).

[0024] With the objective of providing a more human-like glycoprotein derived from a fungal host, U.S. Patent No. 5,834,251 discloses a method for producing a hybrid glycoprotein derived from *Trichoderma reseei*. A hybrid *N*-glycan has only mannose residues on the $\text{Man}\alpha 1\text{-}6$ arm of the core mannose structure and one or two complex antennae on the $\text{Man}\alpha 1\text{-}3$ arm. While this structure has utility, the method has the disadvantage that numerous enzymatic steps must be performed *in*

vitro, which is costly and time-consuming. Isolated enzymes are expensive to prepare and need costly substrates (e.g., UDP-GlcNAc). The method also does not allow for the production of complex glycans on a desired protein.

5 **Intracellular Mannosidase Activity Involved in N-glycan Trimming**

[0025] Alpha-1,2-mannosidase activity is required for the trimming of Man₈GlcNAc₂ to form Man₅GlcNAc₂, which is a major intermediate for complex *N*-glycan formation in mammals. Previous work has shown that truncated murine, fungal and human α -1,2-mannosidase can be expressed in the methylotropic yeast 10 *P. pastoris* and display Man₈GlcNAc₂ to Man₅GlcNAc₂ trimming activity (Lal *et al.* (1998) *Glycobiology* 8(10):981-95; Tremblay *et al.* (1998) *Glycobiology* 8(6):585-95, Callewaert *et al.* (2001) *FEBS Lett.* 503(2-3):173-8). However, to date, no reports exist that show the high level *in vivo* trimming of Man₈GlcNAc₂ to Man₅GlcNAc₂ on a secreted glycoprotein from *P. pastoris*.

15 [0026] Moreover, the mere presence of an α -1,2-mannosidase in the cell does not, by itself, ensure proper intracellular trimming of Man₈GlcNAc₂ to Man₅GlcNAc₂. (*See, e.g.*, Contreras *et al.* WO 02/00856 A2, in which an HDEL tagged mannosidase of *T. reesei* is localized primarily in the ER and co-expressed with an influenza haemagglutinin (HA) reporter protein on which virtually no 20 Man₅GlcNAc₂ could be detected. *See also* Chiba *et al.* (1998) *J. Biol. Chem.* 273(41): 26298-26304, in which a chimeric α -1,2-mannosidase/Och1p transmembrane domain fusion localized in the ER, early Golgi and cytosol of *S. cerevisiae*, had no mannosidase trimming activity). Accordingly, mere localization of a mannosidase in the ER or Golgi is insufficient to ensure activity of the 25 respective enzyme in that targeted organelle. (*See also*, Martinet *et al.* (1998) *Biotech. Letters* 20(12): 1171-1177, showing that α -1,2-mannosidase from *T. reesei*, while localizing intracellularly, increased rather than decreased the extent of mannosylation). To date, there is no report that demonstrates the intracellular localization of an active heterologous α -1,2- mannosidase in either yeast or fungi 30 using a transmembrane localization sequence.

[0027] While it is useful to engineer strains that are able to produce Man₅GlcNAc₂ as the primary *N*-glycan structure, any attempt to further modify

these high mannose precursor structures to more closely resemble human glycans requires additional *in vivo* or *in vitro* steps. Methods to further humanize glycans from fungal and yeast sources *in vitro* are described in U.S. Pat. No. 5,834,251 (*supra*). If $\text{Man}_5\text{GlcNAc}_2$ is to be further humanized *in vivo*, one has to ensure that

5 the generated $\text{Man}_5\text{GlcNAc}_2$ structures are, in fact, generated intracellularly and not the product of mannosidase activity in the medium. Complex *N*-glycan formation in yeast or fungi will require high levels of $\text{Man}_5\text{GlcNAc}_2$ to be generated within the cell because only intracellular $\text{Man}_5\text{GlcNAc}_2$ glycans can be further processed to hybrid and complex *N*-glycans *in vivo*. In addition, one has to

10 demonstrate that the majority of $\text{Man}_5\text{GlcNAc}_2$ structures generated are in fact a substrate for GnTI and thus allow the formation of hybrid and complex *N*-glycans.

[0028] Accordingly, the need exists for methods to produce glycoproteins characterized by a high intracellular $\text{Man}_5\text{GlcNAc}_2$ content which can be further processed into human-like glycoprotein structures in non-human eukaryotic host

15 cells, and particularly in yeast and filamentous fungi.

N-Acetylglucosaminyltransferases

[0029] *N*-Acetylglucosaminyltransferases (“GnTs”) belong to another class of glycosylation enzymes that modify *N*-linked oligosaccharides in the secretory pathway. Such glycosyltransferases catalyze the transfer of a monosaccharide from specific sugar nucleotide donors onto particular hydroxyl position of a monosaccharide in a growing glycan chain in one of two possible anomeric linkages (either α or β). Dennis *et al.* (1999) *Bioessays* 21(5):412-21. Specific GnTs add *N*-acetylglucosamine (“GlcNAc”) onto the $\text{Man}\alpha 1,6$ arm or the $\text{Man}\alpha 1,3$ arm of an *N*-glycan substrate (*e.g.*, $\text{Man}_5\text{GlcNAc}_2$ (“mannose-5 core”) and $\text{Man}_3\text{GlcNAc}_2$ (an “inner core structure”)). The reaction product (*e.g.*, GlcNAc $\text{Man}_5\text{GlcNAc}_2$ or GlcNAc $_2\text{Man}_3\text{GlcNAc}_2$) can then be modified into bi-, tri-, tetra- and penta-antennary *N*-linked oligosaccharide structures.

[0030] *N*-Acetylglucosaminyltransferase III (“GnTIII”) is an enzyme that catalyzes the addition of a GlcNAc, on the middle mannose of the trimannose core ($\text{Man}\alpha 1,6(\text{Man}\alpha 1,3)\text{Man}\beta 1,4 - \text{GlcNAc}\beta 1,4 - \text{GlcNAc}\beta 1,4 - \text{Asn}$) of an *N*-linked oligosaccharide. The addition by GnTIII of a bisecting GlcNAc to an acceptor

substrate (*e.g.* trimannose core) yields a so-called bisected *N*-glycan. For example, the addition by GnTIII of a bisecting GlcNAc to the GlcNAcMan₃GlcNAc₂ structure may yield a bisected *N*-glycan, GlcNAc₂Man₃GlcNAc₂. Similarly, the addition by GnTIII of a bisecting GlcNAc to a GlcNAc₂Man₃GlcNAc₂ structure 5 yields another bisected *N*-glycan, GlcNAc₃Man₃GlcNAc₂. This latter structure has been implicated in greater antibody-dependent cellular cytotoxicity (ADCC).

Umana *et al.* (1999) *Nat. Biotechnol.* 17(2):176-80. Other bisected *N*-glycans can be formed by the action of GnTIII. For example, GlcNAcMan₄GlcNAc₂ can be converted to bisected GlcNAc₂Man₄GlcNAc₂, Man₅GlcNAc₂ can be converted to 10 bisected GlcNAcMan₅GlcNAc₂, and GlcNAcMan₅GlcNAc₂ can be converted to bisected GlcNAc₂Man₅GlcNAc₂. *See, e.g.*, Narasimhan (1982) *J. Biol. Chem.* 257:10235-42. Thus far, GnTIII activity has only been shown in mammalian cells.

[0031] Re-engineering glycoforms of immunoglobulins expressed by mammalian cells is a tedious and cumbersome task. Especially in the case of 15 GnTIII, where over-expression of this enzyme has been implicated in growth inhibition, methods involving regulated (inducible) gene expression had to be employed to produce immunoglobulins with bisected *N*-glycans. Umana *et al.* (1999) *Biotechnol Bioeng.* 65(5):542-9; Umana *et al.* (1999) *Nat. Biotechnol.* 17(2):176-80; Umana *et al.* WO 03/011878; U.S. Patent No. 6,602,684. Such a 20 growth-inhibition effect complicates the ability to coexpress the target protein and GnTIII and may impose an upper limit on GnTIII overexpression. U.S. Patent No. 6,602,684. Careful optimization of the expression levels of GnTIII may be necessary. *Id.* As described above, however, development of the lower eukaryotic host cells used in such a protein production system requires that the endogenous 25 glycosylation pathways of the host cells be further modified.

[0032] The enzymes GnTIV, GnTV and GnTIX expressed in mammalian cells are known to catalyze the transfer of GlcNAc residues in particular conformation onto oligosaccharide substrates producing multiantennary glycan structures. UDP-*N*-acetylglucosamine: α 1,3-D-mannoside β 1,4-*N*-acetylglucosaminyl-transferase 30 (GnTIV; EC 2.4.1.145) catalyzes the transfer of GlcNAc from UDP-GlcNAc in residues in β 1,4 linkage to α 1,3-D-mannoside on GlcNAc β 1-2Man α 1-6(GlcNAc β 1-2Man α 1-3)Man β 1-4GlcNAc β 1-4GlcNAc β 1-Asn (Gleeson and Schachter, *J Biol*

Chem. 1983 May 25;258(10):6162-73; Schachter et al., (1989) *Methods Enzymol.*, 179, 351-397). UDP-*N*-acetylglucosamine: α -6-D-mannoside β 1,6-*N*-acetylglucosaminyltransferase (GnTV; EC 4.1.155) catalyzes the addition of an *N*-acetylglucosamine to the α 1,6 mannosyl core in a β 1,6 linkage forming tri- and 5 tetraantennary *N*-glycans.

[0033] Similarly, the expression of GnTVI in avian cells catalyzes the transfer of GlcNAc residues onto oligosaccharide substrates. Specifically, UDP *N*-acetyl-D-glucosamine (GlcNAc):GlcNAc β 1-6(GlcNAc β 1-2)Man α 1-R[GlcNAc to Man] β 1,4-*N*-acetylglucosaminyltransferase VI(GnTVI) catalyzes the formation of 10 pentaantennary *N*-glycans (Sakamoto et al., *J. Biol. Chem.* 2000 Nov 17;275(46):36029-34). The gene encoding GnTVI has been purified and isolated recently. Taniguchi et al., JP2002209587A2.

[0034] Substrates required to produce complex multiantennary structures have not been synthesized in fungal hosts until recently (Hamilton et al., *Science*. 2003 15 Aug 29;301 (5637): 1244-6). Mammalian cells typically produce an array of complex glycans such as biantennary, triantennary, tetraantennary and even pentaantennary glycoforms through sequential reaction of specific GnTs. In the Golgi apparatus of such cells, *N*-glycan processing of glycoproteins produces biantennary structures predominantly, in addition to the formation of triantennary 20 and tetraantennary structures. It is currently understood that in the formation of complex glycans specific GnTs catalyze specific β -GlcNAc linkages (e.g., β 1,2; β 1,4; β 1,6), producing multiantennary glycans in mammalian cells. These cells, however, are incapable of producing any one homogeneous glycoform in high yield.

[0035] Recently, lower eukaryotes have been engineered to produce complex 25 glycans in homogeneous forms at significant levels (*Science* 2003 Aug 29;301(5637):1244-6). The ability to produce multiantennary complex glycans in lower eukaryotes would provide large amounts of properly folded and glycosylated proteins on an industrial scale at low cost, in faster time, safer and in higher 30 quality. What is needed, therefore, is a protein production system utilizing the inherent capability of robust product titers such as those produced in lower eukaryotic host cells (e.g. yeast and filamentous fungi), which is capable of

producing multiantennary (and optionally, bisected) *N*-linked glycans on proteins, especially therapeutic proteins, expressed in these cells.

SUMMARY OF THE INVENTION

5 [0036] Host cells and cell lines having genetically modified glycosylation pathways that allow them to carry out a sequence of enzymatic reactions which mimic the processing of glycoproteins in mammals, especially in humans, have been developed. Recombinant proteins expressed in these engineered hosts yield glycoproteins more similar, if not substantially identical, to their mammalian, *e.g.*,
10 human counterparts. Host cells of the invention, *e.g.*, lower eukaryotic microorganisms and other non-human, eukaryotic host cells grown in culture, are modified to produce *N*-glycans, such as bisected *N*-glycans, or other structures produced along human glycosylation pathways. This result is achieved using a combination of engineering and/or selection of strains that do not, for example,
15 express enzymes that create the undesirable structures characteristic of the fungal glycoproteins and that do, for example, express heterologous enzymes capable of producing a "human-like" glycoprotein.

[0037] The present invention thus provides a glycoprotein production method using (1) a lower eukaryotic host such as a unicellular or filamentous fungus, or (2)
20 any non-human eukaryotic organism that has a different glycosylation pattern from humans, to modify the glycosylation composition and structures of the proteins made in a host organism ("host cell") so that they resemble more closely carbohydrate structures found in mammalian, *e.g.*, human proteins. The process allows one to obtain an engineered host cell which can be used to express and
25 target any desirable gene(s), *e.g.*, one involved in glycosylation, by methods that are well-established in the scientific literature and generally known to the artisan in the field of protein expression. Host cells with modified oligosaccharides are created or selected. For the production of therapeutic proteins, this method may be adapted to engineer cell lines in which any desired glycosylation structure may be obtained.
30

[0038] Accordingly, in one embodiment, the invention provides methods for making a human-like glycoprotein in a lower eukaryotic host cell by introduction

into the cell of an *N*-acetylglucosaminyltransferase III activity. In a preferred embodiment, the *N*-acetylglucosaminyltransferase III activity is expressed in the cell, and in an even more preferred embodiment, this expression results in the production of *N*-glycans comprising GlcNAc₃Man₃GlcNAc₂,

5 GlcNAc₂Man₃GlcNAc₂, or GlcNAc₂Man₅GlcNAc₂ bisected structures. In another preferred embodiment, the *N*-acetylglucosaminyltransferase III activity is substantially intracellular. In another preferred embodiment of the invention, the glycoprotein including the *N*-glycans with bisected structures is isolated from the lower eukaryotic host cell. In an even more preferred embodiment, the 10 glycoprotein produced in the host cell is a therapeutic protein.

[0039] In another aspect, the invention provides a lower eukaryotic host cell that includes both an *N*-acetylglucosaminyltransferase III activity and an *N*-acetylglucosaminyltransferase II activity. In a preferred embodiment, the host cell including the *N*-acetylglucosaminyltransferase III activity produces *N*-glycans 15 comprising GlcNAcMan₃GlcNAc₂ structures that are capable of reacting with this activity. In a more preferred embodiment, the activity produces a bisected glycan. The lower eukaryotic host cell of some embodiments of the invention may thus include an *N*-glycan with a bisected glycan. In a preferred embodiment, the *N*-glycan includes greater than 10 mole % of the bisected glycan. In some 20 embodiments, the host cell includes an *N*-glycan that comprises GlcNAc₃Man₃GlcNAc₂, GlcNAc₂Man₃GlcNAc₂, or GlcNAc₂Man₅GlcNAc₂ bisected structures. In a preferred embodiment, the host cell includes a Man₅GlcNAc₂ core structure or a Man₃GlcNAc₂ core structure that is modified by a bisecting GlcNAc. In an even more preferred embodiment, the cell produces 25 greater than 10 mole % of the modified structure.

[0040] In another embodiment of the invention, the lower eukaryotic host cell contains an *N*-acetylglucosaminyltransferase I activity in addition to the *N*-acetylglucosaminyltransferase III activity. In a preferred embodiment, the activities are substantially intracellular. In another preferred embodiment, the cell 30 produces *N*-glycans comprising GlcNAcMan₃GlcNAc₂ that are capable of reacting with the GnTIII activity. In an even more preferred embodiment, the GnTIII activity of the cell produces a bisected glycan.

[0041] In another embodiment, the lower eukaryotic host cell of the invention contains both an *N*-acetylglucosaminyltransferase III activity and a mannosidase II activity. In a preferred embodiment, the host cell further contains an *N*-acetylglucosaminyltransferase I activity. In another preferred embodiment, the host cell further contains an *N*-acetylglucosaminyltransferase II activity. In another preferred embodiment, the host cell further contains both an *N*-acetylglucosaminyltransferase I activity and an *N*-acetylglucosaminyltransferase II activity.

[0042] The present invention also provides methods for making a human-like glycoprotein in a lower eukaryotic host cell by introduction into the cell of an *N*-acetylglucosaminyltransferase IV activity. In a preferred embodiment, the *N*-acetylglucosaminyltransferase IV activity is expressed in the cell, and in an even more preferred embodiment, this expression results in the production of *N*-glycans comprising GlcNAc₃Man₃GlcNAc₂ structure. In another preferred embodiment, the *N*-acetylglucosaminyltransferase IV activity is substantially intracellular. In another preferred embodiment of the invention, the glycoprotein including the *N*-glycans with triantennary structures is isolated from the lower eukaryotic host cell. In a more preferred embodiment, the *N*-glycan includes greater than 90 mole % of the triantennary glycan. In an even more preferred embodiment, the glycoprotein produced in the host cell is a therapeutic protein.

[0043] Thus, in another aspect, the invention provides a lower eukaryotic host cell that includes both an *N*-acetylglucosaminyltransferase IV activity and an *N*-acetylglucosaminyltransferase V activity. In a preferred embodiment, the host cell including the *N*-acetylglucosaminyltransferase IV and V activities produces *N*-glycans comprising GlcNAc₄Man₃GlcNAc₂ structures. In a more preferred embodiment, the activity produces a tetraantennary glycan. The lower eukaryotic host cell of some embodiments of the invention may thus include an *N*-glycan with a tetraantennary glycan. In some embodiments, the host cell includes an *N*-glycan that comprises GlcNAc₃Man₃GlcNAc₂ and GlcNAc₄Man₃GlcNAc₂ structures. In a preferred embodiment, the host cell includes a GlcNAcMan₅GlcNAc₂ core structure that is modified by a GnT IV or a GlcNAc₂Man₃GlcNAc₂ core structure that is modified by a GnT IV and GnT V. In a preferred embodiment, the *N*-

glycan includes greater than 70 mole % of the tetraantennary glycan. In an even more preferred embodiment, the cell produces greater than 75 mole % of tetraantennary glycans.

[0044] The present invention also provides a lower eukaryotic host cell that 5 includes an *N*-acetylglucosaminyltransferase VI activity. In a preferred embodiment, the host cell expressing the *N*-acetylglucosaminyltransferase VI activity produces *N*-glycans comprising GlcNAc₅Man₃GlcNAc₂ structures (e.g., pentaantennary glycans). The lower eukaryotic host cell of some embodiments of the invention may thus include an *N*-glycan with a pentaantennary glycan. In some 10 embodiments, the host cell includes an *N*-glycan that comprises GlcNAc₅Man₃GlcNAc₂ structures. In a preferred embodiment, the host cell includes a GlcNAc₂Man₃GlcNAc₂ core structure that is modified by a GnT VI.

[0045] In another embodiment, the invention provides methods for making a human-like glycoprotein in a lower eukaryotic host cell by introduction into the 15 cell of an *N*-acetylglucosaminyltransferase IX activity. In a preferred embodiment, the *N*-acetylglucosaminyltransferase IX activity is expressed in the cell, and in an even more preferred embodiment, this expression results in the production of *N*-glycans comprising GlcNAc₃Man₃GlcNAc₂ and GlcNAc₄Man₃GlcNAc₂ structures. In another preferred embodiment, the *N*-acetylglucosaminyltransferase IX activity 20 is substantially intracellular. In another preferred embodiment of the invention, the glycoprotein including the *N*-glycans with multiantennary structures is isolated from the lower eukaryotic host cell. In an even more preferred embodiment, the glycoprotein produced in the host cell is a therapeutic protein.

[0046] In another embodiment of the invention, the lower eukaryotic host cell 25 contains an *N*-acetylglucosaminyltransferase I activity and an *N*-acetylglucosaminyltransferase II activity. In a preferred embodiment, the activities are substantially intracellular. In another preferred embodiment, the cell produces *N*-glycans comprising GlcNAc₂Man₃GlcNAc₂ that are capable of reacting with the GnTIV activity producing triantennary glycans. In an even more preferred 30 embodiment, the GnTV activity of the cell produces a tetraantennary glycans.

[0047] In another embodiment, the lower eukaryotic host cell of the invention contains both an *N*-acetylglucosaminyltransferase IV activity and a mannosidase II

activity. In a preferred embodiment, the host cell further contains an *N*-acetylglucosaminyltransferase I activity. In another preferred embodiment, the host cell further contains an *N*-acetylglucosaminyltransferase II activity. In another preferred embodiment, the host cell further contains an *N*-acetylglucosaminyltransferase V activity.

5 [0048] In certain preferred embodiments, the host cell of the invention is deficient in an *OCH1* mannosyltransferase activity. Such a cell may, for example, be deficient in a Dol-P-Man:Man₅GlcNAc₂-PP-Dol mannosyltransferase activity. In yet another embodiment, the host cell of the invention may further comprise an 10 α-1,2-mannosidase I activity. In another embodiment, the host cell may further comprise a sugar nucleotide transporter. Preferably, the host cell comprises a UDP-GlcNAc transporter wherein the transfer of GlcNAc residues is facilitated by any one of the above mentioned *N*-acetylglucosaminyltransferase activities.

15 [0049] The present invention also provides glycoproteins that are made by the processes of the invention. In one embodiment, the glycoprotein includes a bisecting GlcNAc on a Man₅GlcNAc₂ or a Man₃GlcNAc₂ core structure and is produced in a lower eukaryotic host cell. In another embodiment, the glycoprotein includes a bisecting GlcNAc attached to a Man₅GlcNAc₂, Man₄GlcNAc₂, Man₃GlcNAc₂, GlcNAcMan₃GlcNAc₂, GlcNAcMan₂GlcNAc₂, or a 20 GlcNAc₂Man₃GlcNAc₂ core structure and is produced in a lower eukaryotic host cell. In a preferred embodiment, greater than 10 mole % of the core structures of the glycoprotein of the invention are modified by the bisecting GlcNAc.

25 [0050] In yet another embodiment, the invention provides a glycoprotein that includes a triantennary structure such as GlcNAc₃Man₃GlcNAc₂ and is produced in a lower eukaryotic host cell. In a preferred embodiment, greater than 90 mole % of the core structures of the glycoprotein of the invention are modified by the GnTIV. In another embodiment, the glycoprotein includes a tetraantennary structure such as GlcNAc₄Man₃GlcNAc₂ and is produced in a lower eukaryotic host cell. In a more preferred embodiment, greater than 75 mole % of the core structures of the 30 glycoprotein of the invention are modified by the GnTV.

[0051] In another aspect, the invention provides pharmaceutical compositions that contain the human-like glycoproteins produced in a lower eukaryotic host cell.

Also provided according to the invention are vectors encoding proteins having one or more *N*-acetylglucosaminyltransferase III, IV, V, VI and IX activities and containing attached targeting peptide sequences. In a preferred embodiment, the proteins encoded by the vectors are localized in a lower eukaryotic host cell such 5 that they produce *N*-glycans having bisected and/or multiantennary structures.

BRIEF DESCRIPTION OF THE DRAWINGS

[0052] **Figure 1A** is a schematic diagram of a typical fungal *N*-glycosylation pathway.

10 [0053] **Figure 1B** is a schematic diagram of a typical human *N*-glycosylation pathway.

[0054] **Figure 2** depicts construction of a combinatorial DNA library of fusion constructs. **Figure 2A** diagrams the insertion of a targeting peptide fragment into pCR2.1-TOPO (Invitrogen, Carlsbad, CA). **Figure 2B** shows the generated 15 targeting peptide sub-library having restriction sites *NotI* - *AscI*. **Figure 2C** diagrams the insertion of a catalytic domain region into pJN347, a modified pUC19 vector. **Figure 2D** shows the generated catalytic domain sub-library having restriction sites *NotI*, *AscI* and *PacI*. **Figure 2E** depicts one particular fusion construct generated from the targeting peptide sub-library and the catalytic 20 domain sub-library.

[0055] **Figure 3** illustrates the *M. musculus* α -1,2-mannosidase IA open reading frame nucleic acid sequence (SEQ ID NO:48) and encoded polypeptide sequence (SEQ ID NO:49). The sequences of the PCR primers used to generate N-terminal truncations are underlined.

25 [0056] **Figure 4** illustrates engineering of vectors with multiple auxotrophic markers and genetic integration of target proteins in the *P. pastoris* *OCH1* locus.

[0057] **Figures 5A – 5E** show MALDI-TOF analysis demonstrating production of kringle 3 domain of human plasminogen (K3) glycoproteins having Man₅GlcNAc₂ as the predominant *N*-glycan structure in *P. pastoris*. **Figure 5A** 30 depicts the standard Man₅GlcNAc₂ [a] glycan (Glyko, Novato, CA) and Man₅GlcNAc₂ + Na⁺ [b]. **Figure 5B** shows PNGase – released glycans from K3 wild type. The *N*-glycans shown are as follows: Man₉GlcNAc₂ [d];

Man₁₀GlcNAc₂ [e]; Man₁₁GlcNAc₂ [f]; Man₁₂GlcNAc₂ [g]. Figure 5C depicts the *och1* deletion resulting in the production of Man₈GlcNAc₂ [c] as the predominant N-glycan. Figures 5D and 5E show the production of Man₅GlcNAc₂ [b] after *in vivo* trimming of Man₈GlcNAc₂ with a chimeric α -1,2-mannosidase. The 5 predominant N-glycan is indicated by a peak with a mass (m/z) of 1253 consistent with its identification as Man₅GlcNAc₂ [b].

[0058] Figures 6A – 6F show MALDI-TOF analysis demonstrating production of IFN- β glycoproteins having Man₅GlcNAc₂ as the predominant N-glycan structure in *P. pastoris*. Figure 6A shows the standard Man₅GlcNAc₂ [a] and 10 Man₅GlcNAc₂ + Na⁺ [b] as the standard (Glyko, Novato, CA). Figure 6B shows PNGase – released glycans from IFN- β wildtype. Figure 6C depicts the *och1* knock-out producing Man₈GlcNAc₂ [c]; Man₉GlcNAc₂ [d]; Man₁₀GlcNAc₂ [e]; Man₁₁GlcNAc₂ [f]; Man₁₂GlcNAc₂ [g]; and no production of Man₅GlcNAc₂ [b]. Figure 6D shows relatively small amount of Man₅GlcNAc₂ [b] among other 15 intermediate N-glycans Man₈GlcNAc₂ [c] to Man₁₂GlcNAc₂ [g]. Figure 6E shows a significant amount of Man₅GlcNAc₂ [b] relative to the other glycans Man₈GlcNAc₂ [c] and Man₉GlcNAc₂ [d] produced by pGC5 (*Saccharomyces MNS1*(m)/mouse mannosidase IB Δ 99). Figure 6F shows predominant production of Man₅GlcNAc₂ [b] on the secreted glycoprotein IFN- β by pFB8 (*Saccharomyces SEC12* (m)/mouse mannosidase IA Δ 187). The N-glycan is indicated by a peak 20 with a mass (m/z) of 1254 consistent with its identification as Man₅GlcNAc₂ [b].

[0059] Figure 7 shows a high performance liquid chromatogram for: (A) Man₉GlcNAc₂ standard labeled with 2-AB (negative control); (B) supernatant of medium *P. pastoris*, Δ *och1* transformed with pFB8 mannosidase, which 25 demonstrates a lack of extracellular mannosidase activity in the supernatant; and (C) Man₉GlcNAc₂ standard labeled with 2-AB after exposure to *T. reesei* mannosidase (positive control).

[0060] Figure 8 shows a high performance liquid chromatogram for: (A) Man₉GlcNAc₂ standard labeled with 2-AB (negative control); (B) supernatant of 30 medium *P. pastoris*, Δ *och1* transformed with pGC5 mannosidase, which demonstrates a lack of extracellular mannosidase activity in the supernatant; and

(C) $\text{Man}_9\text{GlcNAc}_2$ standard labeled with 2-AB after exposure to *T. reesei* mannosidase (positive control).

[0061] Figure 9 shows a high performance liquid chromatogram for: (A) $\text{Man}_9\text{GlcNAc}_2$ standard labeled with 2-AB (negative control); (B) supernatant of medium *P. pastoris*, *Aoch1* transformed with pBC18-5 mannosidase, which demonstrates lack of extracellular mannosidase activity in the supernatant; and (C) supernatant of medium *P. pastoris*, *Aoch1* transformed with pDD28-3, which demonstrates activity in the supernatant (positive control).

[0062] Figures 10A – 10B demonstrate the activity of an UDP-GlcNAc transporter in the production of GlcNAcMan₅GlcNAc₂ in *P. pastoris*. Figure 10A depicts a *P. pastoris* strain (YSH-3) with a human GnTI but without the UDP-GlcNAc transporter resulting in some production of GlcNAcMan₅GlcNAc₂ [b] but a predominant production of Man₅GlcNAc₂ [a]. Figure 10B depicts the addition of UDP-GlcNAc transporter from *K. lactis* in a strain (PEP-3) with the human 15 GnTI, which resulted in the predominant production of GlcNAcMan₅GlcNAc₂ [b]. The single prominent peak of mass (m/z) at 1457 is consistent with its identification as GlcNAcMan₅GlcNAc₂ [b] as shown in Figure 10B.

[0063] Figure 11 shows a pH optimum of a heterologous mannosidase enzyme encoded by pBB27-2 (*Saccharomyces MNN10* (s)/*C. elegans* mannosidase IB Δ31) expressed in *P. pastoris*.

[0064] Figures 12A – 12C show MALDI-TOF analysis of *N*-glycans released from a cell free extract of *K. lactis*. Figure 12A shows the *N*-glycans released from wild-type cells, which includes high-mannose type *N*-glycans. Figure 12B shows the *N*-glycans released from *och1 mnn1* deleted cells, revealing a distinct 25 peak of mass (m/z) at 1908 consistent with its identification as $\text{Man}_9\text{GlcNAc}_2$ [d]. Figure 12C shows the *N*-glycans released from *och1 mnn1* deleted cells after *in vitro* α-1,2-mannosidase digest corresponding to a peak consistent with Man₅GlcNAc₂.

[0065] Figure 13 is a schematic of the structure of the dolichyl pyrophosphate-linked oligosaccharide.

[0066] Figure 14 is a schematic of the generation of GlcNAc₂Man₃GlcNAc₂ *N*-glycans from fungal host cells which are deficient in *alg3*, *alg9*, or *alg12* activities.

[0067] **Figure 15** is a schematic of processing reactions required to produce mammalian-type oligosaccharide structures in a fungal host cell with an *alg3*, *och1* genotype.

[0068] **Figure 16** shows *S. cerevisiae Alg3* Sequence Comparisons (Blast) (SEQ ID NOs:9 - 20, respectively, in order of appearance)

[0069] **Figure 17** shows *S. cerevisiae ALG3* (SEQ ID NO:21) and Alg3p (SEQ ID NO:22) Sequences

[0070] **Figure 18** shows *P. pastoris ALG3* (SEQ ID NO:23) and Alg3p (SEQ ID NO:24) Sequences

[0071] **Figure 19** shows *P. pastoris ALG3* Sequence Comparisons (Blast) (SEQ ID NOs:23 - 31, respectively, in order of appearance)

[0072] **Figure 20** shows *K. lactis ALG3* (SEQ ID NO:33) and Alg3p (SEQ ID NO:34) Sequences

[0073] **Figure 21** shows *K. lactis ALG3* Sequence Comparisons (Blast) (SEQ ID NOs:35 - 40, respectively, in order of appearance)

[0074] **Figure 22** shows a model of an IgG immunoglobulin. Heavy chain and light chain can be, based on similar secondary and tertiary structure, subdivided into domains. The two heavy chains (domains V_H, C_{H1}, C_{H2} and C_{H3}) are linked through three disulfide bridges. The light chains (domains V_L and C_L) are linked by another disulfide bridge to the C_{H1} portion of the heavy chain and, together with the C_{H1} and V_H fragments, make up the Fab region. Antigens bind to the terminal portion of the Fab region. Effector-functions, such as Fc-gamma-Receptor binding have been localized to the C_{H2} domain, just downstream of the hinge region and are influenced by N-glycosylation of asparagine 297 in the heavy chain.

[0075] **Figure 23** is a schematic overview of a modular IgG1 expression vector.

[0076] **Figure 24** shows *M. musculus GnTIII* Nucleic Acid (SEQ ID NO:45) And Amino Acid (SEQ ID NO:46) Sequences

[0077] **Figure 25** (top) is a MALDI-TOF-MS analysis of N-glycans isolated from a kringle 3 glycoprotein produced in a *P. pastoris* YSH-1 displaying a predominant peak at 1461 m/z corresponding to the mass of GlcNAcMan₅GlcNAc₂ [d]; **Figure 25** (bottom) shows a MALDI-TOF-MS analysis of N-glycans isolated from a kringle 3 glycoprotein produced in a *P.*

pastoris YSH-1 transformed with *D. melanogaster* mannosidase II Δ 74/S.

cerevisiae MNN2(s) showing a predominant peak at 1140 m/z corresponding to the mass of GlcNAcMan₃GlcNAc₂ [b] and other peaks corresponding to GlcNAcMan₄GlcNAc₂ [c] at 1303 m/z and GlcNAcMan₅GlcNAc₂ [d] at 1465 m/z.

5 This strain was designated YSH-37.

[0078] Figure 26 (top) is the MALDI-TOF-MS analysis of N-glycans isolated from a kringle 3 glycoprotein produced in *P. pastoris* YSH-1 as shown in Figure 25 (top); Figure 26 (bottom) is a MALDI-TOF-MS analysis of N-glycans isolated from a kringle 3 glycoprotein expressed in *P. pastoris* YSH-1 cells transformed 10 with a pVA53 construct (*S. cerevisiae* MNN2(s)/mGnTIII). The peak at 1463 m/z corresponds the mass of GlcNAcMan₅GlcNAc₂ [d] and the peak at 1666 m/z corresponds to the mass of GlcNAc₂Man₅GlcNAc₂ [a].

[0079] Figure 27 (top) is the MALDI-TOF-MS analysis of N-glycans isolated from a kringle 3 glycoprotein produced in *P. pastoris* YSH-1 as shown in Figure 15 25 (top); Figure 27 (bottom) is a MALDI-TOF-MS analysis of N-glycans isolated from a kringle 3 glycoprotein expressed in *P. pastoris* YSH-1 cells transformed with a pVA55 construct (*S. cerevisiae* MNN2(s)/mGnTIII). The peak at 1463 m/z corresponds to the mass of GlcNAcMan₅GlcNAc₂ [d] and the peak at 1667 m/z corresponds to the mass of GlcNAc₂Man₅GlcNAc₂ [a].

20 [0080] Figure 28 (top) is the MALDI-TOF-MS analysis of N-glycans isolated from a kringle 3 glycoprotein produced in *P. pastoris* YSH-1 as shown in Figure 25 (top); Figure 28 (bottom) is a MALDI-TOF-MS analysis of N-glycans isolated from a kringle 3 glycoprotein expressed in *P. pastoris* YSH-1 cells transformed with a pVB51 construct (*K. lactis* GNTI(s)/mGnTIII). The predominant peak at 25 1463 m/z corresponds to the mass of GlcNAcMan₅GlcNAc₂ [d] and a second peak at 1726 m/z [e], which does not correspond to the mass of GlcNAc₂Man₅GlcNAc₂ is observed.

[0081] Figure 29 is a MALDI-TOF-MS analysis of N-glycans isolated from a kringle 3 glycoprotein expressed in *P. pastoris* YSH-44 cells. The predominant 30 peak at 1356 m/z corresponds to the mass of GlcNAc₂Man₃GlcNAc₂ [x].

[0082] Figure 30 is a MALDI-TOF-MS analysis of N-glycans isolated from a kringle 3 glycoprotein expressed in *P. pastoris* YSH-44 cells transformed with a

pVA53 construct (*S. cerevisiae* MNN2(s)/mGnTIII). The peak at 1340 m/z corresponds to the mass of GlcNAc₂Man₃GlcNAc₂ [x] and the peak at 1542 m/z corresponds to the mass of GlcNAc₃Man₃GlcNAc₂ [y].

[0083] Figure 31 is a MALDI-TOF-MS analysis of *N*-glycans isolated from a 5 kringle 3 glycoprotein expressed in *P. pastoris* PBP6-5 cells. The predominant peak at 1340 m/z corresponds to the mass of GlcNAc₂Man₃GlcNAc₂ [x].

[0084] Figure 32 is a MALDI-TOF-MS analysis of *N*-glycans isolated from a kringle 3 glycoprotein expressed in *P. pastoris* PBP6-5 cells transformed with a 10 pVA53 construct (*S. cerevisiae* MNN2(s)/mGnTIII). The peak at 1340 m/z corresponds to the mass of GlcNAc₂Man₃GlcNAc₂ [x] and the peak at 1543 m/z corresponds to the mass of GlcNAc₃Man₃GlcNAc₂ [y].

[0085] Figure 33 shows a high performance liquid chromatogram, which 15 demonstrates a lack of extracellular GnTIII activity (pVA53) in the supernatant. The *N*-glycan GlcNAcMan₅GlcNAc₂ purified from K3 expressed in PBP-3 strain was added to: BMMY (A); 1 mM UDP-GlcNAc (Sigma Chemical Co., St. Louis, 20 MO)) in BMMY (B); the supernatant of YSH-44 transformed with pVA53 [YSH-57] (C); and the supernatant of YSH-57 + 1 mM UDP-GlcNAc (D).

[0086] Figure 34 shows a high performance liquid chromatogram, which 25 demonstrates a lack of extracellular GnTIII activity (pVA53) in the supernatant. The *N*-glycan GlcNAc₂Man₃GlcNAc₂ purified from K3 expressed in YSH-44 strain was added to: BMMY (A); 1 mM UDP-GlcNAc (Sigma Chemical Co., St. Louis, MO)) in BMMY (B); and the supernatant of YSH-44 transformed with pVA53 [YSH-57] (C).

[0087] Figure 35 is a schematic diagram comparing the normal glycosylation 30 pathways in humans and *P. pastoris* (Panel A) with an engineered humanized *N*-glycosylation pathway in lower eukaryotes (Panel B). The engineered pathway represents the construction of *P. pastoris* strain PBP6-5, which after modification with GnTIII becomes *P. pastoris* strain PBP38.

[0088] Figure 36 is a schematic diagram showing the predominant secreted 35 glycoform produced by each of the designated *P. pastoris* strains and the gene modification used to engineer each of the strains.

[0089] **Figure 37** is a structural representation of the transfer of a GlcNAc to the oligosaccharide intermediate, GlcNAcMan₅GlcNAc₂, produced on glycoproteins in a lower eukaryotic host cell, as catalyzed by GnTIII.

[0090] **Figure 38** is a structural representation of the transfer of a GlcNAc to the 5 oligosaccharide intermediate, GlcNAcMan₃GlcNAc₂, produced on glycoproteins in a lower eukaryotic host cell, as catalyzed by GnTII, and the subsequent transfer of a GlcNAc to the product of that reaction, GlcNAc₂Man₃GlcNAc₂, as catalyzed by GnTIII.

[0091] **Figure 39** is a schematic diagram showing the transfer of GlcNAc 10 residues onto oligosaccharide intermediates catalyzed by GnTIV, GnTV, GnTVI and GnTIX in *P.pastoris*.

[0092] **Figure 40** shows three representative plasmid maps **Figure 40A: pPB144** containing a gene fragment encoding mouse GnTIV; **Figure 40B: pPB140** containing a gene fragment encoding human GnTV; and **Figure 40C: pPB176** 15 containing a gene fragment encoding mouse GnTIX used for transformation in a host *P.pastoris*.

[0093] **Figure 41** shows the gene encoding the *Homo sapiens* mannosyl (alpha-1,3-)glycoprotein beta-1,4-N-acetylglucosaminyltransferase, isoenzyme A (MGAT4A) Accession number NM_012214.

[0094] **Figure 42** shows the gene encoding the *Homo sapiens* mannosyl (alpha-1,3-)glycoprotein beta-1,4-N-acetylglucosaminyltransferase, isoenzyme B (MGAT4B) Accession number NM_014275.

[0095] **Figure 43** shows the gene encoding the *Mus musculus* N-acetylglucosaminyltransferase V (Mgat5) Accession number AF474154.

[0096] **Figure 44** shows the gene encoding the *Gallus gallus* N-acetylglucosaminyltransferase VI Accession number AB040608.

[0097] **Figure 45** shows the gene encoding the *Homo sapiens* N-acetylglucosaminyltransferase IX Accession number AB109185.1.

[0098] **Figure 46** shows the codon optimized DNA fragment encoding part of 30 the human N-acetylglucosaminyltransferase IX lacking the TM domain (Δ 43).

[0099] **Figure 47** is a MALDI-TOF-MS analysis of N-glycans isolated from a kringle 3 glycoprotein expressed in *P.pastoris* PBP43. The yeast strain *P.*

pastoris YSH-44 was transformed with pPB144 containing the *S. cerevisiae* MNN2(s)/human GnTIV fusion construct. The peak at 1543 m/z corresponds to the mass of GlcNAc₃Man₃GlcNAc₂ [y].

[0100] Figure 48 is a MALDI-TOF-MS analysis of N-glycans isolated from a 5 kringle 3 glycoprotein expressed in *P. pastoris* PBP32. The yeast strain *P. pastoris* YSH-44 was transformed with pPB140 containing the *S. cerevisiae* MNN2(s)/mouse GnTV fusion. The peak at 1559 m/z corresponds to the mass of GlcNAc₃Man₃GlcNAc₂ [y].

[0101] Figure 49 is a MALDI-TOF-MS analysis of N-glycans isolated from a 10 kringle 3 glycoprotein expressed in *P. pastoris* PBP46. The yeast strain *P. pastoris* YSH-44 was transformed with pPB140 and pPB144. The peak at 1543 m/z corresponds to the mass of GlcNAc₃Man₃GlcNAc₂ [y] and the peak at 1747 m/z corresponds to the mass of GlcNAc₄Man₃GlcNAc₂ [z].

[0102] Figure 50 is a MALDI-TOF-MS analysis of N-glycans isolated from a 15 kringle 3 glycoprotein expressed in *P. pastoris* PBP94. The yeast strain *P. pastoris* YSH-44 was transformed with pPB128 containing the *S. cerevisiae* MNN2(s)/mouse GnTIVA fusion construct and pPB140 containing the *S. cerevisiae* MNN2(s)/mouse GnTV fusion construct. The predominant peak at 1743 m/z corresponds to the mass of GlcNAc₄Man₃GlcNAc₂ [z].

20

DETAILED DESCRIPTION OF THE INVENTION

[0103] Unless otherwise defined herein, scientific and technical terms used in connection with the present invention shall have the meanings that are commonly understood by those of ordinary skill in the art. Further, unless otherwise required 25 by context, singular terms shall include pluralities and plural terms shall include the singular. The methods and techniques of the present invention are generally performed according to conventional methods well known in the art. Generally, nomenclatures used in connection with, and techniques of biochemistry, enzymology, molecular and cellular biology, microbiology, genetics and protein 30 and nucleic acid chemistry and hybridization described herein are those well-known and commonly used in the art.

[0104] The methods and techniques of the present invention are generally performed according to conventional methods well-known in the art and as described in various general and more specific references that are cited and discussed throughout the present specification unless otherwise indicated. See,

5 e.g., Sambrook *et al.*, *Molecular Cloning: A Laboratory Manual*, 2d ed., Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y. (1989); Ausubel *et al.*, *Current Protocols in Molecular Biology*, Greene Publishing Associates (1992, and Supplements to 2002); Harlow and Lane, *Antibodies: A Laboratory Manual*, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y. (1990); *Introduction to Glycobiology*, Maureen E. Taylor, Kurt Drickamer, Oxford Univ. Press (2003);
10 Worthington Enzyme Manual, Worthington Biochemical Corp. Freehold, NJ; *Handbook of Biochemistry: Section A Proteins*, Vol I 1976 CRC Press; *Handbook of Biochemistry: Section A Proteins*, Vol II 1976 CRC Press; *Essentials of Glycobiology*, Cold Spring Harbor Laboratory Press (1999). The nomenclatures
15 used in connection with, and the laboratory procedures and techniques of, molecular and cellular biology, protein biochemistry, enzymology and medicinal and pharmaceutical chemistry described herein are those well known and commonly used in the art.

[0105] All publications, patents and other references mentioned herein are
20 incorporated by reference.

[0106] The following terms, unless otherwise indicated, shall be understood to have the following meanings:

[0107] As used herein, the term “N-glycan” refers to an N-linked oligosaccharide, e.g., one that is attached by an asparagine-N-acetylglucosamine
25 linkage to an asparagine residue of a polypeptide. N-glycans have a common pentasaccharide core of Man₃GlcNAc₂ (“Man” refers to mannose; “Glc” refers to glucose; and “NAc” refers to N-acetyl; GlcNAc refers to N-acetylglucosamine). The term “trimannose core” used with respect to the N-glycan also refers to the structure Man₃GlcNAc₂ (“Man₃”). The term “pentamannose core” or “Mannose-5
30 core” or “Man₅” used with respect to the N-glycan refers to the structure Man₅GlcNAc₂. N-glycans differ with respect to the number of branches (antennae) comprising peripheral sugars (e.g., GlcNAc, fucose, and sialic acid) that are

attached to the Man₃ core structure. *N*-glycans are classified according to their branched constituents (e.g., high mannose, complex or hybrid).

[0108] A "high mannose" type *N*-glycan has five or more mannose residues. A "complex" type *N*-glycan typically has at least one GlcNAc attached to the 1,3
5 mannose arm and at least one GlcNAc attached to the 1,6 mannose arm of the trimannose core. Complex *N*-glycans may also have galactose ("Gal") residues that are optionally modified with sialic acid or derivatives ("NeuAc", where "Neu" refers to neuraminic acid and "Ac" refers to acetyl). A complex *N*-glycan typically has at least one branch that terminates in an oligosaccharide such as, for example:
10 NeuNAc-; NeuAco2-6GalNAc α 1-; NeuAco2-3Gal β 1-3GalNAc α 1-; NeuAco2-3/6Gal β 1-4GlcNAc β 1-; GlcNAc α 1-4Gal β 1-(mucins only); Fuc α 1-2Gal β 1-(blood group H). Sulfate esters can occur on galactose, GalNAc, and GlcNAc residues, and phosphate esters can occur on mannose residues. NeuAc (Neu: neuraminic acid; Ac:acetyl) can be *O*-acetylated or replaced by NeuGI (*N*-glycolylneuraminic acid). Complex *N*-glycans may also have intrachain substitutions comprising
15 "bisecting" GlcNAc and core fucose ("Fuc"). A "hybrid" *N*-glycan has at least one GlcNAc on the terminal of the 1,3 mannose arm of the trimannose core and zero or more mannoses on the 1,6 mannose arm of the trimannose core.

[0109] The term "predominant" or "predominantly" used with respect to the
20 production of *N*-glycans refers to a structure which represents the major peak detected by matrix assisted laser desorption ionization time of flight mass spectrometry (MALDI-TOF) analysis.

[0110] Abbreviations used herein are of common usage in the art, *see, e.g.*, abbreviations of sugars, above. Other common abbreviations include "PNGase",
25 which refers to peptide *N*-glycosidase F (EC 3.2.2.18); "GlcNAc Tr" or "GnT," which refers to *N*-acetylglucosaminyl Transferase enzymes; "NANA" refers to *N*-acetylneuraminic acid.

[0111] As used herein, a "humanized glycoprotein" or a "human-like glycoprotein" refers alternatively to a protein having attached thereto *N*-glycans
30 having fewer than four mannose residues, and synthetic glycoprotein intermediates (which are also useful and can be manipulated further *in vitro* or *in vivo*) having at least five mannose residues. Preferably, glycoproteins produced according to the

invention contain at least 30 mole %, preferably at least 40 mole % and more preferably 50, 60, 70, 80, 90, or even 100 mole % of the $\text{Man}_5\text{GlcNAc}_2$ intermediate, at least transiently. This may be achieved, *e.g.*, by engineering a host cell of the invention to express a “better”, *i.e.*, a more efficient glycosylation

5 enzyme. For example, a mannosidase is selected such that it will have optimal activity under the conditions present at the site in the host cell where proteins are glycosylated and is introduced into the host cell preferably by targeting the enzyme to a host cell organelle where activity is desired.

[0112] The term “enzyme”, when used herein in connection with altering host
10 cell glycosylation, refers to a molecule having at least one enzymatic activity, and includes full-length enzymes, catalytically active fragments, chimerics, complexes, and the like. A “catalytically active fragment” of an enzyme refers to a polypeptide having a detectable level of functional (enzymatic) activity. Enzyme activity is “substantially intracellular” when less than 10% of the enzyme activity
15 is measurable outside the cell compared to that measurable from lysed cells.

[0113] A lower eukaryotic host cell, when used herein in connection with glycosylation profiles, refers to any eukaryotic cell which ordinarily produces high mannose containing *N*-glycans, and thus is meant to include some animal or plant cells and most typical lower eukaryotic cells, including uni- and multicellular
20 fungal and algal cells.

[0114] As used herein, the term “secretion pathway” refers to the assembly line of various glycosylation enzymes to which a lipid-linked oligosaccharide precursor and an *N*-glycan substrate are sequentially exposed, following the molecular flow of a nascent polypeptide chain from the cytoplasm to the endoplasmic reticulum
25 (ER) and the compartments of the Golgi apparatus. Enzymes are said to be localized along this pathway. An enzyme X that acts on a lipid-linked glycan or an *N*-glycan before enzyme Y is said to be or to act “upstream” to enzyme Y; similarly, enzyme Y is or acts “downstream” from enzyme X.

[0115] The term “targeting peptide” as used herein refers to nucleotide or amino
30 acid sequences encoding a cellular targeting signal peptide which mediates the localization (or retention) of an associated sequence to sub-cellular locations, *e.g.*, organelles.

[0116] The term "polynucleotide" or "nucleic acid molecule" refers to a polymeric form of nucleotides of at least 10 bases in length. The term includes DNA molecules (e.g., cDNA or genomic or synthetic DNA) and RNA molecules (e.g., mRNA or synthetic RNA), as well as analogs of DNA or RNA containing 5 non-natural nucleotide analogs, non-native internucleoside bonds, or both. The nucleic acid can be in any topological conformation. For instance, the nucleic acid can be single-stranded, double-stranded, triple-stranded, quadruplexed, partially double-stranded, branched, hairpinned, circular, or in a padlocked conformation. The term includes single and double stranded forms of DNA. A nucleic acid 10 molecule of this invention may include both sense and antisense strands of RNA, cDNA, genomic DNA, and synthetic forms and mixed polymers of the above. They may be modified chemically or biochemically or may contain non-natural or derivatized nucleotide bases, as will be readily appreciated by those of skill in the art. Such modifications include, for example, labels, methylation, substitution of 15 one or more of the naturally occurring nucleotides with an analog, internucleotide modifications such as uncharged linkages (e.g., methyl phosphonates, phosphotriesters, phosphoramidates, carbamates, etc.), charged linkages (e.g., phosphorothioates, phosphorodithioates, etc.), pendent moieties (e.g., polypeptides), intercalators (e.g., acridine, psoralen, etc.), chelators, alkylators, and 20 modified linkages (e.g., alpha anomeric nucleic acids, etc.) Also included are synthetic molecules that mimic polynucleotides in their ability to bind to a designated sequence via hydrogen bonding and other chemical interactions. Such molecules are known in the art and include, for example, those in which peptide linkages substitute for phosphate linkages in the backbone of the molecule.

25 [0117] Unless otherwise indicated, a "nucleic acid comprising SEQ ID NO:X" refers to a nucleic acid, at least a portion of which has either (i) the sequence of SEQ ID NO:X, or (ii) a sequence complementary to SEQ ID NO:X. The choice between the two is dictated by the context. For instance, if the nucleic acid is used as a probe, the choice between the two is dictated by the requirement that the probe 30 be complementary to the desired target.

[0118] An "isolated" or "substantially pure" nucleic acid or polynucleotide (e.g., an RNA, DNA or a mixed polymer) is one which is substantially separated from

other cellular components that naturally accompany the native polynucleotide in its natural host cell, e.g., ribosomes, polymerases, and genomic sequences with which it is naturally associated. The term embraces a nucleic acid or polynucleotide that (1) has been removed from its naturally occurring environment, (2) is not 5 associated with all or a portion of a polynucleotide in which the "isolated polynucleotide" is found in nature, (3) is operatively linked to a polynucleotide which it is not linked to in nature, or (4) does not occur in nature. The term "isolated" or "substantially pure" also can be used in reference to recombinant or cloned DNA isolates, chemically synthesized polynucleotide analogs, or 10 polynucleotide analogs that are biologically synthesized by heterologous systems.

[0119] However, "isolated" does not necessarily require that the nucleic acid or polynucleotide so described has itself been physically removed from its native environment. For instance, an endogenous nucleic acid sequence in the genome of an organism is deemed "isolated" herein if a heterologous sequence (*i.e.*, a 15 sequence that is not naturally adjacent to this endogenous nucleic acid sequence) is placed adjacent to the endogenous nucleic acid sequence, such that the expression of this endogenous nucleic acid sequence is altered. By way of example, a non-native promoter sequence can be substituted (*e.g.*, by homologous recombination) for the native promoter of a gene in the genome of a human cell, such that this 20 gene has an altered expression pattern. This gene would now become "isolated" because it is separated from at least some of the sequences that naturally flank it.

[0120] A nucleic acid is also considered "isolated" if it contains any 25 modifications that do not naturally occur to the corresponding nucleic acid in a genome. For instance, an endogenous coding sequence is considered "isolated" if it contains an insertion, deletion or a point mutation introduced artificially, *e.g.*, by human intervention. An "isolated nucleic acid" also includes a nucleic acid integrated into a host cell chromosome at a heterologous site, a nucleic acid construct present as an episome. Moreover, an "isolated nucleic acid" can be substantially free of other cellular material, or substantially free of culture medium 30 when produced by recombinant techniques, or substantially free of chemical precursors or other chemicals when chemically synthesized.

[0121] As used herein, the phrase "degenerate variant" of a reference nucleic acid sequence encompasses nucleic acid sequences that can be translated, according to the standard genetic code, to provide an amino acid sequence identical to that translated from the reference nucleic acid sequence.

5 [0122] The term "percent sequence identity" or "identical" in the context of nucleic acid sequences refers to the residues in the two sequences which are the same when aligned for maximum correspondence. The length of sequence identity comparison may be over a stretch of at least about nine nucleotides, usually at least about 20 nucleotides, more usually at least about 24 nucleotides, typically at least
10 about 28 nucleotides, more typically at least about 32 nucleotides, and preferably at least about 36 or more nucleotides. There are a number of different algorithms known in the art which can be used to measure nucleotide sequence identity. For instance, polynucleotide sequences can be compared using FASTA, Gap or Bestfit, which are programs in Wisconsin Package Version 10.0, Genetics Computer
15 Group (GCG), Madison, Wisconsin. FASTA provides alignments and percent sequence identity of the regions of the best overlap between the query and search sequences (Pearson (1990) *Methods Enzymol.* 183:63-98, incorporated herein by reference in its entirety). For instance, percent sequence identity between nucleic acid sequences can be determined using FASTA with its default parameters (a
20 word size of 6 and the NOPAM factor for the scoring matrix) or using Gap with its default parameters as provided in GCG Version 6.1, herein incorporated by reference.

[0123] The term "substantial homology" or "substantial similarity," when referring to a nucleic acid or fragment thereof, indicates that, when optimally
25 aligned with appropriate nucleotide insertions or deletions with another nucleic acid (or its complementary strand), there is nucleotide sequence identity in at least about 50%, more preferably 60% of the nucleotide bases, usually at least about 70%, more usually at least about 80%, preferably at least about 90%, and more preferably at least about 95%, 96%, 97%, 98% or 99% of the nucleotide bases, as
30 measured by any well-known algorithm of sequence identity, such as FASTA, BLAST or Gap, as discussed above.

[0124] Alternatively, substantial homology or similarity exists when a nucleic acid or fragment thereof hybridizes to another nucleic acid, to a strand of another nucleic acid, or to the complementary strand thereof, under stringent hybridization conditions. "Stringent hybridization conditions" and "stringent wash conditions" 5 in the context of nucleic acid hybridization experiments depend upon a number of different physical parameters. Nucleic acid hybridization will be affected by such conditions as salt concentration, temperature, solvents, the base composition of the hybridizing species, length of the complementary regions, and the number of nucleotide base mismatches between the hybridizing nucleic acids, as will be 10 readily appreciated by those skilled in the art. One having ordinary skill in the art knows how to vary these parameters to achieve a particular stringency of hybridization.

[0125] In general, "stringent hybridization" is performed at about 25°C below the thermal melting point (T_m) for the specific DNA hybrid under a particular set of 15 conditions. "Stringent washing" is performed at temperatures about 5°C lower than the T_m for the specific DNA hybrid under a particular set of conditions. The T_m is the temperature at which 50% of the target sequence hybridizes to a perfectly matched probe. See Sambrook *et al.*, *supra*, page 9.51, hereby incorporated by reference. For purposes herein, "high stringency conditions" are defined for 20 solution phase hybridization as aqueous hybridization (*i.e.*, free of formamide) in 6X SSC (where 20X SSC contains 3.0 M NaCl and 0.3 M sodium citrate), 1% SDS at 65°C for 8-12 hours, followed by two washes in 0.2X SSC, 0.1% SDS at 65°C for 20 minutes. It will be appreciated by the skilled artisan that hybridization at 65°C will occur at different rates depending on a number of factors including the 25 length and percent identity of the sequences which are hybridizing.

[0126] The term "mutated" when applied to nucleic acid sequences means that nucleotides in a nucleic acid sequence may be inserted, deleted or changed compared to a reference nucleic acid sequence. A single alteration may be made at a locus (a point mutation) or multiple nucleotides may be inserted, deleted or 30 changed at a single locus. In addition, one or more alterations may be made at any number of loci within a nucleic acid sequence. A nucleic acid sequence may be mutated by any method known in the art including but not limited to mutagenesis

techniques such as "error-prone PCR" (a process for performing PCR under conditions where the copying fidelity of the DNA polymerase is low, such that a high rate of point mutations is obtained along the entire length of the PCR product.

See, e.g., Leung et al. (1989) Technique 1:11-15 and Caldwell and Joyce (1992)

5 *PCR Methods Applic.* 2:28-33); and "oligonucleotide-directed mutagenesis" (a process which enables the generation of site-specific mutations in any cloned DNA segment of interest. *See, e.g., Reidhaar-Olson et al. (1988) Science 241:53-57).*

[0127] The term "vector" as used herein is intended to refer to a nucleic acid molecule capable of transporting another nucleic acid to which it has been linked.

10 One type of vector is a "plasmid", which refers to a circular double stranded DNA loop into which additional DNA segments may be ligated. Other vectors include cosmids, bacterial artificial chromosomes (BAC) and yeast artificial chromosomes (YAC). Another type of vector is a viral vector, wherein additional DNA segments may be ligated into the viral genome (discussed in more detail below). Certain

15 vectors are capable of autonomous replication in a host cell into which they are introduced (*e.g.,* vectors having an origin of replication which functions in the host cell). Other vectors can be integrated into the genome of a host cell upon introduction into the host cell, and are thereby replicated along with the host genome. Moreover, certain preferred vectors are capable of directing the

20 expression of genes to which they are operatively linked. Such vectors are referred to herein as "recombinant expression vectors" (or simply, "expression vectors").

[0128] "Operatively linked" expression control sequences refers to a linkage in which the expression control sequence is contiguous with the gene of interest to control the gene of interest, as well as expression control sequences that act in

25 *trans* or at a distance to control the gene of interest.

[0129] The term "expression control sequence" as used herein refers to polynucleotide sequences which are necessary to affect the expression of coding sequences to which they are operatively linked. Expression control sequences are sequences which control the transcription, post-transcriptional events and

30 translation of nucleic acid sequences. Expression control sequences include appropriate transcription initiation, termination, promoter and enhancer sequences; efficient RNA processing signals such as splicing and polyadenylation signals;

sequences that stabilize cytoplasmic mRNA; sequences that enhance translation efficiency (*e.g.*, ribosome binding sites); sequences that enhance protein stability; and when desired, sequences that enhance protein secretion. The nature of such control sequences differs depending upon the host organism; in prokaryotes, such

5 control sequences generally include promoter, ribosomal binding site, and transcription termination sequence. The term "control sequences" is intended to include, at a minimum, all components whose presence is essential for expression, and can also include additional components whose presence is advantageous, for example, leader sequences and fusion partner sequences.

10 [0130] The term "recombinant host cell" (or simply "host cell"), as used herein, is intended to refer to a cell into which a nucleic acid such as a recombinant vector has been introduced. It should be understood that such terms are intended to refer not only to the particular subject cell but to the progeny of such a cell. Because certain modifications may occur in succeeding generations due to either mutation

15 or environmental influences, such progeny may not, in fact, be identical to the parent cell, but are still included within the scope of the term "host cell" as used herein. A recombinant host cell may be an isolated cell or cell line grown in culture or may be a cell which resides in a living tissue or organism.

[0131] The term "peptide" as used herein refers to a short polypeptide, *e.g.*, one

20 that is typically less than about 50 amino acids long and more typically less than about 30 amino acids long. The term as used herein encompasses analogs and mimetics that mimic structural and thus biological function.

[0132] The term "polypeptide" as used herein encompasses both naturally-occurring and non-naturally-occurring proteins, and fragments, mutants,

25 derivatives and analogs thereof. A polypeptide may be monomeric or polymeric. Further, a polypeptide may comprise a number of different domains each of which has one or more distinct activities.

[0133] The term "isolated protein" or "isolated polypeptide" is a protein or polypeptide that by virtue of its origin or source of derivation (1) is not associated

30 with naturally associated components that accompany it in its native state, (2) when it exists in a purity not found in nature, where purity can be adjudged with respect to the presence of other cellular material (*e.g.*, is free of other proteins from

the same species) (3) is expressed by a cell from a different species, or (4) does not occur in nature (e.g., it is a fragment of a polypeptide found in nature or it includes amino acid analogs or derivatives not found in nature or linkages other than standard peptide bonds). Thus, a polypeptide that is chemically synthesized or 5 synthesized in a cellular system different from the cell from which it naturally originates will be "isolated" from its naturally associated components. A polypeptide or protein may also be rendered substantially free of naturally associated components by isolation, using protein purification techniques well-known in the art. As thus defined, "isolated" does not necessarily require that the 10 protein, polypeptide, peptide or oligopeptide so described has been physically removed from its native environment.

[0134] The term "polypeptide fragment" as used herein refers to a polypeptide that has an amino-terminal and/or carboxy-terminal deletion compared to a full-length polypeptide. In a preferred embodiment, the polypeptide fragment is a 15 contiguous sequence in which the amino acid sequence of the fragment is identical to the corresponding positions in the naturally-occurring sequence. Fragments typically are at least 5, 6, 7, 8, 9 or 10 amino acids long, preferably at least 12, 14, 16 or 18 amino acids long, more preferably at least 20 amino acids long, more preferably at least 25, 30, 35, 40 or 45, amino acids, even more preferably at least 20 50 or 60 amino acids long, and even more preferably at least 70 amino acids long.

[0135] A "modified derivative" refers to polypeptides or fragments thereof that are substantially homologous in primary structural sequence but which include, e.g., *in vivo* or *in vitro* chemical and biochemical modifications or which incorporate amino acids that are not found in the native polypeptide. Such 25 modifications include, for example, acetylation, carboxylation, phosphorylation, glycosylation, ubiquitination, labeling, e.g., with radionuclides, and various enzymatic modifications, as will be readily appreciated by those well skilled in the art. A variety of methods for labeling polypeptides and of substituents or labels useful for such purposes are well-known in the art, and include radioactive isotopes 30 such as ^{125}I , ^{32}P , ^{35}S , and ^3H , ligands which bind to labeled antiligands (e.g., antibodies), fluorophores, chemiluminescent agents, enzymes, and antiligands which can serve as specific binding pair members for a labeled ligand. The choice

of label depends on the sensitivity required, ease of conjugation with the primer, stability requirements, and available instrumentation. Methods for labeling polypeptides are well-known in the art. See Ausubel *et al.*, *Current Protocols in Molecular Biology*, Greene Publishing Associates (1992, and Supplements to 5 2002), hereby incorporated by reference.

[0136] A "polypeptide mutant" or "mutein" refers to a polypeptide whose sequence contains an insertion, duplication, deletion, rearrangement or substitution of one or more amino acids compared to the amino acid sequence of a native or wild-type protein. A mutein may have one or more amino acid point substitutions, 10 in which a single amino acid at a position has been changed to another amino acid, one or more insertions and/or deletions, in which one or more amino acids are inserted or deleted, respectively, in the sequence of the naturally-occurring protein, and/or truncations of the amino acid sequence at either or both the amino or carboxy termini. A mutein may have the same but preferably has a different 15 biological activity compared to the naturally-occurring protein.

[0137] A mutein has at least 70% overall sequence homology to its wild-type counterpart. Even more preferred are muteins having 80%, 85% or 90% overall sequence homology to the wild-type protein. In an even more preferred embodiment, a mutein exhibits 95% sequence identity, even more preferably 97%, 20 even more preferably 98% and even more preferably 99% overall sequence identity. Sequence homology may be measured by any common sequence analysis algorithm, such as Gap or Bestfit.

[0138] Preferred amino acid substitutions are those which: (1) reduce susceptibility to proteolysis, (2) reduce susceptibility to oxidation, (3) alter binding 25 affinity for forming protein complexes, (4) alter binding affinity or enzymatic activity, and (5) confer or modify other physicochemical or functional properties of such analogs.

[0139] As used herein, the twenty conventional amino acids and their abbreviations follow conventional usage. See *Immunology - A Synthesis* (2nd 30 Edition, E.S. Golub and D.R. Gren, Eds., Sinauer Associates, Sunderland, Mass. (1991)), which is incorporated herein by reference. Stereoisomers (e.g., D-amino acids) of the twenty conventional amino acids, unnatural amino acids such as α ,

α -disubstituted amino acids, *N*-alkyl amino acids, and other unconventional amino acids may also be suitable components for polypeptides of the present invention. Examples of unconventional amino acids include: 4-hydroxyproline, γ -carboxyglutamate, ϵ -*N,N,N*-trimethyllysine, ϵ -*N*-acetyllysine, *O*-phosphoserine, 5 5-*N*-acetylserine, *N*-formylmethionine, 3-methylhistidine, 5-hydroxylysine, *N*-methylarginine, and other similar amino acids and imino acids (e.g., 4-hydroxyproline). In the polypeptide notation used herein, the left-hand direction is the amino terminal direction and the right hand direction is the carboxy-terminal direction, in accordance with standard usage and convention.

10 [0140] A protein has "homology" or is "homologous" to a second protein if the nucleic acid sequence that encodes the protein has a similar sequence to the nucleic acid sequence that encodes the second protein. Alternatively, a protein has homology to a second protein if the two proteins have "similar" amino acid sequences. (Thus, the term "homologous proteins" is defined to mean that the two 15 proteins have similar amino acid sequences). In a preferred embodiment, a homologous protein is one that exhibits 60% sequence homology to the wild type protein, more preferred is 70% sequence homology. Even more preferred are homologous proteins that exhibit 80%, 85% or 90% sequence homology to the wild type protein. In a yet more preferred embodiment, a homologous protein 20 exhibits 95%, 97%, 98% or 99% sequence identity. As used herein, homology between two regions of amino acid sequence (especially with respect to predicted structural similarities) is interpreted as implying similarity in function.

[0141] When "homologous" is used in reference to proteins or peptides, it is recognized that residue positions that are not identical often differ by conservative 25 amino acid substitutions. A "conservative amino acid substitution" is one in which an amino acid residue is substituted by another amino acid residue having a side chain (R group) with similar chemical properties (e.g., charge or hydrophobicity). In general, a conservative amino acid substitution will not substantially change the functional properties of a protein. In cases where two or more amino acid 30 sequences differ from each other by conservative substitutions, the percent sequence identity or degree of homology may be adjusted upwards to correct for the conservative nature of the substitution. Means for making this adjustment are

well known to those of skill in the art (*see, e.g.*, Pearson (1990) *Methods Enzymol.* 183:63-98).

[0142] The following six groups each contain amino acids that are conservative substitutions for one another: 1) Serine (S), Threonine (T); 2) Aspartic Acid (D),

5 Glutamic Acid (E); 3) Asparagine (N), Glutamine (Q); 4) Arginine (R), Lysine (K); 5) Isoleucine (I), Leucine (L), Methionine (M), Alanine (A), Valine (V), and 6) Phenylalanine (F), Tyrosine (Y), Tryptophan (W).

[0143] Sequence homology for polypeptides, which is also referred to as percent sequence identity, is typically measured using sequence analysis software. *See,*

10 *e.g.*, the Sequence Analysis Software Package of the Genetics Computer Group (GCG), University of Wisconsin Biotechnology Center, 910 University Avenue, Madison, Wisconsin 53705. Protein analysis software matches similar sequences using measure of homology assigned to various substitutions, deletions and other modifications, including conservative amino acid substitutions. For instance, GCG 15 contains programs such as "Gap" and "Bestfit" which can be used with default parameters to determine sequence homology or sequence identity between closely related polypeptides, such as homologous polypeptides from different species of organisms or between a wild type protein and a mutein thereof. *See, e.g.*, GCG Version 6.1.

20 [0144] A preferred algorithm when comparing a inhibitory molecule sequence to a database containing a large number of sequences from different organisms is the computer program BLAST (Altschul *et al.* (1990) *J. Mol. Biol.* 215:403-410; Gish and States (1993) *Nature Genet.* 3:266-272; Madden *et al.* (1996) *Meth. Enzymol.* 266:131-141; Altschul *et al.* (1997) *Nucleic Acids Res.* 25:3389-3402; Zhang and 25 Madden (1997) *Genome Res.* 7:649-656), especially blastp or tblastn (Altschul *et al.*, 1997). Preferred parameters for BLASTp are: Expectation value: 10 (default); Filter: seg (default); Cost to open a gap: 11 (default); Cost to extend a gap: 1 (default); Max. alignments: 100 (default); Word size: 11 (default); No. of descriptions: 100 (default); Penalty Matrix: BLOWSUM62.

30 [0145] The length of polypeptide sequences compared for homology will generally be at least about 16 amino acid residues, usually at least about 20 residues, more usually at least about 24 residues, typically at least about 28

residues, and preferably more than about 35 residues. When searching a database containing sequences from a large number of different organisms, it is preferable to compare amino acid sequences. Database searching using amino acid sequences can be measured by algorithms other than blastp known in the art. For instance, 5 polypeptide sequences can be compared using FASTA, a program in GCG Version 6.1. FASTA provides alignments and percent sequence identity of the regions of the best overlap between the query and search sequences (*see* Pearson (1990) *Methods Enzymol.* 183:63-98). For example, percent sequence identity between amino acid sequences can be determined using FASTA with its default parameters 10 (a word size of 2 and the PAM250 scoring matrix), as provided in GCG Version 6.1, herein incorporated by reference.

[0146] The term "fusion protein" refers to a polypeptide comprising a polypeptide or fragment coupled to heterologous amino acid sequences. Fusion proteins are useful because they can be constructed to contain two or more desired 15 functional elements from two or more different proteins. A fusion protein comprises at least 10 contiguous amino acids from a polypeptide of interest, more preferably at least 20 or 30 amino acids, even more preferably at least 40, 50 or 60 amino acids, yet more preferably at least 75, 100 or 125 amino acids. Fusion proteins can be produced recombinantly by constructing a nucleic acid sequence 20 which encodes the polypeptide or a fragment thereof in-frame with a nucleic acid sequence encoding a different protein or peptide and then expressing the fusion protein. Alternatively, a fusion protein can be produced chemically by crosslinking the polypeptide or a fragment thereof to another protein.

[0147] The term "region" as used herein refers to a physically contiguous portion 25 of the primary structure of a biomolecule. In the case of proteins, a region is defined by a contiguous portion of the amino acid sequence of that protein.

[0148] The term "domain" as used herein refers to a structure of a biomolecule that contributes to a known or suspected function of the biomolecule. Domains may be co-extensive with regions or portions thereof; domains may also include 30 distinct, non-contiguous regions of a biomolecule. Examples of protein domains include, but are not limited to, an Ig domain, an extracellular domain, a transmembrane domain, and a cytoplasmic domain.

[0149] As used herein, the term "molecule" means any compound, including, but not limited to, a small molecule, peptide, protein, sugar, nucleotide, nucleic acid, lipid, etc., and such a compound can be natural or synthetic.

[0150] Unless otherwise defined, all technical and scientific terms used herein
5 have the same meaning as commonly understood by one of ordinary skill in the art
to which this invention pertains. Exemplary methods and materials are described
below, although methods and materials similar or equivalent to those described
herein can also be used in the practice of the present invention and will be apparent
to those of skill in the art. All publications and other references mentioned herein
10 are incorporated by reference in their entirety. In case of conflict, the present
specification, including definitions, will control. The materials, methods, and
examples are illustrative only and not intended to be limiting.

[0151] Throughout this specification and claims, the word "comprise" or
variations such as "comprises" or "comprising", will be understood to imply the
15 inclusion of a stated integer or group of integers but not the exclusion of any other
integer or group of integers.

Methods For Producing Human-like Glycoproteins In Lower Eukaryotic Host Cells

20 [0152] The invention provides methods for producing a glycoprotein having
human-like glycosylation in a non-human eukaryotic host cell. As described in
more detail below, a eukaryotic host cell that does not naturally express, or which
is engineered not to express, one or more enzymes involved in production of high
mannose structures is selected as a starting host cell. Such a selected host cell is
25 engineered to express one or more enzymes or other factors required to produce
human-like glycoproteins. A desired host strain can be engineered one enzyme or
more than one enzyme at a time. In addition, a nucleic acid molecule encoding one
or more enzymes or activities may be used to engineer a host strain of the
invention. Preferably, a library of nucleic acid molecules encoding potentially
30 useful enzymes (*e.g.*, chimeric enzymes comprising a catalytically active enzyme
fragment ligated in-frame to a heterologous subcellular targeting sequence) is
created (*e.g.*, by ligation of sub-libraries comprising enzymatic fragments and

subcellular targeting sequences), and a strain having one or more enzymes with optimal activities or producing the most "human-like" glycoproteins may be selected by transforming target host cells with one or more members of the library.

[0153] In particular, the methods described herein enable one to obtain, *in vivo*,

5 $\text{Man}_5\text{GlcNAc}_2$ structures in high yield, at least transiently, for the purpose of further modifying it to yield complex *N*-glycans. A successful scheme to obtain suitable $\text{Man}_5\text{GlcNAc}_2$ structures in appropriate yields in a host cell, such as a lower eukaryotic organism, generally involves two parallel approaches: (1) reducing high mannose structures made by endogenous mannosyltransferase
10 activities, if any, and (2) removing 1,2- α -mannose by mannosidases to yield high levels of suitable $\text{Man}_5\text{GlcNAc}_2$ structures which may be further reacted inside the host cell to form complex, human-like glycoforms.

[0154] Accordingly, a first step involves the selection or creation of a eukaryotic host cell, *e.g.*, a lower eukaryote, capable of producing a specific precursor

15 structure of $\text{Man}_5\text{GlcNAc}_2$ that is able to accept *in vivo* GlcNAc by the action of a GlcNAc transferase I ("GnTI"). In one embodiment, the method involves making or using a non-human eukaryotic host cell depleted in a 1,6 mannosyltransferase activity with respect to the *N*-glycan on a glycoprotein. Preferably, the host cell is depleted in an initiating 1,6 mannosyltransferase activity (see below). Such a host
20 cell will lack one or more enzymes involved in the production of high mannose structures which are undesirable for producing human-like glycoproteins.

[0155] One or more enzyme activities are then introduced into such a host cell to produce *N*-glycans within the host cell characterized by having at least 30 mol % of the $\text{Man}_5\text{GlcNAc}_2$ ("Man₅") carbohydrate structures. $\text{Man}_5\text{GlcNAc}_2$ structures

25 are necessary for complex *N*-glycan formation: $\text{Man}_5\text{GlcNAc}_2$ must be formed *in vivo* in a high yield (*e.g.*, in excess of 30%), at least transiently, as subsequent mammalian- and human-like glycosylation reactions require $\text{Man}_5\text{GlcNAc}_2$ or a derivative thereof.

[0156] This step also requires the formation of a particular isomeric structure of

30 $\text{Man}_5\text{GlcNAc}_2$ within the cell at a high yield. While $\text{Man}_5\text{GlcNAc}_2$ structures are necessary for complex *N*-glycan formation, their presence is by no means sufficient. That is because $\text{Man}_5\text{GlcNAc}_2$ may occur in different isomeric forms,

which may or may not serve as a substrate for GlcNAc transferase I. As most glycosylation reactions are not complete, a particular glycosylated protein generally contains a range of different carbohydrate structures (*i.e.*, glycoforms) on its surface. Thus, the mere presence of trace amounts (*i.e.*, less than 5%) of a 5 particular structure like Man₅GlcNAc₂ is of little practical relevance for producing mammalian- or human-like glycoproteins. It is the formation of a GlcNAc transferase I-accepting Man₅GlcNAc₂ intermediate (Figure 1B) in high yield (*i.e.*, above 30%), which is required. The formation of this intermediate is necessary to enable subsequent *in vivo* synthesis of complex *N*-glycans on glycosylated proteins 10 of interest (target proteins).

[0157] Accordingly, some or all of the Man₅GlcNAc₂ produced by the selected host cell must be a productive substrate for enzyme activities along a mammalian glycosylation pathway, *e.g.*, can serve as a substrate for a GlcNAc transferase I activity *in vivo*, thereby forming the human-like *N*-glycan intermediate 15 GlcNAcMan₅GlcNAc₂ in the host cell. In a preferred embodiment, at least 10%, more preferably at least 30% and most preferably 50% or more of the Man₅GlcNAc₂ intermediate produced in the host cell of the invention is a productive substrate for GnTI *in vivo*. It is understood that if, for example, GlcNAcMan₅GlcNAc₂ is produced at 10% and Man₅GlcNAc₂ is produced at 25% 20 on a target protein, that the total amount of transiently produced Man₅GlcNAc₂ is 35% because GlcNAcMan₅GlcNAc₂ is a product of Man₅GlcNAc₂.

[0158] One of ordinary skill in the art can select host cells from nature, *e.g.*, existing fungi or other lower eukaryotes that produce significant levels of Man₅GlcNAc₂ *in vivo*. As yet, however, no lower eukaryote has been shown to 25 provide such structures *in vivo* in excess of 1.8% of the total *N*-glycans (*see e.g.* Maras *et al.* (1997) *Eur. J. Biochem.* 249:701-707). Alternatively, such host cells may be genetically engineered to produce the Man₅GlcNAc₂ structure *in vivo*. Methods such as those described in U.S. Patent No. 5,595,900 may be used to identify the absence or presence of particular glycosyltransferases, mannosidases 30 and sugar nucleotide transporters in a target host cell or organism of interest.

Inactivation of Undesirable Host Cell Glycosylation Enzymes

[0159] The methods of the invention are directed to making host cells which produce glycoproteins having altered, and preferably human-like, *N*-glycan structures. In a preferred embodiment, the methods are directed to making host 5 cells in which oligosaccharide precursors are enriched in $\text{Man}_5\text{GlcNAc}_2$. Preferably, a eukaryotic host cell is used that does not express one or more enzymes involved in the production of high mannose structures. Such a host cell may be found in nature or may be engineered, *e.g.*, starting with or derived from one of many such mutants already described in yeasts. Thus, depending on the 10 selected host cell, one or a number of genes that encode enzymes known to be characteristic of non-human glycosylation reactions will have to be deleted. Such genes and their corresponding proteins have been extensively characterized in a number of lower eukaryotes (*e.g.*, *S. cerevisiae*, *T. reesei*, *A. nidulans*, etc.), thereby providing a list of known glycosyltransferases in lower eukaryotes, their 15 activities and their respective genetic sequence. These genes are likely to be selected from the group of mannosyltransferases, *e.g.* 1,3 mannosyltransferases (*e.g.* *MNN1* in *S. cerevisiae*) (Graham, 1991), 1,2 mannosyltransferases (*e.g.* *KTR/KRE* family from *S. cerevisiae*), 1,6 mannosyltransferases (*OCH1* from *S. cerevisiae*), mannosylphosphate transferases and their regulators (*MNN4* and 20 *MNN6* from *S. cerevisiae*) and additional enzymes that are involved in aberrant, *i.e.*, non-human, glycosylation reactions. Many of these genes have in fact been deleted individually giving rise to viable phenotypes with altered glycosylation profiles. Examples are shown in Table 1.

[0160] Preferred lower eukaryotic host cells of the invention, as described herein 25 to exemplify the required manipulation steps, are hypermannosylation-minus (*och1*) mutants of *Pichia pastoris* or *K. lactis*. Like other lower eukaryotes, *P. pastoris* processes $\text{Man}_9\text{GlcNAc}_2$ structures in the ER with an α -1,2-mannosidase to yield $\text{Man}_8\text{GlcNAc}_2$ (Figure 1A). Through the action of several mannosyltransferases, this structure is then converted to hypermannosylated 30 structures ($\text{Man}_{10}\text{GlcNAc}_2$), also known as mannans (Figure 35A). In addition, it has been found that *P. pastoris* is able to add non-terminal phosphate groups, through the action of mannosylphosphate transferases, to the carbohydrate

structure. This differs from the reactions performed in mammalian cells, which involve the removal rather than addition of mannose sugars (Figure 35A). It is of particular importance to eliminate the ability of the eukaryotic host cell, *e.g.*, fungus, to hypermannosylate an existing $\text{Man}_8\text{GlcNAc}_2$ structure. This can be 5 achieved by either selecting for a host cell that does not hypermannosylate or by genetically engineering such a cell.

[0161] Genes that are involved in the hypermannosylation process have been identified, *e.g.*, in *Pichia pastoris*, and by creating mutations in these genes, one can reduce the production of “undesirable” glycoforms. Such genes can be 10 identified by homology to existing mannosyltransferases or their regulators (*e.g.*, *OCH1*, *MNN4*, *MNN6*, *MNN1*) found in other lower eukaryotes such as *C. albicans*, *Pichia angusta* or *S. cerevisiae* or by mutagenizing the host strain and selecting for a glycosylation phenotype with reduced mannosylation. Based on homologies amongst known mannosyltransferases and mannosylphosphate 15 transferases, one may either design PCR primers (examples of which are shown in Table 2), or use genes or gene fragments encoding such enzymes as probes to identify homologs in DNA libraries of the target or a related organism. Alternatively, one may identify a functional homolog having mannosyltransferase activity by its ability to complement particular glycosylation phenotypes in related 20 organisms.

Table 2. PCR Primers

PCR primer A	PCR primer B	Target Gene(s) in <i>P. pastoris</i>	Homologs
ATGGCGAAGGCA GATGGCAGT (SEQ ID NO:3)	TTAGTCCTTCAA CTTCCTTC (SEQ ID NO:4)	1,6-mannosyltransferase	<i>OCH1</i> <i>S. cerevisiae</i> , <i>Pichia albicans</i>
TAYTGGMNGNTN GARCYNGAYATH AA (SEQ ID NO:5)	GCRTCNCCCCANC KYTCRTA (SEQ ID NO:6)	1,2-mannosyltransferases	KTR/KRE family, <i>S. cerevisiae</i>

Legend: M = A or C, R = A or G, W = A or T, S = C or G, 25 Y = C or T, K = G or T, V = A or C or G, H = A or C or T, D = A or G or T, B = C or G or T, N = G or A or T or C.

[0162] To obtain the gene or genes encoding 1,6-mannosyltransferase activity in *P. pastoris*, for example, one would carry out the following steps: *OCH1* mutants

of *S. cerevisiae* are temperature sensitive and are slow growers at elevated temperatures. One can thus identify functional homologs of *OCH1* in *P. pastoris* by complementing an *OCH1* mutant of *S. cerevisiae* with a *P. pastoris* DNA or cDNA library. Mutants of *S. cerevisiae* are available, e.g., from Stanford University, and are commercially available from ResGen, Invitrogen Corp. (Carlsbad, CA). Mutants that display a normal growth phenotype at elevated temperature, after having been transformed with a *P. pastoris* DNA library, are likely to carry an *OCH1* homolog of *P. pastoris*. Such a library can be created by partially digesting chromosomal DNA of *P. pastoris* with a suitable restriction enzyme and, after inactivating the restriction enzyme, ligating the digested DNA into a suitable vector, which has been digested with a compatible restriction enzyme.

[0163] Suitable vectors include, e.g., pRS314, a low copy (CEN6/ARS4) plasmid based on pBluescript containing the Trp1 marker (Sikorski and Hieter (1989) *Genetics* 122:19-27) and pFL44S, a high copy (2 μ) plasmid based on a modified pUC19 containing the URA3 marker (Bonneaud *et al.* (1991) *Yeast* 7:609-615). Such vectors are commonly used by academic researchers and similar vectors are available from a number of different vendors (e.g., Invitrogen (Carlsbad, CA); Pharmacia (Piscataway, NJ); New England Biolabs (Beverly, MA)). Further examples include pYES/GS, 2 μ origin of replication based yeast expression plasmid from Invitrogen, or Yep24 cloning vehicle from New England Biolabs.

[0164] After ligation of the chromosomal DNA and the vector, one may transform the DNA library into a strain of *S. cerevisiae* with a specific mutation and select for the correction of the corresponding phenotype. After sub-cloning and sequencing the DNA fragment that is able to restore the wild-type phenotype, one may use this fragment to eliminate the activity of the gene product encoded by *OCH1* in *P. pastoris* using *in vivo* mutagenesis and/or recombination techniques well-known to those skilled in the art.

[0165] Alternatively, if the entire genomic sequence of a particular host cell, e.g., fungus, of interest is known, one may identify such genes simply by searching publicly available DNA databases, which are available from several sources, such as NCBI, Swissprot. For example, by searching a given genomic sequence or

database with sequences from a known 1,6 mannosyltransferase gene (*e.g.*, *OCH1* from *S. cerevisiae*), one can identify genes of high homology in such a host cell genome which may (but do not necessarily) encode proteins that have 1,6-mannosyltransferase activity. Nucleic acid sequence homology alone is not enough to prove, however, that one has identified and isolated a homolog encoding an enzyme having the same activity. To date, for example, no data exist to show that an *OCH1* deletion in *P. pastoris* eliminates the crucial initiating 1,6-mannosyltransferase activity (Martinet *et al.* (1998) *Biotech. Letters* 20(12):1171-1177; Contreras *et al.* WO 02/00856 A2). Thus, no data prove that the *P. pastoris* *OCH1* gene homolog actually encodes that function. That demonstration is provided for the first time herein.

[0166] Homologs to several *S. cerevisiae* mannosyltransferases have been identified in *P. pastoris* using these approaches. Homologous genes often have similar functions to genes involved in the mannosylation of proteins in *S. cerevisiae* and thus their deletion may be used to manipulate the glycosylation pattern in *P. pastoris* or, by analogy, in any other host cell, *e.g.*, fungus, plant, insect or animal cells, with similar glycosylation pathways.

[0167] The creation of gene knock-outs, once a given target gene sequence has been determined, is a well-established technique in the art and can be carried out by one of ordinary skill in the art (*see, e.g.*, Rothstein (1991) *Methods in Enzymology* 194:281). The choice of a host organism may be influenced by the availability of good transformation and gene disruption techniques.

[0168] If several mannosyltransferases are to be knocked out, the method developed by Alani and Kleckner (1987) *Genetics* 116:541-545, for example, enables the repeated use of a selectable marker, *e.g.*, the *URA3* marker in yeast, to sequentially eliminate all undesirable endogenous mannosyltransferase activity. This technique has been refined by others but basically involves the use of two repeated DNA sequences, flanking a counter selectable marker. For example: *URA3* may be used as a marker to ensure the selection of a transformants that have integrated a construct. By flanking the *URA3* marker with direct repeats one may first select for transformants that have integrated the construct and have thus disrupted the target gene. After isolation of the transformants, and their

characterization, one may counter select in a second round for those that are resistant to 5-fluoroorotic acid (5-FOA). Colonies that are able to survive on plates containing 5-FOA have lost the *URA3* marker again through a crossover event involving the repeats mentioned earlier. This approach thus allows for the repeated

5 use of the same marker and facilitates the disruption of multiple genes without requiring additional markers. Similar techniques for sequential elimination of genes adapted for use in another eukaryotic host cells with other selectable and counter-selectable markers may also be used.

[0169] Eliminating specific mannosyltransferases, such as 1,6
10 mannosyltransferase (*OCH1*) or mannosylphosphate transferases (*MNN6*, or genes complementing *lbd* mutants) or regulators (*MNN4*) in *P. pastoris* enables one to create engineered strains of this organism which synthesize primarily $\text{Man}_8\text{GlcNAc}_2$ and which can be used to further modify the glycosylation pattern to resemble more complex glycoform structures, e.g., those produced in mammalian,
15 e.g., human cells. A preferred embodiment of this method utilizes DNA sequences encoding biochemical glycosylation activities to eliminate similar or identical biochemical functions in *P. pastoris* to modify the glycosylation structure of glycoproteins produced in the genetically altered *P. pastoris* strain.

[0170] Methods used to engineer the glycosylation pathway in yeasts as
20 exemplified herein can be used in filamentous fungi to produce a preferred substrate for subsequent modification. Strategies for modifying glycosylation pathways in *A. niger* and other filamentous fungi, for example, can be developed using protocols analogous to those described herein for engineering strains to produce human-like glycoproteins in yeast. Undesired gene activities involved in
25 1,2 mannosyltransferase activity, e.g., *KTR/KRE* homologs, are modified or eliminated. A filamentous fungus, such as *Aspergillus*, is a preferred host because it lacks the 1,6 mannosyltransferase activity and as such, one would not expect a hypermannosylating gene activity, e.g. *OCH1*, in this host. By contrast, other desired activities (e.g., α -1,2-mannosidase, UDP-GlcNAc transporter,
30 glycosyltransferase (GnT), galactosyltransferase (GalT) and sialyltransferase (ST)) involved in glycosylation are introduced into the host using the targeting methods of the invention.

**Engineering or Selecting Hosts Having Diminished Initiating
α-1,6 Mannosyltransferase Activity**

[0171] In a preferred embodiment, the method of the invention involves making or using a host cell which is diminished or depleted in the activity of an initiating α-1,6-mannosyltransferase, *i.e.*, an initiation specific enzyme that initiates outer chain mannosylation on the α-1,3 arm of the Man₃GlcNAc₂ core structure. In *S. cerevisiae*, this enzyme is encoded by the *OCH1* gene. Disruption of the *OCH1* gene in *S. cerevisiae* results in a phenotype in which *N*-linked sugars completely lack the poly-mannose outer chain. Previous approaches for obtaining mammalian-type glycosylation in fungal strains have required inactivation of *OCH1* (*see, e.g.*, Chiba *et al.* (1998) *J. Biol. Chem.* 273:26298-304). Disruption of the initiating α-1,6-mannosyltransferase activity in a host cell of the invention may be optional, however (depending on the selected host cell), as the Och1p enzyme requires an intact Man₈GlcNAc₂ for efficient mannose outer chain initiation. Thus, host cells selected or produced according to this invention which accumulate oligosaccharides having seven or fewer mannose residues may produce hypoglycosylated *N*-glycans that will likely be poor substrates for Och1p (*see, e.g.*, Nakayama *et al.* (1997) *FEBS Lett.* 412(3):547-50).

[0172] The *OCH1* gene was cloned from *P. pastoris* (**Example 1**) and *K. lactis* (**Example 9**), as described. The nucleic acid and amino acid sequences of the *OCH1* gene from *K. lactis* are set forth in SEQ ID NOS:7 and 8. Using gene-specific primers, a construct was made from each clone to delete the *OCH1* gene from the genome of *P. pastoris* and *K. lactis* (**Examples 1 and 9**, respectively).

Host cells depleted in initiating α-1,6-mannosyltransferase activity and engineered to produce *N*-glycans having a Man₅GlcNAc₂ carbohydrate structure were thereby obtained (*see, e.g.*, **Figures 5, 6, and 12; Examples 4 and 9**).

[0173] Thus, in another embodiment, the invention provides an isolated nucleic acid molecule having a nucleic acid sequence comprising or consisting of at least forty-five, preferably at least 50, more preferably at least 60 and most preferably 75 or more nucleotide residues of the *K. lactis* *OCH1* gene (SEQ ID NO: 7), and homologs, variants and derivatives thereof. The invention also provides nucleic

acid molecules that hybridize under stringent conditions to the above-described nucleic acid molecules. Similarly, isolated polypeptides (including muteins, allelic variants, fragments, derivatives, and analogs) encoded by the nucleic acid molecules of the invention are provided. Also provided are vectors, including

5 expression vectors, which comprise the above nucleic acid molecules of the invention, as described further herein. Similarly, host cells transformed with the nucleic acid molecules or vectors of the invention are provided.

[0174] The invention further provides methods of making or using a non-human eukaryotic host cell diminished or depleted in an *alg* gene activity (*i.e.*, *alg* activities, including equivalent enzymatic activities in non-fungal host cells) and introducing into the host cell at least one glycosidase activity. In a preferred embodiment, the glycosidase activity is introduced by causing expression of one or more mannosidase activities within the host cell, for example, by activation of a mannosidase activity, or by expression from a nucleic acid molecule of a mannosidase activity, in the host cell.

[0175] In another embodiment, the method involves making or using a host cell diminished or depleted in the activity of one or more enzymes that transfer a sugar residue to the 1,6 arm of lipid-linked oligosaccharide precursors (Figure 13). A host cell of the invention is selected for or is engineered by introducing a mutation 20 in one or more of the genes encoding an enzyme that transfers a sugar residue (*e.g.*, mannosylates) the 1,6 arm of a lipid-linked oligosaccharide precursor. The sugar residue is more preferably mannose, is preferably a glucose, GlcNAc, galactose, sialic acid, fucose or GlcNAc phosphate residue. In a preferred embodiment, the activity of one or more enzymes that mannosylate the 1,6 arm of lipid-linked 25 oligosaccharide precursors is diminished or depleted. The method may further comprise the step of introducing into the host cell at least one glycosidase activity (see below).

[0176] In yet another embodiment, the invention provides a method for producing a human-like glycoprotein in a non-human host, wherein the 30 glycoprotein comprises an N-glycan having at least two GlcNAcs attached to a trimannose core structure.

[0177] In each above embodiment, the method is directed to making a host cell in which the lipid-linked oligosaccharide precursors are enriched in $\text{Man}_x\text{GlcNAc}_2$ structures, where X is 3, 4 or 5 (Figure 14). These structures are transferred in the ER of the host cell onto nascent polypeptide chains by an oligosaccharyl-

5 transferase and may then be processed by treatment with glycosidases (e.g., α -mannosidases) and glycosyltransferases (e.g., GnT1) to produce N-glycans having $\text{GlcNAcMan}_x\text{GlcNAc}_2$ core structures, wherein X is 3, 4 or 5, and is preferably 3 (Figures 14 and 15). As shown in Figure 14, N-glycans having a

10 $\text{GlcNAcMan}_x\text{GlcNAc}_2$ core structure where X is greater than 3 may be converted to $\text{GlcNAcMan}_3\text{GlcNAc}_2$, e.g., by treatment with an α -1,3 and/or α -1,2-1,3 mannosidase activity, where applicable.

[0178] Additional processing of $\text{GlcNAcMan}_3\text{GlcNAc}_2$ by treatment with glycosyltransferases (e.g., GnTII) produces $\text{GlcNAc}_2\text{Man}_3\text{GlcNAc}_2$ core structures which may then be modified, as desired, e.g., by *ex vivo* treatment or by

15 heterologous expression in the host cell of a set of glycosylation enzymes, including glycosyltransferases, sugar transporters and mannosidases (see below), to become human-like N-glycans. Preferred human-like glycoproteins which may be produced according to the invention include those which comprise N-glycans having seven or fewer, or three or fewer, mannose residues; comprise one or more sugars selected from the group consisting of galactose, GlcNAc, sialic acid, and fucose; and comprise at least one oligosaccharide branch comprising the structure NeuNAc-Gal-GlcNAc-Man.

20 [0179] In one embodiment, the host cell has diminished or depleted Dol-P-Man: $\text{Man}_5\text{GlcNAc}_2$ -PP-Dol Mannosyltransferase activity, which is an activity involved in the first mannosylation step from $\text{Man}_5\text{GlcNAc}_2$ -PP-Dol to

25 $\text{Man}_6\text{GlcNAc}_2$ -PP-Dol at the luminal side of the ER (e.g., *ALG3* Figure 13; Figure 14). In *S. cerevisiae*, this enzyme is encoded by the *ALG3* gene. As described above, *S. cerevisiae* cells harboring a leaky *alg3-1* mutation accumulate $\text{Man}_5\text{GlcNAc}_2$ -PP-Dol and cells having a deletion in *alg3* appear to transfer

30 $\text{Man}_5\text{GlcNAc}_2$ structures onto nascent polypeptide chains within the ER. Accordingly, in this embodiment, host cells will accumulate N-glycans enriched in $\text{Man}_5\text{GlcNAc}_2$ structures which can then be converted to $\text{GlcNAc}_2\text{Man}_3\text{GlcNAc}_2$

by treatment with glycosidases (e.g., with α -1,2 mannosidase, α -1,3 mannosidase, or α -1,2-1,3 mannosidase activities) and glycosyltransferase activites (e.g., GnT I, GnT II) (Figure 14; Figure 35B).

[0180] As described in Example 10, degenerate primers were designed based on
5 an alignment of Alg3 protein sequences from *S. cerevisiae*, *D. melanogaster* and
humans (*H. sapiens*) (Figures 16 and 17), and were used to amplify a product from
P. pastoris genomic DNA. The resulting PCR product was used as a probe to
identify and isolate a *P. pastoris* genomic clone comprising an open reading frame
(ORF) that encodes a protein having 35% overall sequence identity and 53%
10 sequence similarity to the *S. cerevisiae* *ALG3* gene (Figures 18 and 19). This *P.*
pastoris gene is referred to herein as “*PpALG3*”. The *ALG3* gene was similarly
identified and isolated from *K. lactis* (Example 10; Figures 20 and 21).

[0181] Thus, in another embodiment, the invention provides an isolated nucleic
acid molecule having a nucleic acid sequence comprising or consisting of at least
15 forty-five, preferably at least 50, more preferably at least 60 and most preferably
75 or more nucleotide residues of the *P. pastoris* *ALG3* gene (Figure 18) and the
K. lactis *ALG3* gene (Figure 20), and homologs, variants and derivatives thereof.
The invention also provides nucleic acid molecules that hybridize under stringent
conditions to the above-described nucleic acid molecules. Similarly, isolated
20 polypeptides (including muteins, allelic variants, fragments, derivatives, and
analog) encoded by the nucleic acid molecules of the invention are provided (*P.*
pastoris and *K. lactis* *ALG3* gene products are shown in Figures 18 and 20). In
addition, also provided are vectors, including expression vectors, which comprise a
nucleic acid molecule of the invention, as described further herein.

25 [0182] Using gene-specific primers, a construct was made to delete the *PpALG3*
gene from the genome of *P. pastoris* (Example 10). This strain was used to
generate a host cell depleted in Dol-P-Man:Man₅GlcNAc₂-PP-Dol
Mannosyltransferase activity and produce lipid-linked Man₅GlcNAc₂-PP-Dol
precursors which are transferred onto nascent polypeptide chains to produce *N*-
30 glycans having a Man₅GlcNAc₂ carbohydrate structure.

[0183] As described in Example 11, such a host cell may be engineered by
expression of appropriate mannosidases to produce *N*-glycans having the desired

Man₃GlcNAc₂ core carbohydrate structure. Expression of GnTs in the host cell (e.g., by targeting a nucleic acid molecule or a library of nucleic acid molecules as described below) enables the modified host cell to produce *N*-glycans having one or two GlcNAc structures attached to each arm of the Man₃ core structure (*i.e.*,

5 GlcNAc₁Man₃GlcNAc₂, GlcNAc₂Man₃GlcNAc₂, or GlcNAc₃Man₃GlcNAc₂; see Figure 15). These structures may be processed further using the methods of the invention to produce human-like *N*-glycans on proteins which enter the secretion pathway of the host cell.

[0184] In a preferred embodiment, the method of the invention involves making
10 or using a host cell which is both (a) diminished or depleted in the activity of an *alg* gene or in one or more activities that mannosylate *N*-glycans on the α -1,6 arm of the Man₃GlcNAc₂ ("Man3") core carbohydrate structure; and (b) diminished or depleted in the activity of an initiating α -1,6-mannosyltransferase, *i.e.*, an initiation specific enzyme that initiates outer chain mannosylation (on the α -1,3 arm of the
15 Man3 core structure). In *S. cerevisiae*, this enzyme is encoded by the *OCH1* gene. Disruption of the *och1* gene in *S. cerevisiae* results in a phenotype in which *N*-linked sugars completely lack the poly-mannose outer chain. Previous approaches for obtaining mammalian-type glycosylation in fungal strains have required inactivation of *OCH1* (see, e.g., Chiba *et al.* (1998) *J. Biol. Chem.* 273:26298-304).
20 Disruption of the initiating α -1,6-mannosyltransferase activity in a host cell of the invention is optional, however (depending on the selected host cell), as the Och1p enzyme requires an intact Man₈GlcNAc for efficient mannose outer chain initiation. Thus, the host cells selected or produced according to this invention, which accumulate lipid-linked oligosaccharides having seven or fewer mannose
25 residues will, after transfer, produce hypoglycosylated *N*-glycans that will likely be poor substrates for Och1p (see, e.g., Nakayama *et al.* (1997) *FEBS Lett.* 412(3):547-50).

30 **Engineering or Selecting Hosts Having *N*-Acetylglucosaminyltransferase III Activity**

[0185] The invention additionally provides a method for producing a human-like glycoprotein in a lower eukaryotic host cell by expressing an *N*-

acetylglucosaminyltransferase III activity (including a full-length enzyme, homologs, variants, derivatives, and catalytically active fragments thereof). In one embodiment, a host cell (*e.g.*, *P. pastoris*) is engineered to produce more human-like *N*-glycans, *e.g.*, by activation of an *N*-acetylglucosaminyltransferase III activity or by expression from a nucleic acid molecule of an *N*-acetylglucosaminyltransferase III activity. Using well-known techniques in the art, gene-specific primers are designed to complement the homologous regions of a GnTIII gene, preferably a mammalian GnTIII gene (*e.g.*, mouse GnTIII) (Figure 24), sequences for which are readily available in the art (*e.g.*, Genbank Accession No. L39373) and are PCR amplified.

[0186] In one embodiment, the invention provides a method for producing a human-like glycoprotein in a lower eukaryote (*e.g.*, *P. pastoris*), wherein the glycoprotein comprises an *N*-glycan exhibiting a bisecting GlcNAc on a trimannose or trimannosyl ($\text{Man}_3\text{GlcNAc}_2$) core structure. In this embodiment, GlcNAc $\text{Man}_3\text{GlcNAc}_2$ (which may be produced by reacting a trimannose core with *N*-acetylglucosaminyltransferase I (“GnTI”) activity, but which is typically produced by trimming of GlcNAc $\text{Man}_5\text{GlcNAc}_2$ by an α -1,3/ α -1,6-mannosidase activity, such as Mannosidase II (Hamilton *et al.* (2003) *Science* 301:1244-46)) is reacted with an *N*-acetylglucosaminyltransferase III activity to produce a bisected GlcNAc $\text{Man}_3\text{GlcNAc}_2$. Accordingly, the invention provides GnTIII activity, which transfers β -1,4 GlcNAc onto substrates that are capable of accepting the bisecting GlcNAc in lower eukaryotes.

[0187] In another embodiment, the invention provides a method for producing a human-like glycoprotein in a lower eukaryote (*e.g.*, *P. pastoris*), wherein the glycoprotein comprises an *N*-glycan exhibiting a bisecting GlcNAc on a trimannose or trimannosyl ($\text{Man}_3\text{GlcNAc}_2$) core structure having at least two GlcNAcs attached to the trimannose core. In this embodiment, Man GlcNAc_2 is reacted with a GnTI activity and then with an *N*-acetylglucosaminyltransferase II (“GnTII”) activity and a GnTIII activity (in either order) to produce a bisected GlcNAc $\text{Man}_3\text{GlcNAc}_2$ (Figure 38). It should be appreciated that the bisected trimannosyl core structure of this embodiment may also contain an additional mannosyl group in place of a GlcNAc residue. For example,

GlcNAcMan₄GlcNAc₂ may be reacted with a GnTIII activity to produce a bisected GlcNAc₂Man₄GlcNAc₂.

[0188] The invention also provides a method for producing a more human-like glycoprotein in a lower eukaryote (*e.g.* *P. pastoris*), wherein the glycoprotein produced comprises an *N*-glycan having at least two GlcNAcs attached to a pentamannose core structure (Man₅GlcNAc₂) and which exhibits a bisected *N*-glycan. Accordingly, in this embodiment, a pentamannose core structure (Man₅GlcNAc₂) is reacted with GnTIII activity to produce a bisected GlcNAcMan₅GlcNAc₂ and GlcNAc₂Man₅GlcNAc₂ structure.

10 [0189] In an alternative embodiment, a pentamannose core structure produced via the mutation of *och1* and *alg3* genes is reacted with α 1,2-mannosidase, GnTI, GnTII and GnTIII activities and UDP-GlcNAc to produce a bisected GlcNAc₃Man₃GlcNAc₂ glycan (Figure 35B). In another embodiment, a pentamannose core structure is reacted with GnTI and GnTIII activities (in either order or in combination) to produce a bisected GlcNAc₂Man₅GlcNAc₂ structure (Figure 37).

15 [0190] In a more preferred embodiment, using the combinatorial DNA library method of the invention, as described below, a pVA53 construct comprising the *S. cerevisiae* *MNN2(s)* leader (GenBank Accession No. NP_009571) fused to a catalytically active GnTIII domain from mouse (GnTIII Δ32) is expressed in a *P. pastoris* strain YSH-1 (Example 13) thereby producing *N*-glycans having a bisected GlcNAc₂Man₅GlcNAc₂ structure (Example 20). Figure 26 (bottom) displays the MALDI-TOF spectrum of *N*-glycans released from a kringle 3 protein expressed in the above-mentioned strain, which is designated PBP26 (Figure 36), exhibiting a predominant peak at 1666 m/z [a], which corresponds to bisected GlcNAc₂Man₅GlcNAc₂. (For comparison, Figure 26 (top) displays the MALDI-TOF spectrum of *N*-glycans released from a kringle 3 protein expressed in strain YSH-1 lacking the pVA53 construct. The predominant peak at 1461 m/z [d] corresponds to the unmodified glycan: GlcNAcMan₅GlcNAc₂.) Accordingly, in one embodiment, a host of the present invention is characterized by its ability to produce, at least transiently, *N*-glycans which exhibit at least 50 mole % of a GlcNAc₂Man₅GlcNAc₂ or at least 50 mole % of a GlcNAc₂Man₃GlcNAc₂.

structure having a bisecting GlcNAc. The mole percent of the glycans is in reference to percent of total neutral glycans as detected by MALDI-TOF. It is understood that if, for example, GlcNAc₂Man₃GlcNAc₂ having a bisecting GlcNAc is produced at 20% and GlcNAc₃Man₃GlcNAc₂ is produced at 25% on a target 5 protein, the total amount of transiently produced GlcNAc₂Man₃GlcNAc₂ having a bisecting GlcNAc is 45%, because GlcNAc₃Man₃GlcNAc₂ is a product of a GlcNAc₂Man₃GlcNAc₂ having a bisecting GlcNAc further reacted with GnTII.

[0191] Similarly, in another embodiment, a pVA55 construct comprising the *S. cerevisiae MNN2(l)* leader (GenBank Accession No. NP_009571) fused to a 10 catalytically active GnTIII domain from mouse (GnTIII Δ32) is expressed in a *P. pastoris* strain (YSH-1) thereby producing *N*-glycans GlcNAcMan₅GlcNAc₂ and bisected *N*-glycans GlcNAc₂Man₅GlcNAc₂ structure. As shown in Figure 27 (bottom), these structures correspond to peaks at 1463 m/z and 1667 m/z, respectively. (For comparison, Figure 27 (top) displays the MALDI-TOF 15 spectrum of *N*-glycans released from a kringle 3 protein expressed in strain YSH-1 lacking the pVA53 construct. The predominant peak corresponds to unmodified GlcNAcMan₅GlcNAc₂ at 1461 m/z [d].) Accordingly, in another embodiment, a host of the present invention is characterized by its ability to produce, at least transiently, *N*-glycans which exhibit at least 20 mole % of a 20 GlcNAc₂Man₅GlcNAc₂ or at least 20 mole % of a GlcNAc₂Man₃GlcNAc₂ structure having a bisecting GlcNAc.

[0192] In an even more preferred embodiment, a pVA53 construct comprising the *S. cerevisiae MNN2(s)* leader (GenBank Accession No. NP_009571) fused to a catalytically active GnTIII domain from mouse (GnTIII Δ32) is expressed in a *P. pastoris* strain YSH-44 (Example 15) thereby producing *N*-glycans having a 25 bisected GlcNAc₃Man₃GlcNAc₂ structure (Example 20). Figure 30 displays the MALDI-TOF spectrum of *N*-glycans released from a kringle 3 protein expressed in the above-mentioned strain designated as YSH-57, exhibiting a predominant peak at 1542 m/z [y], which corresponds to the bisected glycan GlcNAc₃Man₃GlcNAc₂. 30 (For comparison, Figure 29 displays the MALDI-TOF spectrum of *N*-glycans released from a kringle 3 protein expressed in strain YSH-44 lacking the pVA53 construct. The predominant peak at 1356 m/z [x] in Figure 29 corresponds to the

unmodified glycan: GlcNAc₂Man₃GlcNAc₂.) Accordingly, in one embodiment, a host of the present invention is characterized by its ability to produce, at least transiently, *N*-glycans which exhibit at least 80 mole % of a

5 GlcNAc₃Man₃GlcNAc₂ structure having a bisecting GlcNAc. The mole percent of the glycans is in reference to percent of total neutral glycans as detected by MALDI-TOF.

[0193] Alternatively, in another embodiment, a pVA53 construct comprising the *S. cerevisiae MNN2(s)* leader (GenBank Accession No. NP_009571) fused to a catalytically active GnTIII domain from mouse (GnTIII Δ32) is expressed in a *P. pastoris* strain (PBP6-5) (Example 11) thereby producing *N*-glycans having a GlcNAc₂Man₃GlcNAc₂ and a bisected GlcNAc₃Man₃GlcNAc₂ structure. As shown in Figure 32, these structures correspond to peaks at 1340 m/z and 1543 m/z, respectively. Accordingly, in another embodiment, a host of the present invention is characterized by its ability to produce, at least transiently, *N*-glycans which exhibit at least 20 mole % of a GlcNAc₃Man₃GlcNAc₂ structure having a bisecting GlcNAc in an *alg3* mutant host cell.

[0194] The invention provides methods for producing a human-like glycoprotein in a lower eukaryote, wherein the glycoprotein comprises a Man₅GlcNAc₂ core structure or a Man₃GlcNAc₂ core structure, and wherein the core structure is further modified by two or more GlcNAcs. In some embodiments of the invention, 10% or more of the core structures are modified by the two or more GlcNAcs. In other preferred embodiments, 20%, 30%, 40%, 50%, 60%, 70%, 80% or even more of the core structures are so modified. In a highly preferred embodiment, one of the GlcNAcs is a bisecting GlcNAc.

25 [0195] In another aspect of the invention, a combinatorial nucleic acid library which encodes at least one GnTIII catalytic domain is used to express a GnTIII activity in a lower eukaryotic host cell (Example 18). Preferably, a library of the invention comprises a sublibrary of leader sequences fused in frame to a single nucleic acid molecule or a sublibrary of nucleic acid molecules comprising GnTIII 30 sequences, one or more of which encode a catalytic domain having GnTIII activity in the host cell. Alternatively, a single nucleic acid molecule or a sublibrary of nucleic acid molecules comprising leader sequences is fused in frame to a

sublibrary of nucleic acid molecules comprising GnTIII sequences, one or more of which encode a catalytic domain having GnTIII activity in the host cell. (See below.) Expression of these and other such combinatorial libraries is performed in a host cell which expresses a target glycoprotein whose *N*-glycan structures are
5 analyzed to determine whether and how much GnTIII is expressed. A wide range of catalytically active GnTIII enzymes may be produced in a host cell using the methods and libraries of the invention. It is this aspect of the invention that allows a skilled artisan to create and delineate between GnTIII enzymes having little or no activity and those enzymes that are actively expressed and which produce
10 predominant levels of a desired bisected oligosaccharide intermediate such as GlcNAc₂Man₅GlcNAc₂, GlcNAc₃Man₃GlcNAc₂ or GlcNAc₂Man₃GlcNAc₂ in the host cells.

[0196] As described further below, the proper targeting of an enzyme responsible for a given step in the glycosylation pathway to the appropriate subcellular location
15 and the sufficiency of the enzyme's activity at the particular pH of that subcellular location are important factors in the production of glycoproteins having *N*-glycans with the desired structures. The use of combinatorial libraries of fusion proteins to generate diverse populations of enzyme chimeras and the screening of these libraries in transformed cells provides a powerful method to identify host strains
20 with the activity of interest in the appropriate location. In preferred embodiments of the invention, the enzyme activity is located such that an *N*-glycan-containing glycoprotein expressed in the cell is capable of reacting with the activity during the secretion process.

[0197] Not all combinations of leader/catalytic domains produce desired enzyme
25 activities however. A wide variety of leader/catalytic domain combinations is created, only a few of which may be useful in producing the presently desired intermediates. The present invention, nevertheless, encompasses even those combinations that do not presently exhibit a desired enzymatic activity in the exemplified host cell. Figure 28 (bottom) shows a pVB51 construct comprising
30 the *K. lactis* GNT(s) leader (GenBank Accession No. AF106080) fused to a catalytically active GnTIII domain from mouse (GnTIII Δ32) expressed in a *P. pastoris* strain YSH-1, which does not readily exhibit GnTIII activity. (For

comparison, **Figure 28** (top) displays the MALDI-TOF spectrum of *N*-glycans released from a kringle 3 protein expressed in strain YSH-1 lacking the pVA53 construct. The predominant peak corresponds to unmodified GlcNAcMan₅GlcNAc₂ at 1461 m/z.) The predominant peak in **Figure 28** (bottom) at 1463 m/z, which correlates to the mass of GlcNAcMan₅GlcNAc₂, is observed. A second peak at 1726 m/z, which does not correlate to the mass of GlcNAc₂Man₅GlcNAc₂ is also observed. It is contemplated that these and other such combinations may be useful, with or without slight modifications using techniques well known in the art, when they are expressed, e.g., in other host cells including those which have been modified to produce human-like glycoforms.

10 [0198] The use of combinatorial libraries to generate diverse populations of enzyme chimeras and the screening of these libraries in transformed cells further allows strains to be identified in which the enzyme activity is substantially intracellular. **Example 6**, below, provides an example of assay conditions useful for measuring extracellular α -1,2-mannosidase activity. **Examples 22 and 23** also provide examples of assays for glycosyltransferase activity (GnTIII) in the medium. See also **Table 9**, below, and Choi *et al.* (2003) *Proc. Natl. Acad. Sci. U.S.A.* 100(9):5022-27. For purposes of the invention, an enzyme activity is substantially intracellular when less than 10% of the enzyme activity is measurable in the extracellular medium.

15 [0199] As described in **Examples 11, 12, 13, 14, 15, and 19-21**, a host cell may be engineered by the expression of appropriate glycosyltransferases (e.g., *N*-acetylglucosaminyltransferase) to produce *N*-glycans having the desired carbohydrate structures (e.g., GlcNAc₂Man₃GlcNAc₂, GlcNAc₃Man₃GlcNAc₂).

20 Expression of GnTs in the host cell (e.g., by targeting a nucleic acid molecule or a library of nucleic acid molecules as described below and in Choi *et al.* (2003) *Proc. Natl. Acad. Sci. U.S.A.* 100(9):5022-27 and WO 02/00879) enables the modified host cell to produce *N*-glycans having the bisecting GlcNAc on the middle mannose. These structures may be processed further using the methods of the invention to produce human-like *N*-glycans on proteins which enter the secretion pathway of the host cell.

[0200] In a more preferred embodiment, co-expression of appropriate UDP-sugar-transporter(s) and -transferase(s) will cap the terminal α -1,6 and α -1,3 residues as well as the middle mannose with GlcNAc, resulting in the precursor for mammalian-type complex (*e.g.* GlcNAc₃Man₃GlcNAc₂) and hybrid N-glycosylation. These peptide-bound *N*-linked oligosaccharide chains then serve as a precursor for further modification to a mammalian-type oligosaccharide structure. Subsequent expression of galactosyl-transferases and genetically engineering the capacity to transfer sialic acid to the termini (see **Figure 1B**) will produce a mammalian-type (*e.g.*, human-like) *N*-glycan structure.

10

Engineering or Selecting Hosts Having *N*-Acetylglucosaminyltransferase IV, V, VI or IX Activity

[0201] The present invention provides novel lower eukaryotic hosts having *N*-acetylglucosaminyltransferase activity that catalyze the formation of a GlcNAc β 1,4 or a GlcNAc β 1,6 glycosidic linkage on the Man α 1,6 arm and/or Man α 1,3 arm of an oligosaccharide substrate (*e.g.*, GlcNAc₂Man₃GlcNAc₂) in the presence of a sugar nucleotide UDP-GlcNAc. Transfer of the GlcNAc residues is generally preferred in the presence of a UDP-GlcNAc transporter. The present invention provides recombinant nucleic acid molecules encoding proteins having *N*-acetylglucosaminyltransferase activity and methods for expressing active enzyme in the yeast secretory pathway. In addition, the present invention provides oligosaccharide structures produced from the transformed hosts that are useful for therapeutic administration. By catalyzing the transfer of the sugar GlcNAc from UDP-GlcNAc onto the oligosaccharide substrates by an *N*-acetylglucosaminyl-transferase activity, multiantennary glycoforms are formed on a protein, which are then extended by galactosyltransferase and sialyltransferases.

[0202] The invention additionally provides a method for producing a human-like glycoprotein in a lower eukaryotic host cell by expressing *N*-acetylglucosaminyltransferase IV, V, VI or IX activities (including a full-length enzyme, homologs, variants, derivatives, and catalytically active fragments thereof). In one embodiment, a host cell (*e.g.*, *P. pastoris*) is engineered to produce more human-like *N*-glycans, *e.g.*, by activation of an *N*-

acetylglucosaminyltransferase IV, V, VI, IX activities or by expression from a nucleic acid molecule encoding *N*-acetylglucosaminyltransferase IV, V, VI, IX activities (Figure 39). Using well-known techniques in the art, gene-specific primers are designed to hybridize to homologous regions of members of glycosyltransferase family, such as GnTIV, V, VI, IX gene sequences, which are readily available in the art (e.g., Genbank, SwissProt databases) and are PCR amplified.

Expression of *N*-Acetylglucosaminyltransferase IV

[0203] In a first aspect of the invention, a lower eukaryotic host cell is transformed with a nucleotide sequence encoding for the enzyme *N*-Acetylglucosaminyltransferase IV (“GnTIV”) that catalyzes the addition of a sugar residue $\beta(1,4)$ *N*-acetylglucosamine (“GlcNAc”) on the Man α 1,3 arm of the GlcNAc β 1,2 - Man α 1,6 (GlcNAc β 1,2 Man α 1,3) Man β 1,4 - GlcNAc β 1,4 - GlcNAc β 1,4 - Asn of an oligosaccharide substrate. The addition of a GlcNAc β 1,4 by GnTIV onto the Man α 1,3 arm of the acceptor substrate (e.g., GlcNAc₂Man₃GlcNAc₂) yields a so-called triantennary *N*-glycan.

[0204] In one embodiment, the invention provides a method for producing a human-like glycoprotein in a lower eukaryote (e.g., *P. pastoris*), wherein the glycoprotein comprises a triantennary *N*-glycan structure on an oligosaccharide structure (e.g., GlcNAc₃Man₃GlcNAc₂). In this embodiment, the oligosaccharide substrate GlcNAcMan₅GlcNAc₂ (Choi *et al.*, (2003) *Proc Natl Acad Sci U S A* 2003 Apr 29;100(9):5022-7) is trimmed by an α -1,3/ α -1,6-mannosidase activity, such as Mannosidase II (Hamilton *et al.* (2003) *Science* 301:1244 producing the substrate GlcNAc₂Man₃GlcNAc₂, which in turn is reacted with an *N*-acetylglucosaminyltransferase IV activity to produce a triantennary structure: GlcNAc₃Man₃GlcNAc₂ (Figure 39). Two GnTIV isozymes A and B are set forth in Figures 41 and 42. In a preferred embodiment, a gene fragment encoding human GnTIV (Figure 42) ligated in-frame to nucleotides 1-108 of the *S.cerevisiae* *MNN2(s)* targeting peptide sequence is introduced and expressed in *P.pastoris* YSH-44 (Example 15). Thus, in certain embodiments, a host cell of the present invention is characterized by its ability to produce, at least transiently, *N*-glycans which exhibit preferably at least 50, 60, 70, 80, 90 mole % or more of the

desired *N*-glycan structure GlcNAc₃Man₃GlcNAc₂. In an even more preferred embodiment, the oligosaccharide substrate GlcNAc₃Man₃GlcNAc₂ is a substrate for a galactosyltransfer reaction. Accordingly, the invention provides a GnTIV activity, which catalyzes the transfer of GlcNAc residues onto oligosaccharide substrates forming a GlcNAc β -1,4 glycosidic linkage on the Man α 1,3 arm of an oligosaccharide substrate (e.g., GlcNAc₂Man₃GlcNAc₂) in lower eukaryotes.

[0205] In a more preferred embodiment, using a combinatorial DNA library and methods of the invention, pPB144 construct (Figure 40A) comprising the *S. cerevisiae MNN2(s)* leader (GenBank Accession No. NP_009571) fused to a catalytically active GnTIV domain from human (GnTIVB Δ 104) is expressed in a *P. pastoris* strain YSH-44 (Example 15) thereby producing triantennary *N*-glycan GlcNAc₃Man₃GlcNAc₂ structures (Example 25). Figure 47 displays the MALDI-TOF spectrum of *N*-glycans released from a kringle 3 protein expressed in transformed strain designated as PBP43, exhibiting a predominant peak at 1543 m/z [y], which corresponds to triantennary *N*-glycan structure GlcNAc₃Man₃GlcNAc₂. (See Figure 29 for comparison, which displays the MALDI-TOF spectrum of *N*-glycans released from a kringle 3 protein expressed in YSH-44 cells producing GlcNAc₂Man₃GlcNAc₂.) Accordingly, in certain embodiments, a host cell of the present invention is characterized by its ability to produce, at least transiently, *N*-glycans which exhibit preferably at least 50 mole % of a GlcNAc₃Man₃GlcNAc₂ triantennary structure. In another embodiment, the host cell of the present invention produces the triantennary structure GlcNAc₃Man₃GlcNAc₂ catalyzed by GnTIV in an amount that is at least 60, 70, 80, more preferably 90 mole % or greater. Here and throughout, the mole percent of the glycans is in reference to percent of total neutral glycans as detected by MALDI-TOF MS.

Expression of *N*-Acetylglucosaminyltransferase IV and V

[0206] In another aspect of the invention, a lower eukaryotic host cell that is engineered or selected to produce triantennary oligosaccharide substrate (e.g., GlcNAc₃Man₃GlcNAc₂), transformed with a nucleic acid encoding for *N*-acetylglucosaminyltransferase V ("GnTV") activity which catalyzes the addition of

a sugar residue $\beta(1,6)$ N-acetylglucosamine ("GlcNAc") on the Man α 1,6 arm of the GlcNAc β 1,2 - Man α 1,6 (GlcNAc β 1,4 (GlcNAc β 1,2) Man α 1,3) Man β 1,4 - GlcNAc β 1,4 - GlcNAc β 1,4 - Asn of an oligosaccharide substrate. The addition of a GlcNAc β 1,6 by GnTV onto the acceptor substrate (e.g. GlcNAc₃Man₃GlcNAc₂)

5 in the presence of GlcNAc residues yields a tetraantennary N-glycan:
GlcNAc₄Man₃GlcNAc₂. Preferably, the GnTV activity of the present invention is expressed in a lower eukaryotic host cell producing triantennary glycans, for example in *P.pastoris* PBP43 in which the GnTV activity catalyzes the transfer of a GlcNAc residue onto the Man α 1,6 arm of the oligosaccharide substrate

10 GlcNAc₃Man₃GlcNAc₂ forming a GlcNAc β 1,6 glycosidic linkage..

[0207] In one embodiment, using the combinatorial DNA library method of the invention, *P.pastoris* YSH-44 is transformed with the GnTIVB/*S.cerevisiae* MNN2(s) and GnTV/ *S.cerevisiae* MNN2(s) fusion constructs. Figure 49 displays the MALDI-TOF spectrum of N-glycans released from a kringle 3 protein

15 expressed in transformed strain designated as PBP46, exhibiting a predominant peak at 1747 m/z [z], which corresponds to tetraantennary N-glycan structure GlcNAc₄Man₃GlcNAc₂ and a residual peak at 1543 m/z [y], which corresponds to triantennary N-glycan structure GlcNAc₃Man₃GlcNAc₂.

[0208] In another embodiment, using the combinatorial DNA library method of the invention, *P.pastoris* YSH-44 is transformed with a different combination of enzyme and leader fusion: plasmid pPB128 containing GnTIVA(Δ 82)/*S.cerevisiae* MNN2(s) and plasmid pPB140 containing GnTV(Δ 145)/ *S.cerevisiae* MNN2(s) fusion constructs. Figure 50 displays the MALDI-TOF spectrum of N-glycans released from a kringle 3 protein expressed in transformed strain designated as

20 PBP94, exhibiting a predominant peak at 1743 m/z [z], which corresponds to tetraantennary N-glycan structure GlcNAc₄Man₃GlcNAc₂.

[0209] Accordingly, in certain embodiments, the present invention provides a host cell characterized by its ability to produce, at least transiently, N-glycans which exhibit at least 50, 60, 70, 80, 90 mole % or more of the desired N-glycan

25 structure GlcNAc₄Man₃GlcNAc₂. In a more preferred embodiment, by expressing the GnTIV and GnTV activities, the host cell produces the desired multiple antennary N-glycan structure.

[0210] Not all combinations of leader/catalytic domains produce desired enzyme activities in the production of multiantennary glycan structures. A wide variety of leader/catalytic domain combinations is created using the methods and libraries of the invention, only a few of which may be useful in producing the presently desired intermediates. The present invention, nevertheless, encompasses even those combinations that do not presently exhibit a desired enzymatic activity in an exemplified host cell. It is contemplated that these and other such combinations may be useful, with or without slight modifications using techniques well known in the art, when they are expressed, e.g., in other host cells including those which have been modified to produce human-like glycoforms.

Expression of N-Acetylglucosaminyltransferase V

[0211] In another embodiment of the invention, a nucleic acid encoding GnTV activity is expressed in host cells producing the core GlcNAc₂Man₃GlcNAc₂ structures resulting in the formation of triantennary structures GlcNAc₃Man₃GlcNAc₂. Known GnTV sequence is provided in Figure 43. In one embodiment, using the combinatorial DNA library method of the invention, pPB140 construct (Figure 40B) comprising the *S. cerevisiae* MNN2(s) leader (GenBank Accession No. NP_009571) fused to a catalytically active GnTV domain from mouse (GnTV Δ145) is expressed in a *P. pastoris* strain YSH-44 (Example 15) thereby producing triantennary *N*-glycan GlcNAc₃Man₃GlcNAc₂ structures (Example 25). Figure 48 displays the MALDI-TOF spectrum of *N*-glycans released from a kringle 3 protein expressed in transformed strain designated as PBP32, exhibiting a predominant peak at 1559 m/z [y], which corresponds to tetraantennary *N*-glycan structure, and a second peak at 1355 m/z [u] which corresponds to triantennary *N*-glycan structure GlcNAc₃Man₃GlcNAc₂. (See Figure 29 for comparison, which displays the MALDI-TOF spectrum of *N*-glycans released from a kringle 3 protein expressed in YSH-44 cells producing GlcNAc₂Man₃GlcNAc₂.) Accordingly, in certain embodiments, a host of the present invention is characterized by its ability to produce, at least transiently, *N*-glycans which exhibit at least 40 mole % of the triantennary structure: GlcNAc₃Man₃GlcNAc₂. Preferably, the host of the present invention produces at

least 50, 60, 70, 80, 90 mole % or greater triantennary structure catalyzed by GnTV.

Expression of N-Acetylglucosaminyltransferase VI

5 [0212] The invention also provides a lower eukaryotic host cell transformed with a nucleic acid encoding *N*-acetylglucosaminyltransferase VI (“GnVT”) activity, which catalyzes the transfer of a sugar residue β (1,4) *N*-acetylglucosamine on the Man α 1,6 arm of the GlcNAc β 1,4 (GlcNAc β 1,2) Man α 1,6 (GlcNAc β 1,4 (GlcNAc β 1,2) Man α 1,3) Man β 1,4 - GlcNAc β 1,4 - GlcNAc β 1,4 - Asn of an oligosaccharide substrate. The addition of a GlcNAc β 1,4 by GnTVI to an acceptor substrate (e.g. GlcNAc₄Man₃GlcNAc₂) yields a so-called pentaantennary *N*-glycan. The addition of the GlcNAc residue forms a GlcNAc β 1,4 glycosidic linkage on the oligosaccharide substrate. In one embodiment, a nucleic acid encoding GnTVI activity is expressed in a host cell producing pentaantennary *N*-glycans such as

10 GlcNAc₅Man₃GlcNAc₂. In another embodiment, using a DNA fragment encoding the GnTVI activity such as one set forth in **Figure 44**, a plasmid construct comprising the *S. cerevisiae* *MNN2*(s) leader (GenBank Accession No. NP_009571) fused to a catalytically active GnTVI domain from *Gallus gallus* is expressed in a *P. pastoris* strain YSH-44 thereby producing pentaantennary *N*-glycan GlcNAc₅Man₃GlcNAc₂ structures. Accordingly, a host cell of the invention is characterized by its ability to produce, at least transiently, *N*-glycans which produce pentaantennary *N*-glycans in a detectable moiety.

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Expression of N-Acetylglucosaminyltransferase IX

25 [0213] In another embodiment, a nucleic acid encoding GnTIX activity (e.g., Genbank AN NP_945193) is expressed in a host cell in which the GnTIX activity catalyzes the transfer of GlcNAc residues onto complex glycan acceptor substrates (e.g., GlcNAc₂Man₃GlcNAc₂) in the absence of GnTIV, GnTV or GnTVI. The nucleic acid encoding GnTIX activity, which normally appears to be expressed exclusively in the brain, has been shown to catalyze the synthesis of a unique *N*-linked oligosaccharide in CHO mutant cells (Raju et al., (1998) *J Biol Chem* 273, 14090-14098). Expression of a recombinant human GnTIX showed GnTV

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activity, catalyzing the transfer of GlcNAc to the 6-OH position of α 1,6 linked mannose arm of the oligosaccharide GlcNAc₂Man₃GlcNAc₂-PA via β 1,6 linkage in addition to acting on the α 1,3-linked mannose arm (*J Biol Chem.* 2003 Oct 31;278(44):43102-9). The GnTIX is able to catalyze the transfer of GlcNAc to the 5 6-OH position of mannose in the sequence GlcNAc β 1,2-Man α 1.

[0214] Accordingly, the present invention provides a method of producing tetraantennary glycan structures in *P.pastoris* using a nucleic acid encoding GnTIX activity (Figure 45). Preferably, the introduction and expression of GnTIX activity catalyzes the transfer of GlcNAc residues onto the acceptor substrate

10 GlcNAc₂Man₃GlcNAc₂ producing GlcNAc₄Man₃GlcNAc₂ and GlcNAc₃Man₃GlcNAc₂. The host cell expression of GnTIX activity catalyzes the transfer of GlcNAc residues to the 6-OH position preferably on both Man α 1,3 and Man α 1,6 arms of the core GlcNAc₂Man₃GlcNAc₂ oligosaccharide substrate producing the tetraantennary structure GlcNAc₄Man₃GlcNAc₂. In one

15 embodiment, a host cell producing the acceptor substrate GlcNAc₂Man₃GlcNAc₂, such as *P.pastoris* YSH-44, is transformed with a plasmid (such as pPB176) comprising a gene encoding the GnTIX activity (Δ 43) fused in frame to the *S.cerevisiae* MNN2(s) leader (Example 29). The host cell is characterized by its ability to produce, at least transiently, *N*-glycans which exhibit at least 5 mole % or 20 preferably greater GnTIX activity catalyzing the transfer of GlcNAc to the 6-OH position of α 1,6 linked mannose arm of the oligosaccharide GlcNAc₂Man₃GlcNAc₂ via β 1,6 linkage in addition to acting on the α 1,3-linked mannose arm of the oligosaccharide substrate.

[0215] Additionally, the nucleic acid encoding GnTIX activity may be codon 25 optimized for translational efficiency in yeast using well known procedures. Figures 46 provides the codon optimized DNA fragment encoding part of the human GnTIX lacking the TM domain (Δ 43) synthesized from oligonucleotides using PCR.

30 Variations of Multiple Antennary Glycans

[0216] The present invention is adapted to produce a variety of multiple antennary glycans which are suitable for therapeutic purposes. It is contemplated

that glycans having the same GlcNAc β -linkages may increase half-life of the proteins. Accordingly, the invention provides a lower eukaryotic host cell comprising *N*-glycans having at least two GlcNAc residues on either the Man α 1,3 or Man α 1,6 arm of the trimannose core oligosaccharide intermediate (e.g., 5 Man₃GlcNAc₂). In one embodiment, the lower eukaryotic host cell comprises at least two GlcNAc β 1,4 residues on the Man α 1,3 and Man α 1,6 arm of the trimannose core oligosaccharide intermediate (e.g., Man₃GlcNAc₂). In another embodiment, the lower eukaryotic host cell comprises at least two GlcNAc β 1,6 residues on the Man α 1,3 and Man α 1,6 arm of the trimannose core oligosaccharide 10 intermediate (e.g., Man₃GlcNAc₂). In yet another embodiment, the lower eukaryotic host cell comprises at least two GlcNAc β 1,2 residues on the Man α 1,3 and Man α 1,6 arm of the trimannose core oligosaccharide intermediate (e.g., Man₃GlcNAc₂).

[0217] As noted below, while lower eukaryotic host cells are preferred hosts for 15 producing therapeutic proteins using the methods of the invention, the present invention is also useful for modifying *N*-glycan profiles of glycoproteins made in any eukaryotic host cell, preferably in a non-human (e.g., mammalian) host cell.

Host Cells of the Invention

20 [0218] A preferred host cell of the invention is a lower eukaryotic cell, *e.g.*, yeast, a unicellular and multicellular or filamentous fungus. However, a wide variety of host cells are envisioned as being useful in the methods of the invention. Plant cells or insect cells, for instance, may be engineered to express a human-like glycoprotein according to the invention. Likewise, a variety of non-human, 25 mammalian host cells may be altered to express more human-like or otherwise altered glycoproteins using the methods of the invention. As one of skill in the art will appreciate, any eukaryotic host cell (including a human cell) may be used in conjunction with a library of the invention to express one or more chimeric proteins which is targeted to a subcellular location, *e.g.*, organelle, in the host cell 30 where the activity of the protein is modified, and preferably is enhanced. Such a protein is preferably -- but need not necessarily be -- an enzyme involved in protein glycosylation, as exemplified herein. It is envisioned that any protein

coding sequence may be targeted and selected for modified activity in a eukaryotic host cell using the methods described herein.

[0219] Lower eukaryotes that are able to produce glycoproteins having the attached *N*-glycan $\text{Man}_5\text{GlcNAc}_2$ are particularly useful because (a) lacking a high degree of mannosylation (*e.g.*, greater than 8 mannoses per *N*-glycan, or especially 30-40 mannoses), they show reduced immunogenicity in humans; and (b) the *N*-glycan is a substrate for further glycosylation reactions to form an even more human-like glycoform, *e.g.*, by the action of GlcNAc transferase I (Figure 1B; β 1,2 GnTI) to form $\text{GlcNAcMan}_5\text{GlcNAc}_2$. A yield is obtained of greater than 30 mole %, more preferably a yield of 50, 60, 70, 80, 90, or even 100 mole %, glycoproteins with *N*-glycans having a $\text{Man}_5\text{GlcNAc}_2$ structure. In a preferred embodiment, more than 50% of the $\text{Man}_5\text{GlcNAc}_2$ structure is shown to be a substrate for a GnTI activity and can serve as such a substrate *in vivo*.

[0220] Preferred lower eukaryotes of the invention include but are not limited to:

15 *Pichia pastoris*, *Pichia finlandica*, *Pichia trehalophila*, *Pichia koclamae*, *Pichia membranaefaciens*, *Pichia minutia* (*e.g.*, *Ogataea minuta*, *Pichia lindneri*), *Pichia opuntiae*, *Pichia thermotolerans*, *Pichia salictaria*, *Pichia guercuum*, *Pichia pijperi*, *Pichia stiptis*, *Pichia methanolica*, *Pichia* sp., *Saccharomyces cerevisiae*, *Saccharomyces* sp., *Hansenula polymorpha*, *Kluyveromyces* sp., *Candida albicans*,

20 *Aspergillus nidulans*, *Aspergillus niger*, *Aspergillus oryzae*, *Trichoderma reesei*, *Chrysosporium lucknowense*, *Fusarium* sp., *Fusarium gramineum*, *Fusarium venenatum*, and *Neurospora crassa*.

[0221] In each above embodiment, the method is directed to making a host cell in which the oligosaccharide precursors are enriched in $\text{Man}_5\text{GlcNAc}_2$. These structures are desirable because they may then be processed by treatment *in vitro*, for example, using the method of Maras and Contreras, U.S. Patent No. 5,834,251. In a preferred embodiment, however, precursors enriched in $\text{Man}_5\text{GlcNAc}_2$ are processed by at least one further glycosylation reaction *in vivo* -- with glycosidases (*e.g.*, α -mannosidases) and glycosyltransferases (*e.g.*, GnTI) -- to produce human-like *N*-glycans. Oligosaccharide precursors enriched in $\text{Man}_5\text{GlcNAc}_2$, for example, are preferably processed to those having $\text{GlcNAcMan}_X\text{GlcNAc}_2$ core structures, wherein X is 3, 4 or 5, and is preferably 3. *N*-glycans having a

GlcNAcMan_xGlcNAc₂ core structure where X is greater than 3 may be converted to GlcNAcMan₃GlcNAc₂, e.g., by treatment with an α -1,3 and/or α -1,6 mannosidase activity, where applicable. Additional processing of GlcNAcMan₃GlcNAc₂ by treatment with glycosyltransferases (e.g., GnTII)

5 produces GlcNAc₂Man₃GlcNAc₂ core structures which may then be modified, as desired, e.g., by *ex vivo* treatment or by heterologous expression in the host cell of additional glycosylation enzymes, including glycosyltransferases, sugar transporters and mannosidases (see below), to become human-like *N*-glycans.

[0222] Preferred human-like glycoproteins which may be produced according to 10 the invention include those which comprise *N*-glycans having seven or fewer, or three or fewer, mannose residues; and which comprise one or more sugars selected from the group consisting of galactose, GlcNAc, sialic acid, and fucose.

[0223] Another preferred non-human host cell of the invention is a lower 15 eukaryotic cell, e.g., a unicellular or filamentous fungus, which is diminished or depleted in the activity of one or more *alg* gene activities (including an enzymatic activity which is a homolog or equivalent to an *alg* activity). Another preferred host cell of the invention is diminished or depleted in the activity of one or more enzymes (other than *alg* activities) that mannosylate the α -1,6 arm of a lipid-linked oligosaccharide structure.

20 [0224] While lower eukaryotic host cells are preferred, a wide variety of host cells having the aforementioned properties are envisioned as being useful in the methods of the invention. Plant cells, for instance, may be engineered to express a human-like glycoprotein according to the invention. Likewise, a variety of non-human, mammalian host cells may be altered to express more human-like 25 glycoproteins using the methods of the invention. An appropriate host cell can be engineered, or one of the many such mutants already described in yeasts may be used. A preferred host cell of the invention, as exemplified herein, is a hypermannosylation-minus (*OCH1*) mutant in *Pichia pastoris* which has further been modified to delete the *alg3* gene.

30 [0225] The invention additionally provides lower eukaryotic host cells capable of producing glycoproteins having bisected *N*-glycans, such as bisected GlcNAcMan₅GlcNAc₂, GlcNAc₂Man₅GlcNAc₂, GlcNAc₂Man₃GlcNAc₂, and,

preferably, GlcNAc₃Man₃GlcNAc₂. In a preferred embodiment of the invention, the host cells comprise a GnTIII activity. In a more preferred embodiment, the host cells further comprise one or more activities selected from: GnTI, GnTII, GnTIV, and GnTV. Preferred host cells express GnTI, GnTII, and GnTIII. Other 5 preferred host cells additionally express GnTIV and/or GnTV. Even more preferably, the one or more GnT activities of the host cells are substantially intracellular.

[0226] Thus, in preferred embodiments of the invention, host cells comprising the one or more GnT activities produce *N*-glycans comprising structures, including 10 but not limited to GlcNAcMan₃GlcNAc₂, GlcNAcMan₄GlcNAc₂, or GlcNAcMan₅GlcNAc₂, that are capable of reacting with a GnTIII enzyme activity to produce corresponding bisected *N*-glycans. The enzyme activities thereby convert glycoproteins containing these *N*-glycans into forms with new and more desirable properties. Because GnTIII is currently understood to inhibit additional 15 GnT activity in mammalian cells, the skilled artisan should appreciate that sequential glycosylation reaction may or may not be of importance. The present invention contemplates, however, the addition of GnTI and GnTIII in either order or together. It should also be understood that other enzyme activities within the cell, such as, *e.g.*, one or more desired mannosidase activities (*e.g.*, α 1,2 20 mannosidase, Mannosidase I, Mannosidase II), may act in concert with the GnT activities to generate yet other human-like glycoproteins of interest (see Figure 1B).

[0227] In a preferred embodiment, a mannosidase II or a catalytically active fragment thereof is introduced into the host cell to trim the α 1,3 and α 1,6 mannose 25 containing arms of a bisected pentamannose core structure such as GlcNAc₂Man₅GlcNAc₂. The resulting glycans (*e.g.*, bisected GlcNAc₂Man₄GlcNAc₂ and GlcNAc₂Man₃GlcNAc₂) are preferred substrates for subsequent human-like *N*-glycan modification.

[0228] In another embodiment of the invention, the host cells comprise a 30 Man₅GlcNAc₂ core structure or a Man₃GlcNAc₂ core structure modified by two or more GlcNAcs. It should be understood that either core structure may include further modifications in addition to the modification by GlcNAc. Preferably, 10%

or more of the core structures are modified by GlcNAcs. Most preferably, 20%, 30%, 40%, 50%, 60%, 70%, 80% or even more of the core structures contain the GlcNAc modification.

5 Formation of complex *N*-glycans

[0229] Formation of complex *N*-glycan synthesis is a sequential process by which specific sugar residues are removed and attached to the core oligosaccharide structure. In higher eukaryotes, this is achieved by having the substrate sequentially exposed to various processing enzymes. These enzymes carry out 10 specific reactions depending on their particular location within the entire processing cascade. This "assembly line" consists of ER, early, medial and late Golgi, and the trans Golgi network all with their specific processing environment. To re-create the processing of human glycoproteins in the Golgi and ER of lower eukaryotes, numerous enzymes (e.g., glycosyltransferases, glycosidases,

15 phosphatases and transporters) have to be expressed and specifically targeted to these organelles, and preferably, in a location so that they function most efficiently in relation to their environment as well as to other enzymes in the pathway.

[0230] Because one goal of the methods described herein is to achieve a robust protein production strain that is able to perform well in an industrial fermentation 20 process, the integration of multiple genes into the host cell chromosome involves careful planning. As described above, one or more genes which encode enzymes known to be characteristic of non-human glycosylation reactions are preferably deleted. The engineered cell strain is transformed with a range of different genes encoding desired activities, and these genes are transformed in a stable fashion, 25 thereby ensuring that the desired activity is maintained throughout the fermentation process.

[0231] Any combination of the following enzyme activities may be engineered singly or multiply into the host using methods of the invention: sialyltransferases, mannosidases, fucosyltransferases, galactosyltransferases, GlcNAc transferases, 30 ER and Golgi specific transporters (e.g. syn- and antiport transporters for UDP-galactose and other precursors), other enzymes involved in the processing of oligosaccharides, and enzymes involved in the synthesis of activated

oligosaccharide precursors such as UDP-galactose and CMP-N-acetylneurameric acid. Preferably, enzyme activities are introduced on one or more nucleic acid molecules (see also below). Nucleic acid molecules may be introduced singly or multiply, e.g., in the context of a nucleic acid library such as a combinatorial

5 library of the invention. It is to be understood, however, that single or multiple enzymatic activities may be introduced into a host cell in any fashion, including but not limited to protein delivery methods and/or by use of one or more nucleic acid molecules without necessarily using a nucleic acid library or combinatorial library of the invention.

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Expression Of Glycosyltransferases To Produce Complex N-glycans:

[0232] With DNA sequence information, the skilled artisan can clone DNA molecules encoding GnT activities (e.g., Example 3, 8, 11, 15, and 18). Using standard techniques well-known to those of skill in the art, nucleic acid molecules 15 encoding GnTI, II, III, IV or V (or encoding catalytically active fragments thereof) may be inserted into appropriate expression vectors under the transcriptional control of promoters and other expression control sequences capable of driving transcription in a selected host cell of the invention, e.g., a fungal host such as *Pichia* sp., *Kluyveromyces* sp. and *Aspergillus* sp., as described herein, such that 20 one or more of these mammalian GnT enzymes may be actively expressed in a host cell of choice for production of a human-like complex glycoprotein (e.g., Examples 8, 20, and 21).

[0233] Several individual glycosyltransferases have been cloned and expressed in *S. cerevisiae* (GalT, GnTI), *Aspergillus nidulans* (GnTI) and other fungi, 25 without however demonstrating the desired outcome of "humanization" on the glycosylation pattern of the organisms (Yoshida *et al.* (1999) *Glycobiology* 9(1):53-8; Kalsner *et al.* (1995) *Glycoconj. J.* 12(3):360-370). It was speculated that the carbohydrate structure required to accept sugars by the action of such glycosyltransferases was not present in sufficient amounts, which most likely 30 contributed to the lack of complex N-glycan formation.

[0234] A preferred method of the invention provides the functional expression of a GnT, such as GnTI, GnTII, and GnTIII, in the early, medial or late Golgi

apparatus, as well as ensuring a sufficient supply of UDP-GlcNAc (e.g., by expression of a UDP-GlcNAc transporter; see Examples below).

Methods for Providing Sugar Nucleotide Precursors to the Golgi Apparatus:

5 [0235] For a glycosyltransferase to function satisfactorily in the Golgi, the enzyme requires a sufficient concentration of an appropriate nucleotide sugar, which is the high-energy donor of the sugar moiety added to a nascent glycoprotein. In humans, the full range of nucleotide sugar precursors (e.g., UDP-*N*-acetylglucosamine, UDP-*N*-acetylgalactosamine, CMP-*N*-acetylneuraminic acid, 10 UDP-galactose, etc.) are generally synthesized in the cytosol and transported into the Golgi, where they are attached to the core oligosaccharide by glycosyltransferases.

[0236] To replicate this process in non-human host cells, such as lower eukaryotes, sugar nucleoside specific transporters have to be expressed in the 15 Golgi to ensure adequate levels of nucleoside sugar precursors (Sommers and Hirschberg (1981) *J. Cell Biol.* 91(2):A406-A406; Sommers and Hirschberg (1982) *J. Biol. Chem.* 257(18):811-817; Perez and Hirschberg (1987) *Methods in Enzymology* 138:709-715). Nucleotide sugars may be provided to the appropriate compartments, e.g., by expressing in the host microorganism an exogenous gene 20 encoding a sugar nucleotide transporter. The choice of transporter enzyme is influenced by the nature of the exogenous glycosyltransferase being used. For example, a GlcNAc transferase may require a UDP-GlcNAc transporter, a fucosyltransferase may require a GDP-fucose transporter, a galactosyltransferase may require a UDP-galactose transporter, and a sialyltransferase may require a 25 CMP-sialic acid transporter.

[0237] The added transporter protein conveys a nucleotide sugar from the cytosol into the Golgi apparatus, where the nucleotide sugar may be reacted by the glycosyltransferase, e.g., to elongate an *N*-glycan. The reaction liberates a nucleoside diphosphate or monophosphate, e.g., UDP, GDP, or CMP. Nucleoside 30 monophosphates can be directly exported from the Golgi in exchange for nucleoside triphosphate sugars by an antiport mechanism. Accumulation of a nucleoside diphosphate, however, inhibits the further activity of a

glycosyltransferase. As this reaction appears to be important for efficient glycosylation, it is frequently desirable to provide an expressed copy of a gene encoding a nucleotide diphosphatase. The diphosphatase (specific for UDP or GDP as appropriate) hydrolyzes the diphosphonucleoside to yield a nucleoside monophosphate and inorganic phosphate.

[0238] Suitable transporter enzymes, which are typically of mammalian origin, are described below. Such enzymes may be engineered into a selected host cell using the methods of the invention.

[0239] In another example, α 2,3- or α 2,6-sialyltransferase caps galactose residues with sialic acid in the trans-Golgi and TGN of humans leading to a mature form of the glycoprotein (Figure 1B). To reengineer this processing step into a metabolically engineered yeast or fungus will require (1) α 2,3- or α 2,6-sialyltransferase activity and (2) a sufficient supply of CMP-*N*-acetyl neuraminic acid, in the late Golgi of yeast. To obtain sufficient α 2,3-sialyltransferase activity in the late Golgi, for example, the catalytic domain of a known sialyltransferase (e.g. from humans) has to be directed to the late Golgi in fungi (see above). Likewise, transporters have to be engineered to allow the transport of CMP-*N*-acetyl neuraminic acid into the late Golgi. There is currently no indication that fungi synthesize or can even transport sufficient amounts of CMP-*N*-acetyl neuraminic acid into the Golgi. Consequently, to ensure the adequate supply of substrate for the corresponding glycosyltransferases, one has to metabolically engineer the production of CMP-sialic acid into the fungus.

UDP-N-acetylglucosamine

[0240] The cDNA of human UDP-*N*-acetylglucosamine transporter, which was recognized through a homology search in the expressed sequence tags database (dbEST), has been cloned (Ishida (1999) *J. Biochem.* 126(1):68-77). The mammalian Golgi membrane transporter for UDP-*N*-acetylglucosamine was cloned by phenotypic correction with cDNA from canine kidney cells (MDCK) of a recently characterized *Kluyveromyces lactis* mutant deficient in Golgi transport of the above nucleotide sugar (Guillen *et al.* (1998) *Proc. Natl. Acad. Sci. USA* 95(14):7888-7892). Results demonstrate that the mammalian Golgi UDP-GlcNAc

transporter gene has all of the necessary information for the protein to be expressed and targeted functionally to the Golgi apparatus of yeast and that two proteins with very different amino acid sequences may transport the same solute within the same Golgi membrane (Guillen *et al.* (1998) *Proc. Natl. Acad. Sci. USA* 95(14):7888-5 7892).

[0241] Accordingly, one may incorporate the expression of a UDP-GlcNAc transporter in a host cell by means of a nucleic acid construct which may contain, for example: (1) a region by which the transformed construct is maintained in the cell (*e.g.*, origin of replication or a region that mediates chromosomal integration), 10 (2) a marker gene that allows for the selection of cells that have been transformed, including counterselectable and recyclable markers such as *ura3* or *T-urf13* (Soderholm *et al.* (2001) *Biotechniques* 31(2):306-10) or other well characterized selection-markers (*e.g.*, *his4*, *bla*, *Sh ble* etc.), (3) a gene or fragment thereof encoding a functional UDP-GlcNAc transporter (*e.g.*, from *K. lactis*, (Abeijon, 15 (1996) *Proc. Natl. Acad. Sci. U.S.A.* 93:5963-5968), or from *H. sapiens* (Ishida *et al.* (1996) *J. Biochem. (Tokyo)* 120(6):1074-8), and (4) a promoter activating the expression of the above mentioned localization/catalytic domain fusion construct library.

20 *GDP-Fucose*

[0242] The rat liver Golgi membrane GDP-fucose transporter has been identified and purified by Puglielli and Hirschberg (1999) *J. Biol. Chem.* 274(50):35596-25 35600. The corresponding gene has not been identified, however, *N*-terminal sequencing can be used for the design of oligonucleotide probes specific for the corresponding gene. These oligonucleotides can be used as probes to clone the gene encoding for GDP-fucose transporter.

UDP-Galactose

[0243] Two heterologous genes, *gmal2(+)* encoding alpha 1,2-galactosyltransferase (alpha 1,2 GalT) from *Schizosaccharomyces pombe* and (30 *hUGT2*) encoding human UDP-galactose (UDP-Gal) transporter, have been functionally expressed in *S. cerevisiae* to examine the intracellular conditions

required for galactosylation. Correlation between protein galactosylation and UDP-galactose transport activity indicated that an exogenous supply of UDP-Gal transporter, rather than alpha 1,2 GalT played a key role for efficient galactosylation in *S. cerevisiae* (Kainuma (1999) *Glycobiology* 9(2):133-141).

5 Likewise, an UDP-galactose transporter from *S. pombe* was cloned (Segawa (1999) *FEBS Letters* 451(3):295-298).

CMP-N-acetylneurameric acid (CMP-Sialic acid).

[0244] Human CMP-sialic acid transporter (hCST) has been cloned and expressed in Lec 8 CHO cells (Aoki *et al.* (1999) *J. Biochem. (Tokyo)* 126(5):940-50; Eckhardt *et al.* (1997) *Eur. J. Biochem.* 248(1):187-92). The functional expression of the murine CMP-sialic acid transporter was achieved in *Saccharomyces cerevisiae* (Berninsone *et al.* (1997) *J. Biol. Chem.* 272(19):12616-9). Sialic acid has been found in some fungi, however it is not clear whether the chosen host system will be able to supply sufficient levels of CMP-Sialic acid. Sialic acid can be either supplied in the medium or alternatively fungal pathways involved in sialic acid synthesis can also be integrated into the host genome.

Expression of Diphosphatases:

[0245] When sugars are transferred onto a glycoprotein, either a nucleoside diphosphate or monophosphate is released from the sugar nucleotide precursors. While monophosphates can be directly exported in exchange for nucleoside triphosphate sugars by an antiport mechanism, diphosphonucleosides (*e.g.* GDP) have to be cleaved by phosphatases (*e.g.* GDPase) to yield nucleoside monophosphates and inorganic phosphate prior to being exported. This reaction appears to be important for efficient glycosylation, as GDPase from *S. cerevisiae* has been found to be necessary for mannosylation. However, the enzyme only has 10% of the activity towards UDP (Berninsone *et al.* (1994) *J. Biol. Chem.* 269(1):207-211). Lower eukaryotes often do not have UDP-specific diphosphatase activity in the Golgi as they do not utilize UDP-sugar precursors for glycoprotein synthesis in the Golgi. *Schizosaccharomyces pombe*, a yeast which adds galactose residues to cell wall polysaccharides (from UDP-galactose), was found to have

specific UDPase activity, further suggesting the requirement for such an enzyme (Berninsone *et al.* (1994) *J. Biol. Chem.* 269(1):207-211). UDP is known to be a potent inhibitor of glycosyltransferases and the removal of this glycosylation side product is important to prevent glycosyltransferase inhibition in the lumen of the Golgi (Khatara *et al.* (1974) *Eur. J. Biochem.* 44:537-560).

Methods For Altering N-Glycans in a Host By Expressing A Targeted Enzymatic Activity From a Nucleic Acid Molecule

[0246] The present invention further provides a method for producing a human-like glycoprotein in a non-human host cell comprising the step of introducing into the cell one or more nucleic acid molecules which encode an enzyme or enzymes for production of the $\text{Man}_5\text{GlcNAc}_2$ carbohydrate structure. In one preferred embodiment, a nucleic acid molecule encoding one or more mannosidase activities involved in the production of $\text{Man}_5\text{GlcNAc}_2$ from $\text{Man}_8\text{GlcNAc}_2$ or $\text{Man}_9\text{GlcNAc}_2$ is introduced into the host. The invention additionally relates to methods for making altered glycoproteins in a host cell comprising the step of introducing into the host cell a nucleic acid molecule which encodes one or more glycosylation enzymes or activities. Preferred enzyme activities are selected from the group consisting of UDP-GlcNAc transferase, UDP-galactosyltransferase, GDP-fucosyltransferase, CMP-sialyltransferase, UDP-GlcNAc transporter, UDP-galactose transporter, GDP-fucose transporter, CMP-sialic acid transporter, and nucleotide diphosphatases. In a particularly preferred embodiment, the host is selected or engineered to express two or more enzymatic activities in which the product of one activity increases substrate levels of another activity, *e.g.*, a glycosyltransferase and a corresponding sugar transporter, *e.g.*, GnTI and UDP-GlcNAc transporter activities. In another preferred embodiment, the host is selected or engineered to express an activity to remove products which may inhibit subsequent glycosylation reactions, *e.g.* a UDP- or GDP-specific diphosphatase activity.

20 [0247] Preferred methods of the invention involve expressing one or more enzymatic activities from a nucleic acid molecule in a host cell and comprise the step of targeting at least one enzymatic activity to a desired subcellular location

(e.g., an organelle) by forming a fusion protein comprising a catalytic domain of the enzyme and a cellular targeting signal peptide, e.g., a heterologous signal peptide which is not normally ligated to or associated with the catalytic domain. The fusion protein is encoded by at least one genetic construct ("fusion construct")

5 comprising a nucleic acid fragment encoding a cellular targeting signal peptide ligated in the same translational reading frame ("in-frame") to a nucleic acid fragment encoding an enzyme (e.g., glycosylation enzyme), or catalytically active fragment thereof.

[0248] The targeting signal peptide component of the fusion construct or protein

10 is preferably derived from a member of the group consisting of: membrane-bound proteins of the ER or Golgi, retrieval signals, Type II membrane proteins, Type I membrane proteins, membrane spanning nucleotide sugar transporters, mannosidases, sialyltransferases, glucosidases, mannosyltransferases and phosphomannosyltransferases.

15 [0249] The catalytic domain component of the fusion construct or protein is preferably derived from a glycosidase, mannosidase or a glycosyltransferase activity derived from a member of the group consisting of GnT I, GnT II, GnT III, GnT IV, GnT V, GnT VI, GalT, Fucosyltransferase and Sialyltransferase. The catalytic domain preferably has a pH optimum within 1.4 pH units of the average

20 pH optimum of other representative enzymes in the organelle in which the enzyme is localized, or has optimal activity at a pH between 5.1 and 8.0. In a preferred embodiment, the catalytic domain encodes a mannosidase selected from the group consisting of *C. elegans* mannosidase IA, *C. elegans* mannosidase IB, *D. melanogaster* mannosidase IA, *H. sapiens* mannosidase IB, *P. citrinum*

25 mannosidase I, mouse mannosidase IA, mouse mannosidase IB, *A. nidulans* mannosidase IA, *A. nidulans* mannosidase IB, *A. nidulans* mannosidase IC, mouse mannosidase II, *C. elegans* mannosidase II, *H. sapiens* mannosidase II, mannosidase IIX, and mannosidase III.

30 **Selecting a Glycosylation Enzyme: pH Optima and Subcellular Localization**

[0250] In one embodiment of the invention, a human-like glycoprotein is made efficiently in a non-human eukaryotic host cell by introducing into a subcellular

compartment of the cell a glycosylation enzyme selected to have a pH optimum similar to the pH optima of other enzymes in the targeted subcellular compartment.

For example, most enzymes that are active in the ER and Golgi apparatus of *S. cerevisiae* have pH optima that are between about 6.5 and 7.5 (see Table 3).

5 Because the glycosylation of proteins is a highly evolved and efficient process, the internal pH of the ER and the Golgi is likely also in the range of about 6-8. All previous approaches to reduce mannosylation by the action of recombinant mannosidases in fungal hosts, however, have introduced enzymes that have a pH optimum of around pH 5.0 (Martinet *et al.* (1998) *Biotech. Letters* 20(12): 1171-1177, and Chiba *et al.* (1998) *J. Biol. Chem.* 273(41): 26298-26304). At pH 7.0, the *in vitro* determined activity of those mannosidases is reduced to less than 10%, which is likely insufficient activity at their point of use, namely, the ER and early Golgi, for the efficient *in vivo* production of Man₅GlcNAc₂ on *N*-glycans.

10 [0251] Accordingly, a preferred embodiment of this invention targets a selected glycosylation enzyme (or catalytic domain thereof), *e.g.*, an α -mannosidase, to a subcellular location in the host cell (*e.g.*, an organelle) where the pH optimum of the enzyme or domain is within 1.4 pH units of the average pH optimum of other representative marker enzymes localized in the same organelle(s). The pH optimum of the enzyme to be targeted to a specific organelle should be matched 15 with the pH optimum of other enzymes found in the same organelle to maximize the activity per unit enzyme obtained. Table 3 summarizes the activity of mannosidases from various sources and their respective pH optima. Table 4 summarizes their typical subcellular locations.

Table 3. Mannosidases and their pH optimum.

Source	Enzyme	pH optimum	Reference
<i>Aspergillus saitoi</i>	α -1,2-mannosidase	5.0	Ichishima <i>et al.</i> (1999) <i>Biochem. J.</i> 339(Pt 3):589-597
<i>Trichoderma reesei</i>	α -1,2-mannosidase	5.0	Maras <i>et al.</i> (2000) <i>J. Biotechnol.</i> 77(2-3):255-263
<i>Penicillium citrinum</i>	α -D-1,2-mannosidase	5.0	Yoshida <i>et al.</i> (1993) <i>Biochem. J.</i> 290(Pt 2):349-354
<i>C. elegans</i>	α -1,2-mannosidase	5.5	see Figure 11
<i>Aspergillus nidulans</i>	α -1,2-mannosidase	6.0	Eades and Hintz (2000) <i>Gene</i> 255(1):25-34
<i>Homo sapiens</i> IA(Golgi)	α -1,2-mannosidase	6.0	
<i>Homo sapiens</i> IB (Golgi)	α -1,2-mannosidase	6.0	
<i>Lepidopteran</i> insect cells	Type I α -1,2-Man ₆ -mannosidase	6.0	Ren <i>et al.</i> (1995) <i>Biochem.</i> 34(8):2489-2495
<i>Homo sapiens</i>	α -D-mannosidase	6.0	Chandrasekaran <i>et al.</i> (1984) <i>Cancer Res.</i> 44(9):4059-68
<i>Xanthomonas manihotis</i>	α -1,2,3-mannosidase	6.0	U.S. Pat. No. 6,300,113
<i>Mouse</i> IB (Golgi)	α -1,2-mannosidase	6.5	Schneikert and Herscovics (1994) <i>Glycobiology.</i> 4(4):445-50
<i>Bacillus</i> sp. (secreted)	α -D-1,2-mannosidase	7.0	Maruyama <i>et al.</i> (1994) <i>Carbohydrate Res.</i> 251:89-98

[0252] In a preferred embodiment, a particular enzyme or catalytic domain is targeted to a subcellular location in the host cell by means of a chimeric fusion construct encoding a protein comprising a cellular targeting signal peptide not normally associated with the enzymatic domain. Preferably, an enzyme or domain is targeted to the ER, the early, medial or late Golgi, or the trans Golgi apparatus of the host cell.

[0253] In a more preferred embodiment, the targeted glycosylation enzyme is a mannosidase, glycosyltransferase or a glycosidase. In an especially preferred embodiment, mannosidase activity is targeted to the ER or cis Golgi, where the early reactions of glycosylation occur. While this method is useful for producing a

5 human-like glycoprotein in a non-human host cell, it will be appreciated that the method is also useful more generally for modifying carbohydrate profiles of a glycoprotein in any eukaryotic host cell, including human host cells.

[0254] Targeting sequences which mediate retention of proteins in certain organelles of the host cell secretory pathway are well-known and described in the

10 scientific literature and public databases, as discussed in more detail below with respect to libraries for selection of targeting sequences and targeted enzymes. Such subcellular targeting sequences may be used alone or in combination to target a selected glycosylation enzyme (or catalytic domain thereof) to a particular subcellular location in a host cell, *i.e.*, especially to one where the enzyme will

15 have enhanced or optimal activity based on pH optima or the presence of other stimulatory factors.

[0255] When one attempts to trim high mannose structures to yield $\text{Man}_5\text{GlcNAc}_2$ in the ER or the Golgi apparatus of a host cell such as *S. cerevisiae*, for example, one may choose any enzyme or combination of enzymes that (1) has a

20 sufficiently close pH optimum (*i.e.*, between pH 5.2 and pH 7.8), and (2) is known to generate, alone or in concert, the specific isomeric $\text{Man}_5\text{GlcNAc}_2$ structure required to accept subsequent addition of GlcNAc by GnTI. Any enzyme or combination of enzymes that is shown to generate a structure that can be converted to $\text{GlcNAcMan}_5\text{GlcNAc}_2$ by GnTI *in vitro* would constitute an appropriate choice.

25 This knowledge may be obtained from the scientific literature or experimentally. For example, one may determine whether a potential mannosidase can convert $\text{Man}_8\text{GlcNAc}_2\text{-2AB}$ (2-aminobenzamide) to $\text{Man}_5\text{GlcNAc}_2\text{-AB}$ and then verify that the obtained $\text{Man}_5\text{GlcNAc}_2\text{-2AB}$ structure can serve a substrate for GnTI and UDP-GlcNAc to give $\text{GlcNAcMan}_5\text{GlcNAc}_2$ *in vitro*. Mannosidase IA from a

30 human or murine source, for example, would be an appropriate choice (*see, e.g., Example 4*). Examples described herein utilize 2-aminobenzamide labeled *N*-linked oligomannose followed by HPLC analysis to make this determination.

Table 4. Cellular location and pH optima of various glycosylation-related enzymes of *S. cerevisiae*.

Gene	Activity	Location	pH optimum	Reference(s)
<i>KTR1</i>	α - 1,2 mannosyltransferase	Golgi	7.0	Romero <i>et al.</i> (1997) <i>Biochem. J.</i> 321(Pt 2):289-295
<i>MNS1</i>	α - 1,2- mannosidase	ER	6.5	Lipari <i>et al.</i> (1994) <i>Glycobiology.</i> Oct;4(5):697-702
<i>CWH41</i>	glucosidase I mannosyltransferase	ER Golgi	6.8 7-8	Lehele and Tanner (1974) <i>Biochim. Biophys. Acta</i> 350(1):225-235
<i>KRE2</i>	α - 1,2 mannosyltransferase	Golgi	6.5-9.0	Romero <i>et al.</i> (1997) <i>Biochem. J.</i> 321(Pt 2):289-295

5 [0256] Accordingly, a glycosylation enzyme such as an α -1,2-mannosidase enzyme used according to the invention has an optimal activity at a pH of between 5.1 and 8.0. In a preferred embodiment, the enzyme has an optimal activity at a pH of between 5.5 and 7.5. The *C. elegans* mannosidase enzyme, for example, works well in the methods of the invention and has an apparent pH optimum of about 5.5). Preferred mannosidases include those listed in Table 3 having appropriate pH optima, *e.g.* *Aspergillus nidulans*, *Homo sapiens* IA (Golgi), *Homo sapiens* IB (Golgi), *Lepidopteran* insect cells (IPLB-SF21AE), *Homo sapiens*, mouse IB (Golgi), *Xanthomonas manihotis*, *Drosophila melanogaster* and *C. elegans*.

10 15 [0257] An experiment which illustrates the pH optimum for an α -1,2-mannosidase enzyme is described in Example 7. A chimeric fusion protein BB27-2 (*Saccharomyces MNN10* (s)/*C. elegans* mannosidase IB Δ 31), which leaks into the medium was subjected to various pH ranges to determine the optimal activity

of the enzyme. The results of the experiment show that the α -1,2-mannosidase has an optimal pH of about 5.5 for its function (Figure 11).

[0258] In a preferred embodiment, a single cloned mannosidase gene is expressed in the host organism. However, in some cases it may be desirable to 5 express several different mannosidase genes, or several copies of one particular gene, in order to achieve adequate production of $\text{Man}_5\text{GlcNAc}_2$. In cases where multiple genes are used, the encoded mannosidases preferably all have pH optima within the preferred range of about 5.1 to about 8.0, or especially between about 5.5 and about 7.5. Preferred mannosidase activities include α -1,2-mannosidases 10 derived from mouse, human, *Lepidoptera*, *Aspergillus nidulans*, or *Bacillus* sp., *C. elegans*, *D. melanogaster*, *P. citrinum*, *X. laevis* or *A. nidulans*.

In Vivo Alteration of Host Cell Glycosylation Using a Combinatorial DNA Library

15 [0259] Certain methods of the invention are preferably (but need not necessarily be) carried out using one or more nucleic acid libraries. An exemplary feature of a combinatorial nucleic acid library of the invention is that it comprises sequences encoding cellular targeting signal peptides and sequences encoding proteins to be targeted (e.g., enzymes or catalytic domains thereof, including but not limited to 20 those which mediate glycosylation).

[0260] In one embodiment, a combinatorial nucleic acid library comprises: (a) at least two nucleic acid sequences encoding different cellular targeting signal peptides; and (b) at least one nucleic acid sequence encoding a polypeptide to be targeted. In another embodiment, a combinatorial nucleic acid library comprises: 25 (a) at least one nucleic acid sequence encoding a cellular targeting signal peptide; and (b) at least two nucleic acid sequences encoding a polypeptide to be targeted into a host cell. As described further below, a nucleic acid sequence derived from (a) and a nucleic acid sequence derived from (b) are ligated to produce one or more fusion constructs encoding a cellular targeting signal peptide functionally linked to 30 a polypeptide domain of interest. One example of a functional linkage is when the cellular targeting signal peptide is ligated to the polypeptide domain of interest in the same translational reading frame ("in-frame").

[0261] In a preferred embodiment, a combinatorial DNA library expresses one or more fusion proteins comprising cellular targeting signal peptides ligated in-frame to catalytic enzyme domains. The encoded fusion protein preferably comprises a catalytic domain of an enzyme involved in mammalian- or human-like 5 modification of *N*-glycans. In a more preferred embodiment, the catalytic domain is derived from an enzyme selected from the group consisting of mannosidases, glycosyltransferases and other glycosidases which is ligated in-frame to one or more targeting signal peptides. The enzyme domain may be exogenous and/or endogenous to the host cell. A particularly preferred signal peptide is one 10 normally associated with a protein that undergoes ER to Golgi transport.

[0262] The combinatorial DNA library of the present invention may be used for producing and localizing *in vivo* enzymes involved in mammalian- or human-like *N*-glycan modification. The fusion constructs of the combinatorial DNA library are engineered so that the encoded enzymes are localized in the ER, Golgi or the 15 trans-Golgi network of the host cell where they are involved in producing particular *N*-glycans on a glycoprotein of interest. Localization of *N*-glycan modifying enzymes of the present invention is achieved through an anchoring mechanism or through protein-protein interaction where the localization peptide constructed from the combinatorial DNA library localizes to a desired organelle of 20 the secretory pathway such as the ER, Golgi or the trans Golgi network.

[0263] An example of a useful *N*-glycan, which is produced efficiently and in sufficient quantities for further modification by human-like (complex) glycosylation reactions is $\text{Man}_5\text{GlcNAc}_2$. A sufficient amount of $\text{Man}_5\text{GlcNAc}_2$ is needed on a glycoprotein of interest for further human-like processing *in vivo* (e.g., 25 more than 30 mole %). The $\text{Man}_5\text{GlcNAc}_2$ intermediate may be used as a substrate for further *N*-glycan modification to produce $\text{GlcNAcMan}_5\text{GlcNAc}_2$ (Figure 1B; see above). Accordingly, the combinatorial DNA library of the present invention may be used to produce enzymes that subsequently produce $\text{GlcNAcMan}_5\text{GlcNAc}_2$, or other desired complex *N*-glycans, in a useful quantity.

30 [0264] A further aspect of the fusion constructs produced using the combinatorial DNA library of the present invention is that they enable sufficient and often near complete intracellular *N*-glycan trimming activity in the engineered host cell.

Preferred fusion constructs produced by the combinatorial DNA library of the invention encode a glycosylation enzyme, e.g., a mannosidase, which is effectively localized to an intracellular host cell compartment and thereby exhibits very little and preferably no extracellular activity. The preferred fusion constructs of the present invention that encode a mannosidase enzyme are shown to localize where the *N*-glycans are modified, namely, the ER and the Golgi. The fusion enzymes of the present invention are targeted to such particular organelles in the secretory pathway where they localize and act upon *N*-glycans such as $\text{Man}_8\text{GlcNAc}_2$ to produce $\text{Man}_5\text{GlcNAc}_2$ on a glycoprotein of interest.

[0265] GnTIII fusion constructs generated from a combinatorial DNA library to produce bisected glycans were assayed to determine any extracellular activity. An example of a GnTIII fusion constructs exhibiting *in vivo* alteration of host cell glycosylation is designated pVA53. After transforming *P. pastoris* YSH-1 with the fusion construct pVA53, the supernatant was tested to detect any *ex vivo* GnTIII activity. Figure 33 shows no apparent change in the standard substrate GlcNAc $\text{Man}_5\text{GlcNAc}_2$ under conditions that would reveal extracellular GnTIII activity in the medium (Example 22). Similarly, Figure 34 shows no detectable extracellular GnTIII activity in the medium in *P. pastoris* YSH-57 reacting with the substrate GlcNAc $_2\text{Man}_3\text{GlcNAc}_2$ (Example 23).

[0266] Enzymes produced by the combinatorial DNA library of the present invention can modify *N*-glycans on a glycoprotein of interest as shown for K3 or IFN- β proteins expressed in *P. pastoris*, as shown in Figures 5, 6, and 25-34 (see also Examples 2, 4, and 18-23). It is, however, appreciated that other types of glycoproteins, without limitation, including erythropoietin, cytokines such as interferon- α , interferon- β , interferon- γ , interferon- ω , and granulocyte-CSF, coagulation factors such as factor VIII, factor IX, and human protein C, soluble IgE receptor α -chain, IgG, IgG fragments, IgM, interleukins, urokinase, chymase, and urea trypsin inhibitor, IGF-binding protein, epidermal growth factor, growth hormone-releasing factor, annexin V fusion protein, angiostatin, vascular endothelial growth factor-2, myeloid progenitor inhibitory factor-1, osteoprotegerin, α -1 antitrypsin, DNase II, α -feto proteins, AAT, rhTBP-1 (oncept, aka TNF Binding protein 1), TACI-Ig (transmembrane activator and

calcium modulator and cyclophilin ligand interactor), FSH (follicle stimulating hormone), GM-CSF, GLP-1 w/ and w/o FC (glucagon like protein 1) IL-1 receptor agonist, sTNFr (enbrel, aka soluble TNF receptor Fc fusion) ATIII, rhThrombin, glucocerebrosidase and CTLA4-Ig (Cytotoxic T Lymphocyte associated Antigen 4 - Ig) may be glycosylated in this way.

Constructing a Combinatorial DNA Library of Fusion Constructs:

[0267] A combinatorial DNA library of fusion constructs features one or more cellular targeting signal peptides ("targeting peptides") generally derived from *N*-terminal domains of native proteins (e.g., by making C-terminal deletions). Some targeting peptides, however, are derived from the C-terminus of native proteins (e.g. SEC12). Membrane-bound proteins of the ER or the Golgi are preferably used as a source for targeting peptide sequences. These proteins have sequences encoding a cytosolic tail (ct), a transmembrane domain (tmd) and a stem region (sr) which are varied in length. These regions are recognizable by protein sequence alignments and comparisons with known homologs and/or other localized proteins (e.g., comparing hydrophobicity plots).

[0268] The targeting peptides are indicated herein as short (s), medium (m) and long (l) relative to the parts of a type II membrane. The targeting peptide sequence indicated as short (s) corresponds to the transmembrane domain (tmd) of the membrane-bound protein. The targeting peptide sequence indicated as long (l) corresponds to the length of the transmembrane domain (tmd) and the stem region (sr). The targeting peptide sequence indicated as medium (m) corresponds to the transmembrane domain (tmd) and approximately half the length of the stem region (sr). The catalytic domain regions are indicated herein by the number of nucleotide deletion with respect to its wild-type glycosylation enzyme.

Sub-libraries

[0269] In some cases a combinatorial nucleic acid library of the invention may be assembled directly from existing or wild-type genes. In a preferred embodiment, the DNA library is assembled from the fusion of two or more sub-libraries. By the in-frame ligation of the sub-libraries, it is possible to create a

large number of novel genetic constructs encoding useful targeted protein domains such as those which have glycosylation activities.

Catalytic Domain Sub-Libraries Encoding Glycosylation Activities

5 [0270] One useful sub-library includes DNA sequences encoding enzymes such as glycosidases (*e.g.*, mannosidases), glycosyltransferases (*e.g.*, fucosyltransferases, galactosyltransferases, glucosyltransferases), GlcNAc transferases and sialyltransferases. Catalytic domains may be selected from the host to be engineered, as well as from other related or unrelated organisms. Mammalian,

10 plant, insect, reptile, algal or fungal enzymes are all useful and should be chosen to represent a broad spectrum of biochemical properties with respect to temperature and pH optima. In a preferred embodiment, genes are truncated to give fragments some of which encode the catalytic domains of the enzymes. By removing endogenous targeting sequences, the enzymes may then be redirected and

15 expressed in other cellular loci.

[0271] The choice of such catalytic domains may be guided by the knowledge of the particular environment in which the catalytic domain is subsequently to be active. For example, if a particular glycosylation enzyme is to be active in the late Golgi, and all known enzymes of the host organism in the late Golgi have a certain 20 pH optimum, or the late Golgi is known to have a particular pH, then a catalytic domain is chosen which exhibits adequate, and preferably maximum, activity at that pH, as discussed above.

Targeting Peptide Sequence Sub-Libraries

25 [0272] Another useful sub-library includes nucleic acid sequences encoding targeting signal peptides that result in localization of a protein to a particular location within the ER, Golgi, or trans Golgi network. These targeting peptides may be selected from the host organism to be engineered as well as from other related or unrelated organisms. Generally such sequences fall into three

30 categories: (1) *N*-terminal sequences encoding a cytosolic tail (ct), a transmembrane domain (tmd) and part or all of a stem region (sr), which together or individually anchor proteins to the inner (lumenal) membrane of the Golgi; (2)

retrieval signals which are generally found at the C-terminus such as the HDEL (SEQ ID NO:41) or KDEL (SEQ ID NO:42) tetrapeptide; and (3) membrane spanning regions from various proteins, e.g., nucleotide sugar transporters, which are known to localize in the Golgi.

5 [0273] In the first case, where the targeting peptide consists of various elements (ct, tmd and sr), the library is designed such that the ct, the tmd and various parts of the stem region are represented. Accordingly, a preferred embodiment of the sub-library of targeting peptide sequences includes ct, tmd, and/or sr sequences from membrane-bound proteins of the ER or Golgi. In some cases it may be
 10 desirable to provide the sub-library with varying lengths of sr sequence. This may be accomplished by PCR using primers that bind to the 5' end of the DNA encoding the cytosolic region and employing a series of opposing primers that bind to various parts of the stem region.

[0274] Still other useful sources of targeting peptide sequences include retrieval
 15 signal peptides, e.g. the tetrapeptides HDEL or KDEL, which are typically found at the C-terminus of proteins that are transported retrograde into the ER or Golgi. Still other sources of targeting peptide sequences include (a) type II membrane proteins, (b) the enzymes listed in Table 3, (c) membrane spanning nucleotide sugar transporters that are localized in the Golgi, and (d) sequences referenced in
 20 Table 5.

Table 5. Sources of useful compartmental targeting sequences

<i>Gene or Sequence</i>	<i>Organism</i>	<i>Function</i>	<i>Location of Gene Product</i>
<i>MNSI</i>	<i>A. nidulans</i>	α -1,2-mannosidase	ER
<i>MNSI</i>	<i>A. niger</i>	α -1,2-mannosidase	ER
<i>MNSI</i>	<i>S. cerevisiae</i>	α -1,2-mannosidase	ER
<i>GLSI</i>	<i>S. cerevisiae</i>	glucosidase	ER
<i>GLSI</i>	<i>A. niger</i>	glucosidase	ER
<i>GLSI</i>	<i>A. nidulans</i>	glucosidase	ER
HDEL at C-terminus	<i>Universal in fungi</i>	retrieval signal	ER
<i>SEC12</i>	<i>S. cerevisiae</i>	COPII vesicle protein	ER/Golgi

<i>SEC12</i>	<i>A. niger</i>	COPII vesicle protein	ER/Golgi
<i>OCH1</i>	<i>S. cerevisiae</i>	1,6-mannosyltransferase	Golgi (cis)
<i>OCH1</i>	<i>P. pastoris</i>	1,6-mannosyltransferase	Golgi (cis)
<i>MNN9</i>	<i>S. cerevisiae</i>	1,6-mannosyltransferase complex	Golgi
<i>MNN9</i>	<i>A. niger</i>	undetermined	Golgi
<i>VANI</i>	<i>S. cerevisiae</i>	undetermined	Golgi
<i>VANI</i>	<i>A. niger</i>	undetermined	Golgi
<i>ANPI</i>	<i>S. cerevisiae</i>	undetermined	Golgi
<i>HOCl</i>	<i>S. cerevisiae</i>	undetermined	Golgi
<i>MNN10</i>	<i>S. cerevisiae</i>	undetermined	Golgi
<i>MNN10</i>	<i>A. niger</i>	undetermined	Golgi
<i>MNN11</i>	<i>S. cerevisiae</i>	undetermined	Golgi (cis)
<i>MNN11</i>	<i>A. niger</i>	undetermined	Golgi (cis)
<i>MNT1</i>	<i>S. cerevisiae</i>	1,2-mannosyltransferase	Golgi (cis, medial)
<i>KTR1</i>	<i>P. pastoris</i>	undetermined	Golgi (medial)
<i>KRE2</i>	<i>P. pastoris</i>	undetermined	Golgi (medial)
<i>KTR3</i>	<i>P. pastoris</i>	undetermined	Golgi (medial)
<i>MNN2</i>	<i>S. cerevisiae</i>	1,2-mannosyltransferase	Golgi (medial)
<i>KTR1</i>	<i>S. cerevisiae</i>	undetermined	Golgi (medial)
<i>KTR2</i>	<i>S. cerevisiae</i>	undetermined	Golgi (medial)
<i>MNN1</i>	<i>S. cerevisiae</i>	1,3-mannosyltransferase	Golgi (trans)
<i>MNN6</i>	<i>S. cerevisiae</i>	Phosphomannosyltransferase	Golgi (trans)
2,6 ST	<i>H. sapiens</i>	2,6-sialyltransferase	trans Golgi network
UDP-Gal T	<i>S. pombe</i>	UDP-Gal transporter	Golgi

[0275] In any case, it is highly preferred that targeting peptide sequences are selected which are appropriate for the particular enzymatic activity or activities to function optimally within the sequence of desired glycosylation reactions. For example, in developing a modified microorganism capable of terminal sialylation of nascent *N*-glycans, a process which occurs in the late Golgi in humans, it is

desirable to utilize a sub-library of targeting peptide sequences derived from late Golgi proteins. Similarly, the trimming of $\text{Man}_8\text{GlcNAc}_2$ by an α -1,2-mannosidase to give $\text{Man}_5\text{GlcNAc}_2$ is an early step in complex *N*-glycan formation in humans (Figure 1B). It is therefore desirable to have this reaction occur in the ER or early 5 Golgi of an engineered host microorganism. A sub-library encoding ER and early Golgi retention signals is used.

[0276] A series of fusion protein constructs (*i.e.*, a combinatorial DNA library) is then constructed by functionally linking one or a series of targeting peptide sequences to one or a series of sequences encoding catalytic domains. In a 10 preferred embodiment, this is accomplished by the in-frame ligation of a sub-library comprising DNA encoding targeting peptide sequences (above) with a sub-library comprising DNA encoding glycosylation enzymes or catalytically active fragments thereof (see below).

[0277] The resulting library comprises synthetic genes encoding targeting 15 peptide sequence-containing fusion proteins. In some cases it is desirable to provide a targeting peptide sequence at the N-terminus of a fusion protein, or in other cases at the C-terminus. In some cases, targeting peptide sequences may be inserted within the open reading frame of an enzyme, provided the protein structure of individual folded domains is not disrupted. Each type of fusion protein 20 is constructed (in a step-wise directed or semi-random fashion) and optimal constructs may be selected upon transformation of host cells and characterization of glycosylation patterns in transformed cells using methods of the invention.

25 Alteration of Host Cell Glycosylation Using Fusion Constructs From Combinatorial Libraries:

[0278] The construction of a preferred combinatorial DNA library is illustrated schematically in Figure 2 and described in Example 4. The fusion construct may be operably linked to a multitude of vectors, such as expression vectors well-known in the art. A wide variety of such fusion constructs were assembled using representative activities as shown in Table 6. Combinations of targeting 30 peptide/catalytic domains may be assembled for use in targeting mannosidase, glycosyltransferase and glycosidase activities in the ER, Golgi, and the trans Golgi network according to the invention. Surprisingly, the same catalytic domain may

have no effect to a very profound effect on N-glycosylation patterns, depending on the type of targeting peptide used (see, e.g., Table 7, Example 4).

Mannosidase Fusion Constructs

5 [0279] A representative example of a mannosidase fusion construct derived from a combinatorial DNA library of the invention is pFB8, which a truncated *Saccharomyces SEC12*(m) targeting peptide (988-1296 nucleotides of *SEC12* from SwissProt P11655) ligated in-frame to a 187 N-terminal amino acid deletion of a mouse α -mannosidase IA (Genbank AN 6678787). The nomenclature used herein, 10 thus, refers to the targeting peptide/catalytic domain region of a glycosylation enzyme as *Saccharomyces SEC12* (m)/mouse mannosidase IA Δ 187. The encoded fusion protein localizes in the ER by means of the *SEC12* targeting peptide sequence while retaining its mannosidase catalytic domain activity and is capable of producing *in vivo* N-glycans having a Man₅GlcNAc₂ structure (Example 4; 15 Figures 6F and 7B).

[0280] The fusion construct pGC5, *Saccharomyces MNS1*(m)/mouse mannosidase IB Δ 99, is another example of a fusion construct having intracellular mannosidase trimming activity (Example 4; Figures 5D and 8E). Fusion construct pBC18-5 (*Saccharomyces VAN1*(s)/*C. elegans* mannosidase IB Δ 80) is 20 yet another example of an efficient fusion construct capable of producing N-glycans having a Man₅GlcNAc₂ structure *in vivo*. By creating a combinatorial DNA library of these and other such mannosidase fusion constructs according to the invention, a skilled artisan may distinguish and select those constructs having optimal intracellular trimming activity from those having relatively low or no 25 activity. Methods using combinatorial DNA libraries of the invention are advantageous because only a select few mannosidase fusion constructs may produce a particularly desired N-glycan *in vivo*.

[0281] In addition, mannosidase trimming activity may be specific to a particular protein of interest. Thus, it is to be further understood that not all targeting 30 peptide/mannosidase catalytic domain fusion constructs may function equally well to produce the proper glycosylation on a glycoprotein of interest. Accordingly, a protein of interest may be introduced into a host cell transfected with a

combinatorial DNA library to identify one or more fusion constructs which express a mannosidase activity optimal for the protein of interest. One skilled in the art will be able to produce and select optimal fusion construct(s) using the combinatorial DNA library approach described herein.

5 [0282] It is apparent, moreover, that other such fusion constructs exhibiting localized active mannosidase catalytic domains (or more generally, domains of any enzyme) may be made using techniques such as those exemplified in **Example 4** and described herein. It will be a matter of routine experimentation for one skilled in the art to make and use the combinatorial DNA library of the present invention
10 to optimize, for example, $\text{Man}_5\text{GlcNAc}_2$ production from a library of fusion constructs in a particular expression vector introduced into a particular host cell.

Glycosyltransferase Fusion Constructs

[0283] Similarly, a glycosyltransferase combinatorial DNA library was made
15 using the methods of the invention. A combinatorial DNA library of sequences derived from glycosyltransferase I (GnTI) activities were assembled with targeting peptides and screened for efficient production in a lower eukaryotic host cell of a $\text{GlcNAcMan}_5\text{GlcNAc}_2$ N-glycan structure on a marker glycoprotein. A fusion construct shown to produce $\text{GlcNAcMan}_5\text{GlcNAc}_2$ (**pPB104**), *Saccharomyces*
20 *MNN9*(s)/human GnTI Δ 38 was identified (**Example 8**). A wide variety of such GnTI fusion constructs were assembled (**Example 8, Table 10**). Other combinations of targeting peptide/GnTI catalytic domains can readily be assembled by making a combinatorial DNA library. It is also apparent to one skilled in the art that other such fusion constructs exhibiting glycosyltransferase
25 activity may be made as demonstrated in **Example 8**. It will be a matter of routine experimentation for one skilled in the art to use the combinatorial DNA library method described herein to optimize $\text{GlcNAcMan}_5\text{GlcNAc}_2$ production using a selected fusion construct in a particular expression vector and host cell line.
[0284] As stated above for mannosidase fusion constructs, not all targeting
30 peptide/GnTI catalytic domain fusion constructs will function equally well to produce the proper glycosylation on a glycoprotein of interest as described herein. However, one skilled in the art will be able to produce and select optimal fusion

construct(s) using a DNA library approach as described herein. **Example 8** illustrates a preferred embodiment of a combinatorial DNA library comprising targeting peptides and GnTI catalytic domain fusion constructs involved in producing glycoproteins with predominantly GlcNAcMan₅GlcNAc₂ structure.

5

Using Multiple Fusion Constructs to Alter Host Cell Glycosylation

[0285] In another example of using the methods and libraries of the invention to alter host cell glycosylation, a *P. pastoris* strain with an *OCH1* deletion that expresses a reporter protein (K3) was transformed with multiple fusion constructs isolated from combinatorial libraries of the invention to convert high mannose *N*-glycans to human-like *N*-glycans (**Example 8**). First, the mannosidase fusion construct pFB8 (*Saccharomyces SEC12* (m)/mouse mannosidase IA Δ187) was transformed into a *P. pastoris* strain lacking 1,6 initiating mannosyltransferases activity (*i.e.*, *och1* deletion; **Example 1**). Second, pPB103 comprising a *K. lactis* MNN2-2 gene (Genbank AN AF106080) encoding an UDP-GlcNAc transporter was constructed to increase further production of GlcNAcMan₅GlcNAc₂. The addition of the UDP-GlcNAc transporter increased production of GlcNAcMan₅GlcNAc₂ significantly in the *P. pastoris* strain as illustrated in **Figure 10B**. Third, pPB104 comprising *Saccharomyces MNN9* (s)/human GnTI Δ38 was introduced into the strain. This *P. pastoris* strain is referred to as “PBP-3.” (See **Figure 36**.)

[0286] It is understood by one skilled in the art that host cells such as the above-described yeast strains can be sequentially transformed and/or co-transformed with one or more expression vectors. It is also understood that the order of transformation is not particularly relevant in producing the glycoprotein of interest. The skilled artisan recognizes the routine modifications of the procedures disclosed herein may provide improved results in the production of the glycoprotein of interest.

[0287] The importance of using a particular targeting peptide sequence with a particular catalytic domain sequence becomes readily apparent from the experiments described herein. The combinatorial DNA library provides a tool for constructing enzyme fusions that are involved in modifying *N*-glycans on a

glycoprotein of interest, which is especially useful in producing human-like glycoproteins. (Any enzyme fusion, however, may be selected using libraries and methods of the invention.) Desired transformants expressing appropriately targeted, active α -1,2-mannosidase produce K3 with N-glycans of the structure

5 $\text{Man}_5\text{GlcNAc}_2$ as shown in Figures 5D and 5E. This confers a reduced molecular mass to the cleaved glycan compared to the K3 of the parent *OCH1* deletion strain, as was detected by MALDI-TOF mass spectrometry in Figure 5C.

[0288] Similarly, the same approach was used to produce another secreted glycoprotein: IFN- β comprising predominantly $\text{Man}_5\text{GlcNAc}_2$. The 10 $\text{Man}_5\text{GlcNAc}_2$ was removed by PNGase digestion (Papac *et al.* (1998) *Glycobiology* 8:445-454) and subjected to MALDI-TOF as shown in Figures 6A – 6F. A single prominent peak at 1254 (m/z) confirms $\text{Man}_5\text{GlcNAc}_2$ production on IFN- β in Figures 6E (pGCS) (*Saccharomyces MNS1*(m)/mouse mannosidase IB $\Delta 99$) and 6F (pFE8) (*Saccharomyces SEC12* (m)/mouse mannosidase IA $\Delta 187$). 15 Furthermore, in the *P. pastoris* strain PBP-3 comprising pFB8 (*Saccharomyces SEC12* (m)/mouse mannosidase IA $\Delta 187$), pPB104 (*Saccharomyces MNN9* (s)/human GnTI $\Delta 38$) and pPB103 (*K. lactis MNN2-2* gene), the hybrid N-glycan GlcNAcMan₅GlcNAc₂ [b] was detected by MALDI-TOF (Figure 10).

[0289] After identifying transformants with a high degree of mannose trimming, 20 additional experiments were performed to confirm that mannosidase (trimming) activity occurred *in vivo* and was not predominantly the result of extracellular activity in the growth medium (Example 6; Figures 7-9).

[0290] Although the present invention is exemplified using a *P. pastoris* host organism, it is understood by those skilled in the art that other eukaryotic host 25 cells, including other species of yeast and fungal hosts, may be altered as described herein to produce human-like glycoproteins. The techniques described herein for identification and disruption of undesirable host cell glycosylation genes, *e.g.* *OCH1*, is understood to be applicable for these and/or other homologous or functionally related genes in other eukaryotic host cells such as other yeast and 30 fungal strains. As described in Example 9, *och1 mnn1* genes were deleted from *K. lactis* to engineer a host cell leading to N-glycans that are completely converted to Man₅GlcNAc₂ by 1,2-mannosidase (Figure 12C).

[0291] The *MNN1* gene was cloned from *K. lactis* as described in Example 9. The nucleic acid and deduced amino acid sequences of the *K. lactis MNN1* gene are shown in SEQ ID NOs:43 and 44, respectively. Using gene-specific primers, a construct was made to delete the *MNN1* gene from the genome of *K. lactis*

5 (Example 9). Host cells depleted in *och1* and *mnn1* activities produce *N*-glycans having a Man₉GlcNAc₂ carbohydrate structure (see, e.g., Figure 12B). Such host cells may be engineered further using, e.g., methods and libraries of the invention, to produce mammalian- or human-like glycoproteins.

[0292] Thus, in another embodiment, the invention provides an isolated nucleic acid molecule having a nucleic acid sequence comprising or consisting of at least forty-five, preferably at least 50, more preferably at least 60 and most preferably 75 or more nucleotide residues of the *K. lactis MNN1* gene (SEQ ID NO: 43), and homologs, variants and derivatives thereof. The invention also provides nucleic acid molecules that hybridize under stringent conditions to the above-described 10 nucleic acid molecules. Similarly, isolated polypeptides (including muteins, allelic variants, fragments, derivatives, and analogs) encoded by the nucleic acid molecules of the invention are provided. In addition, also provided are vectors, including expression vectors, which comprise a nucleic acid molecule of the invention, as described further herein. Similarly host cells transformed with the 15 nucleic acid molecules or vectors of the invention are provided.

[0293] Another aspect of the present invention thus relates to a non-human eukaryotic host strain expressing glycoproteins comprising modified *N*-glycans that resemble those made by human-cells. Performing the methods of the invention in species other than yeast and fungal cells is thus contemplated and 20 encompassed by this invention. It is contemplated that a combinatorial nucleic acid library of the present invention may be used to select constructs that modify the glycosylation pathway in any eukaryotic host cell system. For example, the combinatorial libraries of the invention may also be used in plants, algae and insects, and in other eukaryotic host cells, including mammalian and human cells, 25 to localize proteins, including glycosylation enzymes or catalytic domains thereof, in a desired location along a host cell secretory pathway. Preferably, glycosylation enzymes or catalytic domains and the like are targeted to a subcellular location

along the host cell secretory pathway where they are capable of functioning, and preferably, where they are designed or selected to function most efficiently.

[0294] Plant and insect cells may also be engineered to alter the glycosylation of expressed proteins using the combinatorial library and methods of the invention.

5 Furthermore, glycosylation in mammalian cells, including human cells, may also be modified using the combinatorial library and methods of the invention. It may be possible, for example, to optimize a particular enzymatic activity or to otherwise modify the relative proportions of various *N*-glycans made in a mammalian host cell using the combinatorial library and methods of the invention.

10 [0295] Examples of modifications to glycosylation which can be affected using a method according to this embodiment of the invention are: (1) engineering a eukaryotic host cell to trim mannose residues from $\text{Man}_8\text{GlcNAc}_2$ to yield a $\text{Man}_5\text{GlcNAc}_2$ *N*-glycan; (2) engineering eukaryotic host cell to add an *N*-acetylglucosamine (GlcNAc) residue to $\text{Man}_5\text{GlcNAc}_2$ by action of GlcNAc

15 transferase I; (3) engineering a eukaryotic host cell to functionally express an enzyme such as an *N*-acetylglucosaminyl Transferase (GnTI, GnTII, GnTIII, GnTIV, GnTV, GnTVI), mannosidase II, fucosyltransferase (FT), galactosyl transferase (GalT) or a sialyltransferase (ST).

[0296] By repeating the method, increasingly complex glycosylation pathways 20 can be engineered into a target host, such as a lower eukaryotic microorganism. In one preferred embodiment, the host organism is transformed two or more times with DNA libraries including sequences encoding glycosylation activities.

Selection of desired phenotypes may be performed after each round of transformation or alternatively after several transformations have occurred.

25 Complex glycosylation pathways can be rapidly engineered in this manner.

Sequential Glycosylation Reactions

[0297] In a preferred embodiment, such targeting peptide/catalytic domain 30 libraries are designed to incorporate existing information on the sequential nature of glycosylation reactions in higher eukaryotes. Reactions known to occur early in the course of glycoprotein processing require the targeting of enzymes that catalyze such reactions to an early part of the Golgi or the ER. For example, the

trimming of $\text{Man}_8\text{GlcNAc}_2$ to $\text{Man}_5\text{GlcNAc}_2$ by mannosidases is an early step in complex *N*-glycan formation (Figures 1B and 35A). Because protein processing is initiated in the ER and then proceeds through the early, medial and late Golgi, it is desirable to have this reaction occur in the ER or early Golgi. When designing a 5 library for mannosidase I localization, for example, one thus attempts to match ER and early Golgi targeting signals with the catalytic domain of mannosidase I.

Generating Additional Sequence Diversity

[0298] The method of this embodiment is most effective when a nucleic acid, 10 *e.g.*, a DNA library transformed into the host contains a large diversity of sequences, thereby increasing the probability that at least one transformant will exhibit the desired phenotype. Single amino acid mutations, for example, may drastically alter the activity of glycoprotein processing enzymes (Romero *et al.* (2000) *J. Biol. Chem.* 275(15):11071-4). Accordingly, prior to transformation, a 15 DNA library or a constituent sub-library may be subjected to one or more techniques to generate additional sequence diversity. For example, one or more rounds of gene shuffling, error prone PCR, *in vitro* mutagenesis or other methods for generating sequence diversity, may be performed to obtain a larger diversity of sequences within the pool of fusion constructs.

20

Expression Control Sequences

[0299] In addition to the open reading frame sequences described above, it is generally preferable to provide each library construct with expression control sequences, such as promoters, transcription terminators, enhancers, ribosome 25 binding sites, and other functional sequences as may be necessary to ensure effective transcription and translation of the fusion proteins upon transformation of fusion constructs into the host organism.

[0300] Suitable vector components, *e.g.*, selectable markers, expression control sequences (*e.g.*, promoter, enhancers, terminators and the like) and, optionally, 30 sequences required for autonomous replication in a host cell, are selected as a function of which particular host cell is chosen. Selection criteria for suitable vector components for use in a particular mammalian or a lower eukaryotic host

cell are routine. Preferred lower eukaryotic host cells of the invention include *Pichia pastoris*, *Pichia finlandica*, *Pichia trehalophila*, *Pichia koclamae*, *Pichia membranaefaciens*, *Pichia opuntiae*, *Pichia thermotolerans*, *Pichia salictaria*, *Pichia guercuum*, *Pichia pijperi*, *Pichia stiptis*, *Pichia methanolica*, *Pichia* sp., 5 *Saccharomyces cerevisiae*, *Saccharomyces* sp., *Hansenula polymorpha*, *Kluyveromyces* sp., *Kluyveromyces lactis*, *Candida albicans*, *Aspergillus nidulans*, *Aspergillus niger*, *Aspergillus oryzae*, *Trichoderma reesei*, *Chrysosporium lucknowense*, *Fusarium* sp. *Fusarium gramineum*, *Fusarium venenatum* and *Neurospora crassa*. Where the host is *Pichia pastoris*, suitable promoters include, 10 for example, the *AOX1*, *AOX2*, *GAPDH* and *P40* promoters.

Selectable Markers

[0301] It is also preferable to provide each construct with at least one selectable marker, such as a gene to impart drug resistance or to complement a host metabolic 15 lesion. The presence of the marker is useful in the subsequent selection of transformants; for example, in yeast the *URA3*, *HIS4*, *SUC2*, *G418*, *BLA*, or *SH BLE* genes may be used. A multitude of selectable markers are known and available for use in yeast, fungi, plant, insect, mammalian and other eukaryotic host cells.

20

Transformation

[0302] The nucleic acid library is then transformed into the host organism. In yeast, any convenient method of DNA transfer may be used, such as electroporation, the lithium chloride method, or the spheroplast method. In 25 filamentous fungi and plant cells, conventional methods include particle bombardment, electroporation and agrobacterium mediated transformation. To produce a stable strain suitable for high-density culture (e.g., fermentation in yeast), it is desirable to integrate the DNA library constructs into the host chromosome. In a preferred embodiment, integration occurs via homologous 30 recombination, using techniques well-known in the art. For example, DNA library elements are provided with flanking sequences homologous to sequences of the

host organism. In this manner, integration occurs at a defined site in the host genome, without disruption of desirable or essential genes.

[0303] In an especially preferred embodiment, library DNA is integrated into the site of an undesired gene in a host chromosome, effecting the disruption or deletion 5 of the gene. For example, integration into the sites of the *OCH1*, *MNN1*, or *MNN4* genes allows the expression of the desired library DNA while preventing the expression of enzymes involved in yeast hypermannosylation of glycoproteins. In other embodiments, library DNA may be introduced into the host via a nucleic acid molecule, plasmid, vector (*e.g.*, viral or retroviral vector), chromosome, and may 10 be introduced as an autonomous nucleic acid molecule or by homologous or random integration into the host genome. In any case, it is generally desirable to include with each library DNA construct at least one selectable marker gene to allow ready selection of host organisms that have been stably transformed. Recyclable marker genes such as *ura3*, which can be selected for or against, are 15 especially suitable.

Screening and Selection Processes

[0304] After transformation of the host strain with the DNA library, transformants displaying a desired glycosylation phenotype are selected. Selection 20 may be performed in a single step or by a series of phenotypic enrichment and/or depletion steps using any of a variety of assays or detection methods. Phenotypic characterization may be carried out manually or using automated high-throughput screening equipment. Commonly, a host microorganism displays protein *N*-glycans on the cell surface, where various glycoproteins are localized.

[0305] One may screen for those cells that have the highest concentration of terminal GlcNAc on the cell surface, for example, or for those cells which secrete 25 the protein with the highest terminal GlcNAc content. Such a screen may be based on a visual method, like a staining procedure, the ability to bind specific terminal GlcNAc binding antibodies or lectins conjugated to a marker (such lectins are available from E.Y. Laboratories Inc., San Mateo, CA), the reduced ability of 30 specific lectins to bind to terminal mannose residues, the ability to incorporate a radioactively labeled sugar *in vitro*, altered binding to dyes or charged surfaces, or

may be accomplished by using a Fluorescence Assisted Cell Sorting (FACS) device in conjunction with a fluorophore labeled lectin or antibody (Guillen *et al.* (1998) *Proc. Natl. Acad. Sci. USA* 95(14):7888-7892).

[0306] Accordingly, intact cells may be screened for a desired glycosylation phenotype by exposing the cells to a lectin or antibody that binds specifically to the desired *N*-glycan. A wide variety of oligosaccharide-specific lectins are available commercially (*e.g.*, from EY Laboratories, San Mateo, CA). Alternatively, antibodies to specific human or animal *N*-glycans are available commercially or may be produced using standard techniques. An appropriate lectin or antibody 10 may be conjugated to a reporter molecule, such as a chromophore, fluorophore, radioisotope, or an enzyme having a chromogenic substrate (Guillen *et al.*, 1998. *Proc. Natl. Acad. Sci. USA* 95(14): 7888-7892).

[0307] Screening may then be performed using analytical methods such as spectrophotometry, fluorimetry, fluorescence activated cell sorting, or scintillation 15 counting. In other cases, it may be necessary to analyze isolated glycoproteins or *N*-glycans from transformed cells. Protein isolation may be carried out by techniques known in the art. In a preferred embodiment, a reporter protein is secreted into the medium and purified by affinity chromatography (*e.g.* Ni-affinity or glutathione -S-transferase affinity chromatography). In cases where an isolated 20 *N*-glycan is preferred, an enzyme such as endo- β -*N*-acetylglucosaminidase (Genzyme Co., Boston, MA; New England Biolabs, Beverly, MA) may be used to cleave the *N*-glycans from glycoproteins. Isolated proteins or *N*-glycans may then be analyzed by liquid chromatography (*e.g.*, HPLC), mass spectroscopy, or other suitable means. U.S. Patent No. 5,595,900 teaches several methods by which cells 25 with desired extracellular carbohydrate structures may be identified. In a preferred embodiment, MALDI-TOF mass spectrometry is used to analyze the cleaved *N*-glycans.

[0308] Prior to selection of a desired transformant, it may be desirable to deplete the transformed population of cells having undesired phenotypes. For example, 30 when the method is used to engineer a functional mannosidase activity into cells, the desired transformants will have lower levels of mannose in cellular glycoprotein. Exposing the transformed population to a lethal radioisotope of

mannose in the medium depletes the population of transformants having the undesired phenotype, *i.e.*, high levels of incorporated mannose (Huffaker and Robbins (1983) *Proc Natl Acad Sci U S A.* 80(24):7466-70). Alternatively, a cytotoxic lectin or antibody, directed against an undesirable *N*-glycan, may be used

5 to deplete a transformed population of undesired phenotypes (*e.g.*, Stanley and Siminovitch (1977) *Somatic Cell Genet* 3(4):391-405). U.S. Patent No. 5,595,900 teaches several methods by which cells with a desired extracellular carbohydrate structures may be identified. Repeatedly carrying out this strategy allows for the sequential engineering of more and more complex glycans in lower eukaryotes.

10 [0309] To detect host cells having on their surface a high degree of the human-like *N*-glycan intermediate GlcNAcMan₃GlcNAc₂, for example, one may select for transformants that allow for the most efficient transfer of GlcNAc by GlcNAc Transferase from UDP-GlcNAc in an *in vitro* cell assay. This screen may be carried out by growing cells harboring the transformed library under selective

15 pressure on an agar plate and transferring individual colonies into a 96-well microtiter plate. After growing the cells, the cells are centrifuged, the cells resuspended in buffer, and after addition of UDP-GlcNAc and GnTII, the release of UDP is determined either by HPLC or an enzyme linked assay for UDP. Alternatively, one may use radioactively labeled UDP-GlcNAc and GnTII, wash

20 the cells and then look for the release of radioactive GlcNAc by *N*-acetylglucosaminidase. All this may be carried manually or automated through the use of high throughput screening equipment. Transformants that release more UDP, in the first assay, or more radioactively labeled GlcNAc in the second assay, are expected to have a higher degree of GlcNAcMan₃GlcNAc₂ on their surface and

25 thus constitute the desired phenotype. Similar assays may be adapted to look at the *N*-glycans on secreted proteins as well.

[0310] Alternatively, one may use any other suitable screen such as a lectin binding assay that is able to reveal altered glycosylation patterns on the surface of transformed cells. In this case the reduced binding of lectins specific to terminal mannosides may be a suitable selection tool. *Galanthus nivalis* lectin binds specifically to terminal α -1,3 mannose, which is expected to be reduced if sufficient mannosidase II activity is present in the Golgi. One may also enrich for

desired transformants by carrying out a chromatographic separation step that allows for the removal of cells containing a high terminal mannose content. This separation step would be carried out with a lectin column that specifically binds cells with a high terminal mannose content (e.g., *Galanthus nivalis* lectin bound to 5 agarose, Sigma, St.Louis, MO) over those that have a low terminal mannose content.

[0311] In addition, one may directly create such fusion protein constructs, as additional information on the localization of active carbohydrate modifying enzymes in different lower eukaryotic hosts becomes available in the scientific 10 literature. For example, it is known that human β 1,4-GalTr can be fused to the membrane domain of *MNT*, a mannosyltransferase from *S. cerevisiae*, and localized to the Golgi apparatus while retaining its catalytic activity (Schwientek *et al.* (1995) *J. Biol. Chem.* 270(10):5483-9). If *S. cerevisiae* or a related organism is 15 the host to be engineered one may directly incorporate such findings into the overall strategy to obtain complex *N*-glycans from such a host. Several such gene fragments in *P. pastoris* have been identified that are related to glycosyltransferases in *S. cerevisiae* and thus could be used for that purpose.

Integration Sites

20 [0312] As one ultimate goal of this genetic engineering effort is a robust protein production strain that is able to perform well in an industrial fermentation process, the integration of multiple genes into the host (e.g., fungal) chromosome preferably involves careful planning. The engineered strain may likely have to be transformed with a range of different genes, and these genes will have to be 25 transformed in a stable fashion to ensure that the desired activity is maintained throughout the fermentation process. As described herein, any combination of various desired enzyme activities may be engineered into the fungal protein expression host, e.g., sialyltransferases, mannosidases, fucosyltransferases, galactosyltransferases, glucosyltransferases, GlcNAc transferases, ER and Golgi 30 specific transporters (e.g. syn and antiport transporters for UDP-galactose and other precursors), other enzymes involved in the processing of oligosaccharides, and enzymes involved in the synthesis of activated oligosaccharide precursors such

as UDP-galactose, CMP-N-acetylneurameric acid. Examples of preferred methods for modifying glycosylation in a lower eukaryotic host cell, such as *Pichia pastoris*, are shown in Table 6.

5 Table 6. Some preferred embodiments for modifying glycosylation in a lower eukaryotic microorganism

Desired Structure	Suitable Catalytic Activities	Suitable Sources of Localization Sequences	Suitable Gene Deletions	Suitable Transporters and/or Phosphatases
Man ₅ GlcNAc ₂	α-1,2-mannosidase (murine, human, <i>Bacillus</i> sp., <i>A.nidulans</i>)	Mns1 (<i>N</i> -terminus, <i>S. cerevisiae</i>) Och1 (<i>N</i> -terminus, <i>S. cerevisiae</i> , <i>P. pastoris</i>) Ktr1 Mnn9 Mnt1 (<i>S. cerevisiae</i>) KDEL, HDEL (<i>C</i> -terminus)	<i>OCH1</i> <i>MNN4</i> <i>MNN6</i>	none
GlcNAcMan ₅ GlcNAc ₂	GlcNAc Transferase I, (human, murine, rat etc.)	Och1 (<i>N</i> -terminus, <i>S. cerevisiae</i> , <i>P. pastoris</i>) KTR1 (<i>N</i> -terminus) Mnn1 (<i>N</i> -terminus, <i>S. cerevisiae</i>) Mnt1 (<i>N</i> -terminus, <i>S. cerevisiae</i>) GDPase (<i>N</i> -terminus, <i>S. cerevisiae</i>)	<i>OCH1</i> <i>MNN4</i> <i>MNN6</i>	UDP-GlcNAc transporter (human, murine, <i>K. lactis</i>) UDPase (human)
GlcNAcMan ₃ GlcNAc ₂	mannosidase II	Ktr1 Mnn1 (<i>N</i> -terminus, <i>S. cerevisiae</i>) Mnt1 (<i>N</i> -terminus, <i>S. cerevisiae</i>) Kre2/Mnt1 (<i>S. cerevisiae</i>) Kre2 (<i>P. pastoris</i>) Ktr1 (<i>S. cerevisiae</i>) Ktr1 (<i>P. pastoris</i>) Mnn1 (<i>S. cerevisiae</i>)	<i>OCH1</i> <i>MNN4</i> <i>MNN6</i>	UDP-GlcNAc transporter (human, murine, <i>K. lactis</i>) UDPase (human)
GlcNAc _{(2-4)‑} Man ₃ GlcNAc ₂	GlcNAc Transferase II, III, IV, V	Mnn1 (<i>N</i> -terminus, <i>S. cerevisiae</i>) Mnt1 (<i>N</i> -terminus,	<i>OCH1</i> <i>MNN4</i> <i>MNN6</i>	UDP-GlcNAc transporter (human, murine,

	(human, murine)	<i>S. cerevisiae</i>) Kre2/Mnt1 (<i>S.</i> <i>cerevisiae</i>) Kre2 (<i>P. pastoris</i>) Ktr1 (<i>S. cerevisiae</i>) Ktr1 (<i>P. pastoris</i>) Mnn1 (<i>S.</i> <i>cerevisiae</i>)		<i>K. lactis</i>) UDPase (human)
Gal ₍₁₋₄₎ GlcNAc ₍₂₋₄₎ - Man ₃ GlcNAc ₂	β-1,4- Galactosyl transferase (human)	Mnn1 (<i>N</i> -terminus, <i>S. cerevisiae</i>) Mnt1 (<i>N</i> -terminus, <i>S.</i> <i>cerevisiae</i>) Kre2/Mnt1 (<i>S.</i> <i>cerevisiae</i>) Kre2 (<i>P. pastoris</i>) Ktr1 (<i>S. cerevisiae</i>) Ktr1 (<i>P. pastoris</i>) Mnn1 (<i>S.</i> <i>cerevisiae</i>)	<i>OCH1</i> <i>MNN4</i> <i>MNN6</i>	UDP-Galactose transporter (human, <i>S.pombe</i>)
NANA ₍₁₋₄₎ - Gal ₍₁₋₄₎ GlcNAc ₍₂₋₄₎ - Man ₃ GlcNAc ₂	α-2,6- Sialyltransfe rase (human) α-2,3- Sialyltransfe rase	KTR1 MNN1 (<i>N</i> -terminus, <i>S. cerevisiae</i>) MNT1 (<i>N</i> -terminus, <i>S. cerevisiae</i>) Kre2/Mnt1 (<i>S.</i> <i>cerevisiae</i>) Kre2 (<i>P. pastoris</i>) Ktr1 (<i>S. cerevisiae</i>) Ktr1 (<i>P. pastoris</i>) MNN1 (<i>S.</i> <i>cerevisiae</i>)	<i>OCH1</i> <i>MNN4</i> <i>MNN6</i>	CMP-Sialic acid transporter (human)

[0313] As any strategy to engineer the formation of complex *N*-glycans into a host cell such as a lower eukaryote involves both the elimination as well as the addition of particular glycosyltransferase activities, a comprehensive scheme will attempt to coordinate both requirements. Genes that encode enzymes that are undesirable serve as potential integration sites for genes that are desirable. For example, 1,6 mannosyltransferase activity is a hallmark of glycosylation in many known lower eukaryotes. The gene encoding alpha-1,6 mannosyltransferase (*OCH1*) has been cloned from *S. cerevisiae* and mutations in the gene give rise to a viable phenotype with reduced mannosylation. The gene locus encoding alpha-1,6 mannosyltransferase activity therefore is a prime target for the integration of genes encoding glycosyltransferase activity. In a similar manner, one can choose a range

of other chromosomal integration sites that, based on a gene disruption event in that locus, are expected to: (1) improve the cells ability to glycosylate in a more human-like fashion, (2) improve the cells ability to secrete proteins, (3) reduce proteolysis of foreign proteins and (4) improve other characteristics of the process
5 that facilitate purification or the fermentation process itself.

Target Glycoproteins

[0314] The methods described herein are useful for producing glycoproteins, especially glycoproteins used therapeutically in humans. Glycoproteins having
10 specific glycoforms may be especially useful, for example, in the targeting of therapeutic proteins. For example, mannose-6-phosphate has been shown to direct proteins to the lysosome, which may be essential for the proper function of several enzymes related to lysosomal storage disorders such as Gaucher's, Hunter's, Hurler's, Scheie's, Fabry's and Tay-Sachs disease, to mention just a few.
15 Likewise, the addition of one or more sialic acid residues to a glycan side chain may increase the lifetime of a therapeutic glycoprotein *in vivo* after administration. Accordingly, host cells (e.g., lower eukaryotic or mammalian) may be genetically engineered to increase the extent of terminal sialic acid in glycoproteins expressed in the cells. Alternatively, sialic acid may be conjugated to the protein of interest
20 *in vitro* prior to administration using a sialic acid transferase and an appropriate substrate. Changes in growth medium composition may be employed in addition to the expression of enzyme activities involved in human-like glycosylation to produce glycoproteins more closely resembling human forms (Weikert *et al.*
25 (1999) *Nature Biotechnology* 17, 1116-1121; Werner *et al.* (1998)
27 *Arzneimittelforschung* 48(8):870-880; Andersen and Goochee (1994) *Cur. Opin. Biotechnol.* 5:546-549; Yang and Butler (2000) *Biotechnol.Bioengin.* 68(4):370-380). Specific glycan modifications to monoclonal antibodies (e.g. the addition of a bisecting GlcNAc) have been shown to improve antibody dependent cell
30 cytotoxicity (Umana *et al.* (1999) *Nat. Biotechnol.* 17(2):176-80), which may be desirable for the production of antibodies or other therapeutic proteins.
[0315] Therapeutic proteins are typically administered by injection, orally, pulmonary, or other means. Examples of suitable target glycoproteins which may

be produced according to the invention include, without limitation: erythropoietin, cytokines such as interferon- α , interferon- β , interferon- γ , interferon- ω , and granulocyte-CSF, coagulation factors such as factor VIII, factor IX, and human protein C, soluble IgE receptor α -chain, IgG, IgG fragments, IgM, interleukins, 5 urokinase, chymase, and urea trypsin inhibitor, IGF-binding protein, epidermal growth factor, growth hormone-releasing factor, annexin V fusion protein, angiostatin, vascular endothelial growth factor-2, myeloid progenitor inhibitory factor-1, osteoprotegerin, α -1 antitrypsin, DNase II, α -feto proteins, AAT, rhTBP-1 (onercept, aka TNF Binding protein 1), TACI-Ig (transmembrane activator and 10 calcium modulator and cyclophilin ligand interactor), FSH (follicle stimulating hormone), GM-CSF, GLP-1 w/ and w/o FC (glucagon like protein 1) IL-1 receptor agonist, sTNFr (enbrel, aka soluble TNF receptor Fc fusion) ATIII, rhThrombin, glucocerebrosidase and CTLA4-Ig (Cytotoxic T Lymphocyte associated Antigen 4 - Ig).

15

Expression Of GnT-III To Boost Antibody Functionality

[0316] The addition of *N*-acetylglucosamine residues to the GlcNAcMan₃GlcNAc₂ structure by *N*-acetylglucosaminyltransferases II and III yields a so-called bisected *N*-glycan GlcNAc₃Man₃GlcNAc₂ (Figure 15). This 20 structure has been implicated in greater antibody-dependent cellular cytotoxicity (ADCC) (Umana *et al.* (1999) *Nat. Biotechnol.* 17(2):176-80). Re-engineering glycoforms of immunoglobulins expressed by mammalian cells is a tedious and cumbersome task. Especially in the case of GnTIII, where over-expression of this enzyme has been implicated in growth inhibition, methods involving regulated 25 (inducible) gene expression had to be employed to produce immunoglobulins with bisected *N*-glycans (Umana *et al.* (1999) *Biotechnol Bioeng.* 65(5):542-9; Umana *et al.* (1999) *Nat. Biotechnol.* 17(2):176-80); Umana *et al.* WO 03/011878; U.S. Patent No. 6,602,684.

[0317] Accordingly, in another embodiment, the invention provides systems and 30 methods for producing human-like *N*-glycans having bisecting *N*-acetylglucosamine (GlcNAc) on a trimannose or pentamannose core structure. In a preferred embodiment, the invention provides a system and method for producing

immunoglobulins having bisected *N*-glycans. The systems and methods described herein will not suffer from previous problems, *e.g.*, cytotoxicity associated with overexpression of GnTIII or ADCC, as the host cells of the invention are engineered and selected to be viable and preferably robust cells which produce *N*-glycans having substantially modified human-type glycoforms such as GlcNAc₂Man₃GlcNAc₂. Thus, addition of a bisecting *N*-acetylglucosamine in a host cell of the invention will have a negligible effect on the growth-phenotype or viability of those host cells.

[0318] In addition, work by others has shown that there is no linear correlation between GnTIII expression levels and the degree of ADCC. Umana *et al.* (1999) *Nature Biotechnol.* 17:176-80. Thus, finding the optimal expression level in mammalian cells and maintaining it throughout an FDA approved fermentation process seems to be a challenge. However, in cells of the invention, such as fungal cells, finding a promoter of appropriate strength to establish a robust, reliable and optimal GnTIII expression level is a comparatively easy task for one of skill in the art.

[0319] A host cell such as a yeast strain capable of producing glycoproteins with bisecting *N*-glycans is engineered according to the invention, by introducing into the host cell a GnTIII activity (**Example 12**). Preferably, the host cell is transformed with a nucleic acid that encodes GnTIII (*see, e.g., Figure 24*) or a domain thereof having enzymatic activity, optionally fused to a heterologous cell signal targeting peptide (*e.g.*, using the libraries and associated methods of the invention.) Host cells engineered to express GnTIII will produce higher antibody titers than mammalian cells are capable of. They will also produce antibodies with higher potency with respect to ADCC.

[0320] Antibodies produced by mammalian cell lines transfected with GnTIII have been shown to be as effective as antibodies produced by non-transfected cell-lines, but at a 10-20 fold lower concentration (Davies *et al.* (2001) *Biotechnol. Bioeng.* 74(4):288-94). An increase of productivity of the production vehicle of the invention over mammalian systems by a factor of twenty, and a ten-fold increase of potency will result in a net-productivity improvement of two hundred. The invention thus provides a system and method for producing high titers of an

antibody having high potency (e.g., up to several orders of magnitude more potent than what can currently be produced). The system and method is safe and provides high potency antibodies at low cost in short periods of time. Host cells engineered to express GnTIII according to the invention produce immunoglobulins having bisected N-glycans at rates of at least 50 mg/liter/day to at least 500 mg/liter/day. In addition, each immunoglobulin (Ig) molecule (comprising bisecting GlcNAcs) is more potent than the same Ig molecule produced without bisecting GlcNAcs.

10 **Production of Multiantennary Structures For Improved Glycoprotein Functionality**

[0321] Synthesis of tetraantennary structures has been found to be important for *in vivo* biological activity of a variety of proteins such as EPO and α_1 -acid glycoproteins. Takeuchi et al., Proc Natl Acad Sci U S A. 1989 Oct;86(20):7819-22; Boris et al., Inflammation (1990) 14, 315-323. Pharmacokinetics studies have shown that the bulky structure of the tetraantennary branching prevents EPO from filtering out into the urine. Modification of proteins, for example, with chemical conjugates (e.g., polyethylene glycol), has been devised to retard the clearance of potentially therapeutic glycoproteins. Accordingly, in one embodiment, the present invention provides methods to synthesize glycoproteins comprising multiple antennary structures in lower eukaryotes (*P.pastoris*) that have better *in vivo* biological activity and are less rapidly cleared than the same glycoprotein having reduced antennarity. In essence, the glycoproteins produced according to the methods of the present invention (see also WO 02/00879 and WO 03/056914 incorporated herein by reference) have improved therapeutic efficacy.

20 [0322] The following are examples which illustrate various aspects of the invention. These examples should not be construed as limiting: the examples are included for the purposes of illustration only.

EXAMPLE 1
Cloning and Disruption of the *OCH1* gene in *P. pastoris*

Generation of an *OCH1* mutant of *P. pastoris*:

[0323] A 1215 bp ORF of the *P. pastoris* *OCH1* gene encoding a putative α -1,6-mannosyltransferase was amplified from *P. pastoris* genomic DNA (strain X-33, Invitrogen, Carlsbad, CA) using the oligonucleotides 5'-ATGGCGAAGGCAGATGGCAGT-3' (SEQ ID NO:3) and 5'-TTAGTCCTTCCAACTTCCCTTC-3' (SEQ ID NO:4) which were designed based on the *P. pastoris* *OCH1* sequence (Japanese Patent Application Publication No. 8-336387). Subsequently, 2685 bp upstream and 1175 bp downstream of the ORF of the *OCH1* gene were amplified from a *P. pastoris* genomic DNA library (Boehm, T. et al. (1999) *Yeast* 15(7):563-72) using the internal oligonucleotides 5'-ACTGCCATCTGCCTTCGCCAT-3' (SEQ ID NO:47) in the *OCH1* gene, and 5'-GTAATACGACTCACTATAAGGGC-3' T7 (SEQ ID NO:48) and 5'-AATTAACCCTCACTAAAGGG-3' T3 (SEQ ID NO:49) oligonucleotides in the backbone of the library bearing plasmid lambda ZAP II (Stratagene, La Jolla, CA). The resulting 5075 bp fragment was cloned into the pCR2.1-TOPO vector (Invitrogen, Carlsbad, CA) and designated pBK9.

[0324] After assembling a gene knockout construct that substituted the *OCH1* reading frame with a *HIS4* resistance gene, *P. pastoris* was transformed and colonies were screened for temperature sensitivity at 37°C. *OCH1* mutants of *S. cerevisiae* are temperature sensitive and are slow growers at elevated temperatures. One can thus identify functional homologs of *OCH1* in *P. pastoris* by complementing an *OCH1* mutant of *S. cerevisiae* with a *P. pastoris* DNA or cDNA library. About 20 temperature sensitive strains were further subjected to a colony PCR screen to identify colonies with a deleted *och1* gene. Several *och1* deletions were obtained.

[0325] The linearized pBK9.1, which has 2.1 kb upstream sequence and 1.5 kb downstream sequence of *OCH1* gene cassette carrying *Pichia HIS4* gene, was transformed into *P. pastoris* BK1 [GS115 (*his4* Invitrogen Corp., San Diego, CA) carrying the human IFN- β gene in the *AOX1* locus] to knock out the wild-type *OCH1* gene. The initial screening of transformants was performed using histidine

drop-out medium followed by replica plating to select the temperature sensitive colonies. Twenty out of two hundred histidine-positive colonies showed a temperature sensitive phenotype at 37°C. To exclude random integration of pBK9.1 into the *Pichia* genome, the 20 temperature-sensitive isolates were 5 subjected to colony PCR using primers specific to the upstream sequence of the integration site and to *HIS4* ORF. Two out of twenty colonies were *och1* defective and further analyzed using a Southern blot and a Western blot indicating the functional *och1* disruption by the *och1* knock-out construct. Genomic DNA were digested using two separate restriction enzymes *BglII* and *ClaI* to confirm the *och1* 10 knock-out and to confirm integration at the open reading frame. The Western Blot showed *och1* mutants lacking a discrete band produced in the GS115 wild type at 46.2 kDa.

EXAMPLE 2

15 **Engineering of *P. pastoris* with α -1,2-Mannosidase to Produce $\text{Man}_5\text{GlcNAc}_2$ - Containing IFN- β Precursors**

[0326] An α -1,2-mannosidase is required for the trimming of $\text{Man}_8\text{GlcNAc}_2$ to yield $\text{Man}_5\text{GlcNAc}_2$, an essential intermediate for complex *N*-glycan formation. While the production of a $\text{Man}_5\text{GlcNAc}_2$ precursor is essential, it is not necessarily 20 sufficient for the production of hybrid and complex glycans because the specific isomer of $\text{Man}_5\text{GlcNAc}_2$ may or may not be a substrate for GnTI. An *och1* mutant of *P. pastoris* is engineered to express secreted human interferon- β under the control of an *aox* promoter. A DNA library is constructed by the in-frame ligation of the catalytic domain of human mannosidase IB (an α -1,2-mannosidase) with a 25 sub-library including sequences encoding early Golgi and ER localization peptides. The DNA library is then transformed into the host organism, resulting in a genetically mixed population wherein individual transformants each express interferon- β as well as a synthetic mannosidase gene from the library. Individual transformant colonies are cultured and the production of interferon is induced by 30 addition of methanol. Under these conditions, over 90% of the secreted protein is glycosylated interferon- β .

[0327] Supernatants are purified to remove salts and low-molecular weight contaminants by C₁₈ silica reversed-phase chromatography. Desired transformants expressing appropriately targeted, active α -1,2-mannosidase produce interferon- β including N-glycans of the structure Man₅GlcNAc₂, which has a reduced molecular mass compared to the interferon- β of the parent strain. The purified interferon- β is analyzed by MALDI-TOF mass spectroscopy and colonies expressing the desired form of interferon- β are identified.

EXAMPLE 3

10 **Generation of an och1 Mutant Strain Expressing an α -1,2-Mannosidase, GnT I and GnT II for Production of a Human-Like Glycoprotein.**

[0328] The 1215 bp open reading frame of the *P. pastoris* *OCH1* gene as well as 2685 bp upstream and 1175 bp downstream was amplified by PCR (*see also* WO 02/00879), cloned into the pCR2.1-TOPO vector (Invitrogen) and designated pBK9. To create an *och1* knockout strain containing multiple auxotrophic markers, 100 μ g of pJN329, a plasmid containing an *och1::URA3* mutant allele flanked with *Sfi*I restriction sites was digested with *Sfi*I and used to transform *P. pastoris* strain JC308 (Cereghino *et al.* (2001) *Gene* 263:159-169) by electroporation. Following incubation on defined medium lacking uracil for 10 days at room temperature, 1000 colonies were picked and re-streaked. URA⁺ clones that were unable to grow at 37°C, but grew at room temperature, were subjected to colony PCR to test for the correct integration of the *och1::URA3* mutant allele. One clone that exhibited the expected PCR pattern was designated YJN153. The Kringle 3 domain of human plasminogen (K3) was used as a model protein. A Neo^R marked plasmid containing the K3 gene was transformed into strain YJN153 and a resulting strain, expressing K3, was named BK64-1.

[0329] Plasmid pPB103, containing the *Kluyveromyces lactis* *MNN2-2* gene which encodes a Golgi UDP-N-acetylglucosamine transporter was constructed by cloning a blunt *Bgl*II-*Hind*III fragment from vector pDL02 (Abeijon *et al.* (1996) *Proc. Natl. Acad. Sci. U.S.A.* 93:5963-5968) into *Bgl*II and *Bam*HI digested and blunt ended pBLADE-SX containing the *P. pastoris* *ADE1* gene (Cereghino *et al.* (2001) *Gene* 263:159-169). This plasmid was linearized with *Eco*NI and

transformed into strain BK64-1 by electroporation and one strain confirmed to contain the *MNN2-2* by PCR analysis was named **PBP1**.

[0330] A library of mannosidase constructs was generated, comprising in-frame fusions of the leader domains of several type I or type II membrane proteins from *S. cerevisiae* and *P. pastoris* fused with the catalytic domains of several α -1,2-mannosidase genes from human, mouse, fly, worm and yeast sources (see, e.g., WO02/00879, incorporated herein by reference). This library was created in a *P. pastoris* *HIS4* integration vector and screened by linearizing with *SalI*, transforming by electroporation into strain **PBP1**, and analyzing the glycans released from the K3 reporter protein. One active construct chosen was a chimera of the 988-1296 nucleotides (*C*-terminus) of the yeast *SEC12* gene fused with a *N*-terminal deletion of the mouse α -1,2-mannosidase IA gene (Figure 3), which was missing the 187 nucleotides. A *P. pastoris* strain expressing this construct was named **PBP2**.

10 [0331] A library of GnTI constructs was generated, comprising in-frame fusions of the same leader library with the catalytic domains of GnTI genes from human, worm, frog and fly sources (WO 02/00879). This library was created in a *P. pastoris* *ARG4* integration vector and screened by linearizing with *AatII*, transforming by electroporation into strain **PBP2**, and analyzing the glycans released from K3. One active construct chosen was a chimera of the first 120 bp of the *S. cerevisiae* *MNN9* gene fused to a deletion of the human GnTI gene, which was missing the first 154 bp. A *P. pastoris* strain expressing this construct was named **PBP-3**. (See also Figure 36.)

15 [0332] A library of GnTII constructs was generated, which comprised in-frame fusions of the leader library with the catalytic domains of GnTII genes from human and rat sources (WO 02/00879). This library was created in a *P. pastoris* integration vector containing the *NST^R* gene conferring resistance to the drug nourseothricin. The library plasmids were linearized with *EcoRI*, transformed into strain **RDP27** by electroporation, and the resulting strains were screened by analysis of the released glycans from purified K3.

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Materials for the Following Reactions

[0333] MOPS, sodium cacodylate, manganese chloride, UDP-galactose and CMP-N-acetylneuraminic acid were from Sigma. Trifluoroacetic acid (TFA) was from Sigma/Aldrich, Saint Louis, MO. Recombinant rat α 2,6-sialyltransferase 5 from *Spodoptera frugiperda* and β 1,4-galactosyltransferase from bovine milk were from Calbiochem (San Diego, CA). Protein N-glycosidase F, mannosidases, and oligosaccharides were from Glyko (San Rafael, CA). DEAE ToyoPearl resin was from TosoHaas. Metal chelating ‘HisBind’ resin was from Novagen (Madison, WI). 96-well lysate-clearing plates were from Promega (Madison, WI). Protein-10 binding 96-well plates were from Millipore (Bedford, MA). Salts and buffering agents were from Sigma (St. Louis, MO). MALDI matrices were from Aldrich (Milwaukee, WI).

Protein Purification

15 [0334] Kringle 3 was purified using a 96-well format on a Beckman BioMek 2000 sample-handling robot (Beckman/Coulter Ranch Cucamonga, CA). Kringle 3 was purified from expression media using a C-terminal hexa-histidine tag. The robotic purification is an adaptation of the protocol provided by Novagen for their HisBind resin. Briefly, a 150uL (μ L) settled volume of resin is poured into the 20 wells of a 96-well lysate-binding plate, washed with 3 volumes of water and charged with 5 volumes of 50mM NiSO₄ and washed with 3 volumes of binding buffer (5mM imidazole, 0.5M NaCl, 20mM Tris-HCL pH7.9). The protein expression media is diluted 3:2, media/PBS (60mM PO₄, 16mM KCl, 822mM NaCl pH7.4) and loaded onto the columns. After draining, the columns are 25 washed with 10 volumes of binding buffer and 6 volumes of wash buffer (30mM imidazole, 0.5M NaCl, 20mM Tris-HCl pH7.9) and the protein is eluted with 6 volumes of elution buffer (1M imidazole, 0.5M NaCl, 20mM Tris-HCl pH7.9). The eluted glycoproteins are evaporated to dryness by lyophilization.

30 Release of N-linked Glycans

[0335] The glycans are released and separated from the glycoproteins by a modification of a previously reported method (Papac, *et al.* A. J. S. (1998)

Glycobiology 8, 445-454). The wells of a 96-well MultiScreen IP (Immobilon-P membrane) plate (Millipore) are wetted with 100uL of methanol, washed with 3X150uL of water and 50uL of RCM buffer (8M urea, 360mM Tris, 3.2mM EDTA pH8.6), draining with gentle vacuum after each addition. The dried protein samples are dissolved in 30uL of RCM buffer and transferred to the wells containing 10uL of RCM buffer. The wells are drained and washed twice with RCM buffer. The proteins are reduced by addition of 60uL of 0.1M DTT in RCM buffer for 1hr at 37°C. The wells are washed three times with 300uL of water and carboxymethylated by addition of 60uL of 0.1M iodoacetic acid for 30min in the dark at room temperature. The wells are again washed three times with water and the membranes blocked by the addition of 100uL of 1% PVP 360 in water for 1hr at room temperature. The wells are drained and washed three times with 300uL of water and deglycosylated by the addition of 30uL of 10mM NH₄HCO₃ pH 8.3 containing one milliunit of *N*-glycanase (Glyko). After 16 hours at 37°C, the solution containing the glycans was removed by centrifugation and evaporated to dryness.

Matrix Assisted Laser Desorption Ionization Time of Flight Mass Spectrometry

[0336] Molecular weights of the glycans were determined using a Voyager DE PRO linear MALDI-TOF (Applied Biosciences) mass spectrometer using delayed extraction. The dried glycans from each well were dissolved in 15uL of water and 0.5uL spotted on stainless steel sample plates and mixed with 0.5uL of S-DHB matrix (9mg/mL of dihydroxybenzoic acid, 1mg/mL of 5-methoxysalicylic acid in 1:1 water/acetonitrile 0.1% TFA) and allowed to dry.

[0337] Ions were generated by irradiation with a pulsed nitrogen laser (337nm) with a 4ns pulse time. The instrument was operated in the delayed extraction mode with a 125ns delay and an accelerating voltage of 20kV. The grid voltage was 93.00%, guide wire voltage was 0.10%, the internal pressure was less than 5 X 10⁻⁷ torr, and the low mass gate was 875Da. Spectra were generated from the sum of 100-200 laser pulses and acquired with a 2 GHz digitizer. Man₅GlcNAc₂ oligosaccharide was used as an external molecular weight standard. All spectra

were generated with the instrument in the positive ion mode. The estimated mass accuracy of the spectra was 0.5%.

[0338] The mass of the *N*-glycans eluted from the column is generally associated with a positive ion adduct, which increases the mass by the molecular weight of the 5 positive ion. The most common adducts are H⁺, Na⁺ and K⁺.

EXAMPLE 4

Engineering of *P. pastoris* to Produce Man₅GlcNAc₂ as the Predominant *N*-Glycan Structure Using a Combinatorial DNA Library.

10 [0339] An *och1* mutant of *P. pastoris* (see Examples 1 and 3) was engineered to express and secrete proteins such as the kringle 3 domain of human plasminogen (K3) under the control of the inducible *AOX1* promoter. The Kringle 3 domain of human plasminogen (K3) was used as a model protein. A DNA fragment encoding the K3 was amplified using Pfu turbo polymerase (Stratagene, La Jolla, CA) and 15 cloned into *EcoRI* and *XbaI* sites of pPICZαA (Invitrogen, Carlsbad, CA), resulting in a C-terminal 6- His tag. In order to improve the *N*-linked glycosylation efficiency of K3 (Hayes *et al.* 1975 *J. Arch. Biochem. Biophys.* 171, 651-655), Pro₄₆ was replaced with Ser₄₆ using site-directed mutagenesis. The resulting plasmid was designated pBK64. The correct sequence of the PCR construct was 20 confirmed by DNA sequencing.

[0340] A combinatorial DNA library was constructed by the in-frame ligation of murine α-1,2-mannosidase IB (Genbank AN 6678787) and IA (Genbank AN 6754619) catalytic domains with a sub-library including sequences encoding Cop II vesicle, ER, and early Golgi localization peptides according to Table 6. The 25 combined DNA library was used to generate individual fusion constructs, which were then transformed into the K3 expressing host organism, resulting in a genetically mixed population wherein individual transformants each express K3 as well as a localization signal/mannosidase fusion gene from the library. Individual transformants were cultured and the production of K3 was induced by transfer to a 30 methanol containing medium. Under these conditions, after 24 hours of induction, over 90% of the protein in the medium was K3. The K3 reporter protein was purified from the supernatant to remove salts and low-molecular weight contaminants by Ni-affinity chromatography. Following affinity purification, the

protein was desalted by size exclusion chromatography on a Sephadex G10 resin (Sigma, St. Louis, MO) and either directly subjected to MALDI-TOF analysis described below or the *N*-glycans were removed by PNGase digestion as described below (Release of *N*-glycans) and subjected to MALDI-TOF analysis Miele *et al.*

5 (1997) *Biotechnol. Appl. Biochem.* 25:151-157.

[0341] Following this approach, a diverse set of transformants were obtained; some showed no modification of the *N*-glycans compared to the *ochI* knockout strain; and others showed a high degree of mannose trimming (Figures 5D and 5E). Desired transformants expressing appropriately targeted, active α -1,2-mannosidase produced K3 with *N*-glycans of the structure $\text{Man}_5\text{GlcNAc}_2$. This confers a reduced molecular mass to the glycoprotein compared to the K3 of the parent *ochI* deletion strain, a difference which was readily detected by MALDI-TOF mass spectrometry (Figure 5). Table 7 indicates the relative $\text{Man}_5\text{GlcNAc}_2$ production levels.

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Table 7. A representative combinatorial DNA library of localization sequences/catalytic domains exhibiting relative levels of $\text{Man}_5\text{GlcNAc}_2$ production.

		<i>Targeting peptide sequences</i>				
		<i>MNS1(s)</i>	<i>MNS1(m)</i>	<i>MNS1(l)</i>	<i>SEC12(s)</i>	<i>SEC12(m)</i>
Catalytic Domains	Mouse mannosidase 1A $\Delta 187$	FB4 ++	FB5 +	FB6 -	FB7 ++	FB8 ++++
	Mouse mannosidase 1B $\Delta 58$	GB4 ++	GB5 +	GB6 +	GB7 ++	GB8 +
	Mouse mannosidase 1B $\Delta 99$	GC4 -	GC5 +++	GC6 +	GC7 +	GC8 +
	Mouse mannosidase 1B $\Delta 170$	GD4 -	GD5 -	GD6 -	GD7 +	GD8 +

Table 8. Another combinatorial DNA library of localization sequences/catalytic domains exhibiting relative levels of $\text{Man}_5\text{GlcNAc}_2$ production.

		<i>Targeting peptide sequences</i>					
Catalytic Domains		<i>VANI(s)</i>	<i>VANI(m)</i>	<i>VANI(l)</i>	<i>MNN10(s)</i>	<i>MNN10(m)</i>	<i>MNN10(l)</i>
	<i>C. elegans</i> mannosidase 1B $\Delta 80$	BC18-5	BC19	BC20	BC27	BC28	BC29
		+++++	++++	+++	+++++	++++	+++
	<i>C. elegans</i> mannosidase 1B $\Delta 31$	BB18	BB19	BB20	BB18	BB19	BB20
		+++++	++++	+++	+++++	++++	++++

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[0342] Targeting peptides were selected from *MNS I* (SwissProt P32906) in *S. cerevisiae* (long, medium and short) (see supra Nucleic Acid Libraries; Combinatorial DNA Library of Fusion Constructs) and *SEC12* (SwissProt P11655) in *S. cerevisiae* (988-1140 nucleotides: short) and (988-1296: medium). Although majority of the targeting peptide sequences were *N*-terminal deletions, some targeting peptide sequences, such as *SEC12* were *C*-terminal deletions. Catalytic domains used in this experiment were selected from mouse mannosidase 1A with a 187 amino acid *N*-terminal deletion; and mouse mannosidase 1B with a 58, 99 and 170 amino acid deletion. The number of (+)s, as used herein, indicates the relative levels of $\text{Man}_5\text{GlcNAc}_2$ production. The notation (-) indicates no apparent production of $\text{Man}_5\text{GlcNAc}_2$. The notation (+) indicates less than 10% production of $\text{Man}_5\text{GlcNAc}_2$. The notation (++) indicates about 10-20% production of $\text{Man}_5\text{GlcNAc}_2$. The notation with (+++) indicates about 20-40% production of $\text{Man}_5\text{GlcNAc}_2$. The notation with (++++) indicates about 50% production of $\text{Man}_5\text{GlcNAc}_2$. The notation with (+++++) indicates greater than 50% production of $\text{Man}_5\text{GlcNAc}_2$.

[0343] Table 9 shows relative amount of $\text{Man}_5\text{GlcNAc}_2$ on secreted K3. Six hundred and eight (608) different strains of *P. pastoris*, $\Delta och1$ were generated by transforming them with a single construct of a combinatorial genetic library that

was generated by fusing nineteen (19) α -1,2 mannosidase catalytic domains to thirty-two (32) fungal ER, and cis-Golgi leaders.

Table 9

Amount of Man ₅ GlcNAc ₂ on secreted K3 (% of total glycans)	Number of constructs (%)
N.D.*	19 (3.1)
0-10%	341 (56.1)
10-20%	50 (8.2)
20-40&	75 (12.3)
40-60%	72 (11.8)
More than 60%	51 (8.4) [†]
Total	608 (100)

5

* Several fusion constructs were not tested because the corresponding plasmids could not be propagated in *E. coli* prior to transformation into *P. pastoris*.

10 [†] Clones with the highest degree of Man₅GlcNAc₂ trimming (30/51) were further analyzed for mannosidase activity in the supernatant of the medium. The majority (28/30) displayed detectable mannosidase activity in the supernatant (e.g. Figure 4B). Only two constructs displayed high Man₅GlcNAc₂ levels, while lacking mannosidase activity in the medium (e.g. Figure 4C).

[0344] Table 7 shows two constructs pFB8 and pGC5, among others, displaying Man₅GlcNAc₂. Table 8 shows a more preferred construct, pBC18-5, a *S. cerevisiae VAN1(s)* targeting peptide sequence (from SwissProt 23642) ligated in-frame to a *C. elegans* mannosidase IB (Genbank AN CAA98114) 80 amino acid N-terminal deletion (*Saccharomyces Van1(s)/ C.elegans mannosidase IB Δ80*). This fusion construct also produces a predominant Man₅GlcNAc₂ structure, as shown in Figure 5E. This construct was shown to produce greater than 50% Man₅GlcNAc₂ (+++++).

Generation of a combinatorial localization/mannosidase library:

[0345] Generating a combinatorial DNA library of α -1,2-mannosidase catalytic domains fused to targeting peptides required the amplification of mannosidase domains with varying lengths of N-terminal deletions from a number of organisms. To approach this goal, the full length open reading frames (ORFs) of α -1,2-mannosidases were PCR amplified from either cDNA or genomic DNA obtained from the following sources: *Homo sapiens*, *Mus musculus*, *Drosophila*

melanogaster, *Caenorhabditis elegans*, *Aspergillus nidulans* and *Penicillium citrinum*. In each case, DNA was incubated in the presence of oligonucleotide primers specific for the desired mannosidase sequence in addition to reagents required to perform the PCR reaction. For example, to amplify the ORF of the *M.*

5 *musculus* α -1,2-mannosidase IA, the 5'-primer

ATGCCCGTGGGGGGCCTGTTGCCGCTTCAGTAGC (SEQ ID NO:52) and the 3'-primer

TCATTTCTCTTGCCATCAATTCCCTTCTGTTCACGG (SEQ ID NO:53) were incubated in the presence of Pfu DNA polymerase (Stratagene, La Jolla, CA)

10 and amplified under the conditions recommended by Stratagene using the cycling parameters: 94°C for 1min (1 cycle); 94°C for 30 sec, 68°C for 30 sec, 72°C for 3min (30 cycles). Following amplification the DNA sequence encoding the ORF was incubated at 72 °C for 5 min with 1U Taq DNA polymerase (Promega, Madison, WI) prior to ligation into pCR2.1-TOPO (Invitrogen, Carlsbad, CA) and 15 transformed into TOP10 chemically competent *E. coli*, as recommended by Invitrogen. The cloned PCR product was confirmed by ABI sequencing using primers specific for the mannosidase ORF.

[0346] To generate the desired N-terminal truncations of each mannosidase, the complete ORF of each mannosidase was used as the template in a subsequent 20 round of PCR reactions wherein the annealing position of the 5'-primer was specific to the 5'-terminus of the desired truncation and the 3'-primer remained specific for the original 3'-terminus of the ORF. To facilitate subcloning of the truncated mannosidase fragment into the yeast expression vector, pJN347 (Figure 2C) *Ascl* and *PacI* restriction sites were engineered onto each truncation product, 25 at the 5'- and 3'-termini respectively. The number and position of the N-terminal truncations generated for each mannosidase ORF depended on the position of the transmembrane (TM) region in relation to the catalytic domain (CD). For instance, if the stem region located between the TM and CD was less than 150bp, then only one truncation for that protein was generated. If, however, the stem region was 30 longer than 150bp then either one or two more truncations were generated depending on the length of the stem region.

[0347] An example of how truncations for the *M. musculus* mannosidase IA (Genbank AN 6678787) were generated is described herein, with a similar approach being used for the other mannosidases. **Figure 3** illustrates the ORF of the *M. musculus* α -1,2-mannosidase IA with the predicted transmembrane and 5 catalytic domains being highlighted in bold. Based on this structure, three 5'- primers were designed (annealing positions underlined in **Figure 3**) to generate the Δ 65-, Δ 105- and Δ 187-N-terminal deletions. Using the Δ 65 N-terminal deletion as an example the 5'-primer used was 5'-
GGCGCGCCGACTCCTCCAAGCTGCTCAGCGGGTCCTGTTCCAC-3'

10 (SEQ ID NO:54) (with the *AscI* restriction site highlighted in bold) in conjunction with the 3'-primer 5'-
CCTTAATTAATCATTCTCTTGCCATCAATTCCCTTCTGTTCACGG-
3' (SEQ ID NO:55) (with the *PacI* restriction site highlighted in bold). Both of these primers were used to amplify a 1561 bp fragment under the conditions
15 outlined above for amplifying the full length *M. musculus* mannosidase 1A ORF. Furthermore, like the product obtained for the full length ORF, the truncated product was also incubated with Taq DNA polymerase, ligated into pCR2.1-TOPO (Invitrogen, Carlsbad, CA), transformed into TOP10 and ABI sequenced. After having amplified and confirmed the sequence of the truncated mannosidase
20 fragment, the resulting plasmid, pCR2.1- Δ 65mMannIA, was digested with *AscI* and *PacI* in New England Biolabs buffer #4 (Beverly, MA) for 16h at 37°C. In parallel, the pJN347 (**Figure 2C**) was digested with the same enzymes and incubated as described above. Post-digestion, both the pJN347 (**Figure 2C**) backbone and the truncated catalytic domain were gel extracted and ligated using the
25 Quick Ligation Kit (New England Biolabs, Beverly, MA), as recommended by the manufacturers, and transformed into chemically competent DH5 α cells (Invitrogen, Carlsbad, CA). Colony PCR was used to confirm the generation of the pJN347-mouse Mannosidase IA Δ 65 construct.

[0348] Having generated a library of truncated α -1,2-mannosidase catalytic 30 domains in the yeast expression vector pJN347 (**Figure 2C**) the remaining step in generating the targeting peptide/catalytic domain library was to clone in-frame the targeting peptide sequences (**Figure 2**). Both the pJN347-mannosidase constructs

(Figure 2D) and the pCR2.1TOPO-targeting peptide constructs (Figure 2B) such as were incubated overnight at 37°C in New England Biolabs buffer #4 in the presence of the restriction enzymes *NotI* and *Ascl*. Following digestion, both the pJN347-mannosidase back-bone and the targeting peptide regions were gel-extracted and ligated using the Quick Ligation Kit (New England Biolabs, Beverly, MA), as recommended by the manufacturers, and transformed into chemically competent DH5 α cells (Invitrogen, Carlsbad, CA). Subsequently, the pJN347-targeting peptide/mannosidase constructs were ABI sequenced to confirm that the generated fusions were in-frame. The estimated size of the final targeting peptide/alpha-1,2-mannosidase library contains over 1300 constructs generated by the approach described above. Figure 2 illustrates construction of the combinatorial DNA library.

15 Engineering a *P. pastoris* *OCH1* knock-out strain with multiple auxotrophic markers.

[0349] The first step in plasmid construction involved creating a set of universal plasmids containing DNA regions of the *KEX1* gene of *P. pastoris* (Boehm *et al.* Yeast 1999 May;15(7):563-72) as space holders for the 5' and 3' regions of the genes to be knocked out. The plasmids also contained the *S. cerevisiae* Ura-blaster (Alani *et al.* (1987) Genetics 116:541-545) as a space holder for the auxotrophic markers, and an expression cassette with a multiple cloning site for insertion of a foreign gene. A 0.9-kb fragment of the *P. pastoris* *KEX1*-5' region was amplified by PCR using primers

GGCGAGCTCGGCTACCCGGCCAAGGCTGAGATCATTGTCCAGCTTCA
25 GA (SEQ ID NO:56) and
GCCCACGTCGACGGATCCGGTTAACATCGATTGGAGAGGCTGACACC
GCTACTA (SEQ ID NO:57) and *P. pastoris* genomic DNA as a template and cloned into the *SacI*, *Sall* sites of pUC19 (New England Biolabs, Beverly, MA). The resulting plasmid was cut with *BamHI* and *Sall*, and a 0.8-kb fragment of the 30 *KEX1*-3' region that had been amplified using primers
CGGGATCCACTAGTATTAAATCATATGTGCGAGTGTACAACTCTTCCC
ACATGG (SEQ ID NO:58) and

GGACGCGTCGACGGCCTACCCGGCCGTACGAGGAATTCTCGG
ATGACTTTTC (SEQ ID NO:59) was cloned into the open sites creating
pJN262. This plasmid was cut with *BamHI* and the 3.8-kb *BamHI*, *BglII* fragment
of pNKY51 (Alani *et al.* (1987) *Genetics* 116:541-545) was inserted in both
5 possible orientations resulting in plasmids pJN263 (**Figure 4A**) and pJN284
(**Figure 4B**).

[0350] An expression cassette was created with *NotI* and *PacI* as cloning sites.
The GAPDH promoter of *P. pastoris* was amplified using primers
CGGGATCCCTCGAGAGATCTTTGTAGAAATGTCTGGTGCCT (SEQ
10 ID NO:60) and
GGACATGCATGCACTAGTGCGGCCACGTGATAGTTGTTCA
ATTGATTGAAATAGGGACAA (SEQ ID NO:61) and plasmid pGAPZ-A
(Invitrogen) as template and cloned into the *BamHI*, *SphI* sites of pUC19 (New
England Biolabs, Beverly, MA) (**Figure 4B**). The resulting plasmid was cut with
15 *SpeI* and *SphI* and the CYC1 transcriptional terminator region ("TT") that had been
amplified using primers
CCTGCTAGCTTAATTAACCGCGGCACGTCCGACGGCGGCCA
CGGGTCCCA (SEQ ID NO:62) and
GGACATGCATGCGGATCCCTTAAGAGCCGGCAGCTTGCAAATT
20 AAAGCCTTCGAGCGTCCC (SEQ ID NO:63) and plasmid pPICZ-A
(Invitrogen) as a template was cloned into the open sites creating pJN261 (**Figure**
4B).

[0351] A knockout plasmid for the *P. pastoris* *OCH1* gene was created by
digesting pJN263 with *SalI* and *SpeI* and a 2.9-kb DNA fragment of the *OCH1*-5'
25 region, which had been amplified using the primers
GAACCACGTCGACGGCATTGCGGCCAAAACCTTTTCCTATT
CAAACACAAGGCATTGC (SEQ ID NO:64) and
CTCCAATACTAGTCGAAGATTATCTTCTACGGTGCCTGGACTC (SEQ ID
NO:65) and *P. pastoris* genomic DNA as a template, was cloned into the open sites
30 (**Figure 4C**). The resulting plasmid was cut with *EcoRI* and *PmeI* and a 1.0-kb
DNA fragment of the *OCH1*-3' region that had been generated using the primers
TGGAAAGGTTAACAAAGCTAGAGTAAAATAGATATAGCGAG

ATTAGAGAATG (SEQ ID NO:66) and
AAGAATTCGGCTGGAAGGCCTTGTACCTTGATGTAGTTCCCGTT
TTCATC (SEQ ID NO:67) was inserted to generate pJN298 (**Figure 4C**). To
allow for the possibility to simultaneously use the plasmid to introduce a new gene,
5 the *BamHI* expression cassette of pJN261 (**Figure 4B**) was cloned into the unique
BamHI site of pJN298 (**Figure 4C**) to create pJN299 (**Figure 4E**).
[0352] The *P. pastoris* Ura3-blaster cassette was constructed using a similar
strategy as described in Lu *et al.* (1998) *Appl. Microbiol. Biotechnol.* 49:141-146.
A 2.0-kb *PstI*, *SpeI* fragment of *P. pastoris* *URA3* was inserted into the *PstI*, *XbaI*
10 sites of pUC19 (New England Biolabs, Beverly, MA) to create pJN306 (**Figure**
4D). Then a 0.7-kb *SacI*, *PvuII* DNA fragment of the *lacZ* open reading frame was
cloned into the *SacI*, *SmaI* sites to yield pJN308 (**Figure 4D**). Following digestion
of pJN308 (**Figure 4D**) with *PstI*, and treatment with T4 DNA polymerase, the
SacI - *PvuII* fragment from *lacZ* that had been blunt-ended with T4 DNA
15 polymerase was inserted generating pJN315 (**Figure 4D**). The *lacZ*/URA3
cassette was released by digestion with *SacI* and *SphI*, blunt ended with T4 DNA
polymerase and cloned into the backbone of pJN299 that had been digested with
PmeI and *AflII* and blunt ended with T4 DNA polymerase. The resulting plasmid
was named pJN329 (**Figure 4E**).
20 [0353] A *HIS4* marked expression plasmid was created by cutting pJN261
(**Figure 4F**) with EcoICRI (**Figure 4F**). A 2.7kb fragment of the *Pichia pastoris*
HIS4 gene that had been amplified using the primers
GCCCAAGCCGGCCTTAAGGGATCTCCTGATGACTGACTCACTGATAATA
AAAATACGG (SEQ ID NO:68) and
25 GGGCGCGTATTAAATACTAGTGGATCTATCGAATCTAAATGTAAGTTA
AAATCTCTAA (SEQ ID NO:69) cut with *NgoMIV* and *SwaI* and then blunt-
ended using T4 DNA polymerase, was then ligated into the open site. This
plasmid was named pJN337 (**Figure 4F**). To construct a plasmid with a multiple
cloning site suitable for fusion library construction, pJN337 was cut with *NotI* and
30 *PacI* and the two oligonucleotides
GGCCGCCTGCAGATTAAATGAATTCGCGCGCCTTAAT (SEQ ID
NO:70) and TAAGGCGCGCCGAATTCATTAAATCTGCAGGGC (SEQ ID

NO:71) that had been annealed *in vitro* were ligated into the open sites, creating pJN347 (**Figure 4F**).

[0354] To create an *och1* knockout strain containing multiple auxotrophic markers, 100 µg of pJN329 was digested with *SfiI* and used to transform *P. pastoris* strain JC308 (Cereghino *et al.* (2001) *Gene* 263:159-169) by electroporation. Following transformation, the URA dropout plates were incubated at room temperature for 10 days. One thousand (1000) colonies were picked and restreaked. All 1000 clones were then streaked onto 2 sets of URA dropout plates. One set was incubated at room temperature, whereas the second set was incubated at 37°C. The clones that were unable to grow at 37°C, but grew at room temperature, were subjected to colony PCR to test for the correct *OCH1* knockout. One clone that showed the expected PCR signal (about 4.5 kb) was designated YJN153.

EXAMPLE 5
Characterization of the Combinatorial DNA Library

[0355] Positive transformants screened by colony PCR confirming integration of the mannosidase construct into the *P. pastoris* genome were subsequently grown at room temperature in 50ml BMGY buffered methanol-complex medium consisting of 1% yeast extract, 2% peptone, 100 mM potassium phosphate buffer, pH 6.0, 1.34% yeast nitrogen base, 4 X 10⁻⁵% biotin, and 1% glycerol as a growth medium until OD_{600nm} 2-6 at which point they were washed with 10ml BMMY (buffered methanol-complex medium consisting of 1% yeast extract, 2% peptone, 100 mM potassium phosphate buffer, pH 6.0, 1.34% yeast nitrogen base, 4 X 10⁻⁵% biotin, and 1.5% methanol as a growth medium) media prior to induction of the reporter protein for 24 hours at room temperature in 5ml BMMY. Consequently, the reporter protein was isolated and analyzed as described in Example 3 to characterize its glycan structure. Using the targeting peptides in Table 6, mannosidase catalytic domains localized to either the ER or the Golgi showed significant level of trimming of a glycan predominantly containing Man₉GlcNAc₂ to a glycan predominantly containing Man₅GlcNAc₂. This is evident when the glycan structure of the reporter glycoprotein is compared between that of *P. pastoris och1* knock-out in **Figures 5C and 6C** and the same strain transformed

with *M. musculus* mannosidase constructs as shown in Figures 5D, 5E, 6D – 6F. Figures 5 and 6 show expression of constructs generated from the combinatorial DNA library which show significant mannosidase activity in *P. pastoris*.

Expression of pGC5 (*Saccharomyces MNS1(m)*/mouse mannosidase IB Δ99) 5 (Figures 5D and 6E) produced a protein which has approximately 30% of all glycans trimmed to Man₅GlcNAc₂, while expression of pFB8 (*Saccharomyces SEC12(m)*/mouse mannosidase IA Δ187) (Figure 6F) produced approximately 50% Man₅GlcNAc₂ and expression of pBC18-5 (*Saccharomyces VAN1(s)*/ *C. elegans* mannosidase IB Δ80) (Figure 5E) produced 70% Man₅GlcNAc₂.

10

EXAMPLE 6 Trimming *in vivo* by α-1,2-mannosidase

[0356] To ensure that the novel engineered strains of Example 4 in fact produced the desired Man₅GlcNAc₂ structure *in vivo*, cell supernatants were tested 15 for mannosidase activity (see Figures 7 – 9). For each construct/host strain described below, HPLC was performed at 30°C with a 4.0mm x 250 mm column of Altech (Avondale, PA, USA) Econosil-NH₂ resin (5μm) at a flow rate of 1.0 ml/min for 40 min. In Figures 7 and 8, degradation of the standard Man₉GlcNAc₂ [b] was shown to occur resulting in a peak which correlates to Man₈GlcNAc₂. In 20 Figure 7, the Man₉GlcNAc₂ [b] standard eluted at 24.61 min and Man₅GlcNAc₂ [a] eluted at 18.59 min. In Figure 8, Man₉GlcNAc₂ eluted at 21.37 min and Man₅GlcNAc₂ at 15.67 min. In Figure 9, the standard Man₈GlcNAc₂ [b] was shown to elute at 20.88 min.

[0357] *P. pastoris* cells comprising plasmid pFB8 (*Saccharomyces SEC12(m)*/mouse mannosidase IA Δ187) were grown at 30°C in BMGY to an OD600 of about 10. Cells were harvested by centrifugation and transferred to BMMY to induce the production of K3 (kringle 3 from human plasminogen) under control of an *AOX1* promoter. After 24 hours of induction, cells were removed by centrifugation to yield an essentially clear supernatant. An aliquot of the 25 supernatant was removed for mannosidase assays and the remainder was used for the recovery of secreted soluble K3. A single purification step using CM-sepharose chromatography and an elution gradient of 25mM NaAc, pH5.0 to 30

25mM NaAc, pH5.0, 1M NaCl, resulted in a 95% pure K3 eluting between 300-500mM NaCl. *N*-glycan analysis of the K3 derived glycans is shown in Figure 6F. The earlier removed aliquot of the supernatant was further tested for the presence of secreted mannosidase activity. A commercially available standard of 5 2-aminobenzamide-labeled N-linked-type oligomannose 9 (Man9-2-AB) (Glyko, Novato, CA) was added to: BMMY (Figure 7A), the supernatant from the above aliquot (Figure 7B), and BMMY containing 10ng of 75mU/mL of α -1,2-mannosidase from *Trichoderma reesei* (obtained from Contreras *et al.*, WO 02/00856 A2) (Figure 7C). After incubation for 24 hours at room temperature, 10 samples were analyzed by amino silica HPLC to determine the extent of mannosidase trimming.

[0358] *P. pastoris* cells comprising plasmid pGC5 (*Saccharomyces MNS1(m)/mouse mannosidase IB Δ99*) were similarly grown and assayed. Cells were grown at room temperature in BMGY to an OD600 of about 10. Cells were 15 harvested by centrifugation and transferred to BMMY to induce the production of K3 under control of an *AOX1* promoter. After 24 hours of induction, cells were removed by centrifugation to yield an essentially clear supernatant. An aliquot of the supernatant was removed for mannosidase assays and the remainder was used for the recovery of secreted soluble K3. A single purification step using CM-20 sepharose chromatography and an elution gradient of 25mM NaAc, pH5.0 to 25mM NaAc, pH5.0, 1M NaCl, resulted in a 95% pure K3 eluting between 300-500mM NaCl. *N*-glycan analysis of the K3 derived glycans is shown in Figure 5D. The earlier removed aliquot of the supernatant was further tested for the presence of secreted mannosidase activity as shown in Figure 8B. A 25 commercially available standard of Man9-2-AB (Glyko, Novato, CA) were added to: BMMY (Figure 8A), supernatant from the above aliquot (Figure 8B), and BMMY containing 10ng of 75mU/mL of α -1,2-mannosidase from *Trichoderma reesei* (obtained from Contreras *et al.*, WO 02/00856 A2) (Figure 8C). After 30 incubation for 24 hours at room temperature, samples were analyzed by amino silica HPLC to determine the extent of mannosidase trimming.

[0359] Man9-2-AB was used as a substrate and it is evident that after 24 hours of incubation, mannosidase activity was virtually absent in the supernatant of the

pFB8 (*Saccharomyces SEC12* (m)/mouse mannosidase IA Δ187) strain digest (Figure 7B) and pGC5 (*Saccharomyces MNS1*(m)/mouse mannosidase IB Δ99) strain digest (Figure 8B) whereas the positive control (purified α-1,2-mannosidase from *T. reesei* obtained from Contreras) leads to complete conversion of 5 Man₉GlcNAc₂ to Man₅GlcNAc₂ under the same conditions, as shown in Figures 7C and 8C. This is conclusive data showing *in vivo* mannosidase trimming in *P. pastoris* pGC5 strain; and pFB8 strain, which is distinctly different from what has been reported to date (Contreras *et al.*, WO 02/00856 A2).

[0360] [0360] Figure 9 further substantiates localization and activity of the mannosidase 10 enzyme. *P. pastoris* comprising pBC18-5 (*Saccharomyces VAN1*(s)/*C. elegans* mannosidase IB Δ80) was grown at room temperature in BMGY to an OD600 of about 10. Cells were harvested by centrifugation and transferred to BMMY to induce the production of K3 under control of an *AOX1* promoter. After 24 hours of induction, cells were removed by centrifugation to yield an essentially clear 15 supernatant. An aliquot of the supernatant was removed for mannosidase assays and the remainder was used for the recovery of secreted soluble K3. A single purification step using CM-sepharose chromatography and an elution gradient 25mM NaAc, pH5.0 to 25mM NaAc, pH5.0, 1M NaCl, resulted in a 95% pure K3 eluting between 300-500mM NaCl. *N*-glycan analysis of the K3 derived glycans is 20 shown in Figure 5E. The earlier removed aliquot of the supernatant was further tested for the presence of secreted mannosidase activity as shown in Figure 9B. A commercially available standard of Man8-2-AB (Glyko, Novato, CA) was added to: BMMY (Figure 9A), supernatant from the above aliquot pBC18-5 (*Saccharomyces VAN1*(s)/*C. elegans* mannosidase IB Δ80) (Figure 9B), and 25 BMMY containing media from a different fusion construct pDD28-3 (*Saccharomyces MNN10*(m) (from SwissProt 50108)/*H. sapiens* mannosidase IB Δ99) (Figure 9C). After incubation for 24 hours at room temperature, samples were analyzed by amino silica HPLC to determine the extent of mannosidase trimming. Figure 9B demonstrates intracellular mannosidase activity in 30 comparison to a fusion construct pDD28-3 (*Saccharomyces MNN10*(m) *H. sapiens* mannosidase IB Δ99) exhibiting a negative result (Figure 9C).

EXAMPLE 7
pH Optimum Assay of Engineered α -1,2-mannosidase

[0361] *P. pastoris* cells comprising plasmid pBB27-2 (*Saccharomyces MNN10* (s) (from SwissProt 50108)/*C. elegans* mannosidase IB Δ31) were grown at room temperature in BMGY to an OD600 of about 17. About 80 μ L of these cells were inoculated into 600 μ L BMGY and were grown overnight. Subsequently, cells were harvested by centrifugation and transferred to BMMY to induce the production of K3 (kringle 3 from human plasminogen) under control of an *AOX1* promoter. After 24 hours of induction, cells were removed by centrifugation to yield an essentially clear supernatant (pH 6.43). The supernatant was removed for mannosidase pH optimum assays. Fluorescence-labeled Man₈GlcNAc₂ (0.5 μ g) was added to 20 μ L of supernatant adjusted to various pH (Figure 11) and incubated for 8 hours at room temperature. Following incubation the sample was analyzed by HPLC using an Econosil NH2 4.6 X 250 mm, 5 micron bead, amino-bound silica column (Altech, Avondale, PA). The flow rate was 1.0 ml/min for 40 min and the column was maintained to 30°C. After eluting isocratically (68% A:32% B) for 3 min, a linear solvent gradient (68% A:32% B to 40% A:60% B) was employed over 27 min to elute the glycans (18). Solvent A (acetonitrile) and solvent B (ammonium formate, 50 mM, pH 4.5. The column was equilibrated with solvent (68% A:32% B) for 20 min between runs.

EXAMPLE 8
Engineering of *P. pastoris* to Produce N-glycans with the Structure GlcNAcMan₅GlcNAc₂

[0362] GlcNAc Transferase I activity is required for the maturation of complex and hybrid N-glycans (U.S. Pat. No. 5,834,251). Man₅GlcNAc₂ may only be trimmed by mannosidase II, a necessary step in the formation of human glycoforms, after the addition of N-acetylglucosamine to the terminal α-1,3 mannose residue of the trimannose stem by GlcNAc Transferase I (Schachter, 1991 Glycobiology 1(5):453-461). Accordingly, a combinatorial DNA library was prepared including DNA fragments encoding suitably targeted catalytic domains of GlcNAc Transferase I genes from *C. elegans* and *Homo sapiens*; and localization sequences from *GLS*, *MNS*, *SEC*, *MNN9*, *VAN1*, *ANP1*, *HOC1*, *MNN10*, *MNN11*.

MNT1, KTR1, KTR2, MNN2, MNN5, YURI, MNN1, and MNN6 from *S. cerevisiae* and *P. pastoris* putative α -1,2-mannosyltransferases based on the homology from *S. cerevisiae*: D2, D9 and J3, which are *KTR* homologs. Table 10 includes but does not limit targeting peptide sequences such as *SEC* and *OCHI*, from *P. pastoris* and *K. lactis* GnTI, (See Table 6 and Table 10)

Table 10. A representative combinatorial library of targeting peptide sequences/ catalytic domain for UDP-N-Acetylglucosaminyl Transferase I (GnTI)

Catalytic Domain		Targeting peptide				
		<i>OCHI</i> (s)	<i>OCHI</i> (m)	<i>OCHI</i> (l)	<i>MNN9</i> (s)	<i>MNN9</i> (m)
Human, GnTI, Δ38	PB105	PB106	PB107	PB104	N/A	
Human, GnTI, Δ86	NB12	NB13	NB14	NB15	NB	
<i>C. elegans</i> , GnTI, Δ88	OA12	OA13	OA14	OA15	OA16	
<i>C. elegans</i> , GnTI, Δ35	PA12	PA13	PA14	PA15	PA16	
<i>C. elegans</i> , GnTI, Δ63	PB12	PB13	PB14	PB15	PB16	
<i>X. laevis</i> , GnTI, Δ33	QA12	QA13	QA14	QA15	QA16	
<i>X. laevis</i> , GnTI, Δ103	QB12	QB13	QB14	QB15	QB 16	

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[0363] Targeting peptide sequences were selected from *OCHI* in *P. pastoris* (long, medium and short) (see Example 4) and *MNN9* (SwissProt P39107) in *S. cerevisiae* (short and medium). Catalytic domains were selected from human GnTI with a 38 and 86 amino acid *N*-terminal deletion, *C. elegans* (gly-12) GnTI with a 35 and 63 amino acid deletion as well as *C. elegans* (gly-14) GnTI with a 88 amino acid *N*-terminal deletion and *X. laevis* GnTI with a 33 and 103 amino acid *N*-terminal deletion, respectively.

[0364] A portion of the gene encoding human *N*-acetylglucosaminyl Transferase I (MGATI, Accession# NM002406), lacking the first 154 bp, was amplified by PCR using oligonucleotides 5'-TGGCAGGCAGCGCCTCAGTCAGCGCTCTCG-3' (SEQ ID NO:72) and 5'-AGGTAAATTA AGTGCTAATTCCAGCTAGG-3' (SEQ ID NO:73) and vector pHG4.5 (ATCC# 79003) as template. The resulting PCR product was cloned into pCR2.1-TOPO and the correct sequence was confirmed. Following digestion with *AscI* and *PacI* the truncated GnTI was inserted into plasmid pJN346 to create pNA. After digestion of pJN271 with *NotI*

and *Ascl*, the 120 bp insert was ligated into pNA to generate an in-frame fusion of the *MNN9* transmembrane domain with the GnTI, creating pNA15.

[0365] The host organism is a strain of *P. pastoris* that is deficient in hypermannosylation (e.g. an *och1* mutant), provides the substrate UDP-GlcNAc in 5 the Golgi and/or ER (i.e., contains a functional UDP-GlcNAc transporter), and provides *N*-glycans of the structure Man₅GlcNAc₂ in the Golgi and/or ER (e.g. *P. pastoris* pFB8 (*Saccharomyces SEC12* (m)/mouse mannosidase IA Δ187) from above). First, *P. pastoris* pFB8 was transformed with pPB103 containing the 10 *Kluyveromyces lactis MNN2-2* gene (Genbank AN AF106080) (encoding UDP-GlcNAc transporter) cloned into *BamHI* and *BglII* site of pBLADE-SX plasmid (Cereghino *et al.* (2001) *Gene* 263:159-169). Then the aforementioned combinatorial DNA library encoding a combination of exogenous or endogenous 15 GnTI/localization genes was transformed and colonies were selected and analyzed for the presence of the GnTI construct by colony PCR. Our transformation and integration efficiency was generally above 80% and PCR screening can be omitted once robust transformation parameters have been established.

Protein Purification

[0366] K3 was purified from the medium by Ni-affinity chromatography 20 utilizing a 96-well format on a Beckman BioMek 2000 laboratory robot. The robotic purification is an adaptation of the protocol provided by Novagen for their HisBind resin. Another screening method may be performed using a specific terminal GlcNAc binding antibody, or a lectin such as the GSII lectin from 25 *Griffonia simplifolia*, which binds terminal GlcNAc (EY Laboratories, San Mateo, CA). These screens can be automated by using lectins or antibodies that have been modified with fluorescent labels such as FITC or analyzed by MALDI-TOF.

[0367] Secreted K3 can be purified by Ni-affinity chromatography, quantified 30 and equal amounts of protein can be bound to a high protein binding 96-well plate. After blocking with BSA, plates can be probed with a GSII-FACS lectin and screened for maximum fluorescent response. A preferred method of detecting the above glycosylated proteins involves the screening by MALDI-TOF mass

spectrometry following the affinity purification of secreted K3 from the supernatant of 96-well cultured transformants. Transformed colonies were picked and grown to an OD₆₀₀ of 10 in a 2ml, 96-well plate in BMGY at 30°C. Cells were harvested by centrifugation, washed in BMMY and resuspended in 250ul of 5 BMMY. Following 24 hours of induction, cells were removed by centrifugation, the supernatant was recovered and K3 was purified from the supernatant by Ni affinity chromatography. The N-glycans were released and analyzed by MALDI-TOF delayed extraction mass spectrometry as described herein.

[0368] In summary, the methods of the invention yield strains of *P. pastoris* that 10 produce GlcNAcMan₅GlcNAc₂ in high yield, as shown in Figure 10B. At least 60% of the N-glycans are GlcNAcMan₅GlcNAc₂. To date, no report exists that describes the formation of GlcNAcMan₅GlcNAc₂ on secreted soluble 15 glycoproteins in any yeast. Results presented herein show that addition of the UDP-GlcNAc transporter along with GnTI activity produces a predominant GlcNAcMan₅GlcNAc₂ structure, which is confirmed by the peak at 1457 (m/z) (Figure 10B).

Construction of strain PBP-3:

[0369] The *P. pastoris* strain expressing K3, (*Δoch1*, arg-, ade-, his-) was 20 transformed successively with the following vectors. First, pFB8 (*Saccharomyces SEC12* (m)/mouse mannosidase IA Δ187) was transformed in the *P. pastoris* strain by electroporation. Second, pPB103 containing *Kluyveromyces lactis* MNN2-2 gene (Genbank AN AF106080) (encoding UDP-GlcNAc transporter) cloned into pBLADE-SX plasmid (Cereghino *et al.* (2001) *Gene* 263:159-169) digested with 25 *Bam*H I and *Bgl*II enzymes was transformed in the *P. pastoris* strain. Third, pPB104 containing *Saccharomyces* MNN9(s)/human GnTI Δ38 encoding gene cloned as *Not*I-*Pac*I fragment into pJN336 was transformed into the *P. pastoris* strain.

EXAMPLE 9**Engineering *K. lactis* Cells to Produce N-glycans with the Structure
 $\text{Man}_5\text{GlcNAc}_2$** Identification and Disruption of the *K. lactis* OCH1 gene

5 [0370] The *OCH1* gene of the budding yeast *S. cerevisiae* encodes a 1,6-mannosyltransferase that is responsible for the first Golgi localized mannose addition to the $\text{Man}_8\text{GlcNAc}_2$ *N*-glycan structure on secreted proteins (Nakanishi-Shindo *et al.* (1993) *J. Biol. Chem.*; 268(35):26338-45). This mannose transfer is generally recognized as the key initial step in the fungal specific

10 polymannosylation of *N*-glycan structures (Nakanishi-Shindo *et al.* (1993) *J. Biol. Chem.* 268(35):26338-26345; Nakayama *et al.* (1992) *EMBO J.* 11(7):2511-19; Morin-Ganet *et al* (2000) *Traffic* 1(1):56-68). Deletion of this gene in *S. cerevisiae* results in a significantly shorter *N*-glycan structure that does not include this typical polymannosylation or a growth defect at elevated temperatures (Nakayama *et al.* (1992) *EMBO J.* 11(7):2511-19).

15 [0371] The *Och1p* sequence from *S. cerevisiae* was aligned with known homologs from *Candida albicans* (Genbank accession # AAL49987), and *P. pastoris* along with the *Hoc1* proteins of *S. cerevisiae* (Neiman *et al* (1997) *Genetics* 145(3):637-45 and *K. lactis* (PENDANT EST database) which are related

20 but distinct mannosyltransferases. Regions of high homology that were in common among *Och1p* homologs but distinct from the *Hoc1p* homologs were used to design pairs of degenerate primers that were directed against genomic DNA from the *K. lactis* strain MG1/2 (Bianchi *et al* (1987) *Current Genetics* 12:185-192). PCR amplification with primers RCD33

25 (CCAGAAGAATTCAATTYTGYCARTGG) (SEQ ID NO:74) and RCD34 (CAGTGAAAATACCTGGNCCNGTCCA) (SEQ ID NO:75) resulted in a 302 bp product that was cloned and sequenced and the predicted translation was shown to have a high degree of homology to *Och1* proteins (>55% to *S. cerevisiae Och1p*). [0372] The 302 bp PCR product was used to probe a Southern blot of genomic

30 DNA from *K. lactis* strain (MG1/2) with high stringency (Sambrook *et al.*, *Molecular Cloning: A Laboratory Manual*, 2nd ed., Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, 1989). Hybridization was observed in a pattern consistent with a single gene indicating that this 302 bp segment

corresponds to a portion of the *K. lactis* genome and *K. lactis* (*KLOCH1*) contains a single copy of the gene. To clone the entire *KLOCH1* gene, the Southern blot was used to map the genomic locus. Accordingly, a 5.2 kb *Bam*H*I/Pst**I* fragment was cloned by digesting genomic DNA and ligating those fragments in the range of 5.2
5 kb into pUC19 (New England Biolabs, Beverly, MA) to create a *K. lactis* subgenomic library. This subgenomic library was transformed into *E. coli* and several hundred clones were tested by colony PCR using RCD 33/34. The 5.2 kb clone containing the predicted *KLOCH1* gene was sequenced and an open reading frame of 1362 bp encoding a predicted protein that is 46.5% identical to the *S.*
10 *cerevisiae* *OCH1* gene. The 5.2 kb sequence was used to make primers for construction of an *och1::KAN^R* deletion allele using a PCR overlap method (Davidson *et al.* (2002) *Microbiol.* 148(Pt 8):2607-15). This deletion allele was transformed into two *K. lactis* strains and G418 resistant colonies selected. These colonies were screened by both PCR and for temperature sensitivity to obtain a
15 strain deleted for the *OCH1* ORF. The results of the experiment show strains which reveal a mutant PCR pattern, which were characterized by analysis of growth at various temperatures and *N*-glycan carbohydrate analysis of secreted and cell wall proteins following PNGase digestion. The *och1* mutation conferred a temperature sensitivity which allowed strains to grow at 30°C but not at 35°C.
20 **Figure 12A** shows a MALDI-TOF analysis of a wild type *K. lactis* strain producing *N*-glycans of $\text{Man}_8\text{GlcNAc}_2$ [c] and higher.

Identification, Cloning, and Disruption of the *K. lactis* *MNN1* gene

[0373] *S. cerevisiae MNN1* is the structural gene for the Golgi α-1,3-mannosyltransferase. The product of *MNN1* is a 762-amino acid type II membrane protein (Yip *et al.* (1994) *Proc Natl Acad Sci U S A.* 91(7):2723-7). Both *N*-linked and *O*-linked oligosaccharides isolated from *mnn1* mutants lack α-1,3-mannose linkages (Raschke *et al.* (1973) *J Biol Chem.* 248(13):4660-66).
25 [0374] The *Mnn1p* sequence from *S. cerevisiae* was used to search the *K. lactis* translated genomic sequences (PEDANT). One 405 bp DNA sequence encoding a putative protein fragment of significant similarity to *Mnn1p* was identified. An internal segment of this sequence was subsequently PCR amplified with primers
30

KMN1 (TGCCATCTTTAGGTCCAGGCCGTTC) (SEQ ID NO:76) and KMN2 (GATCCCACGACGCATCGTATTCTTTC), (SEQ ID NO:77) and used to probe a Southern blot of genomic DNA from *K. lactis* strain (MG1/2). Based on the Southern hybridization data a 4.2 Kb *BamHI-PstI* fragment was cloned by 5 generating a size-selected library as described herein. A single clone containing the *K. lactis MNN1* gene was identified by whole colony PCR using primers KMN1 and KMN2 and sequenced. Within this clone a 2241 bp ORF was identified encoding a predicted protein that was 34% identical to the *S. cerevisiae MNN1* gene. Primers were designed for construction of a *mnn1::NAT^R* deletion 10 allele using the PCR overlap method (Davidson *et al.* (2002) *Microbiol.* 148(Pt 8):2607-15).

[0375] This disruption allele was transformed into a strain of *K. lactis* by electroporation and nourseothricin resistant transformants were selected and PCR amplified for homologous insertion of the disruption allele. Strains that reveal a 15 mutant PCR pattern may be subjected to *N*-glycan carbohydrate analysis of a known reporter gene.

[0376] Figure 12B depicts the *N*-glycans from the *K. lactis och1 mnn1* deletion strain observed following PNGase digestion the MALDI-TOF as described herein. The predominant peak at 1908 (m/z) indicated as [d] is consistent with the mass of 20 *Man₉GlcNAc₂*.

[0377] Additional methods and reagents which can be used in the methods for modifying the glycosylation are described in the literature, such as U.S. Patent No. 5,955,422, U.S. Patent No. 4,775,622, U.S. Patent No. 6,017,743, U.S. Patent No. 4,925,796, U.S. Patent No. 5,766,910, U.S. Patent No. 5,834,251, U.S. Patent No. 25 5,910,570, U.S. Patent No. 5,849,904, U.S. Patent No. 5,955,347, U.S. Patent No. 5,962,294, U.S. Patent No. 5,135,854, U.S. Patent No. 4,935,349, U.S. Patent No. 5,707,828, and U.S. Patent No. 5,047,335. Appropriate yeast expression systems can be obtained from sources such as the American Type Culture Collection, Rockville, MD. Vectors are commercially available from a variety of sources.

EXAMPLE 10**Identification, cloning and deletion of the *ALG3* gene in *P. pastoris* and *K. lactis*.**

[0378] Degenerate primers were generated based on an alignment of Alg3 protein sequences from *S. cerevisiae*, *H. sapiens*, and *D. melanogaster* and were used to amplify an 83 bp product from *P. pastoris* genomic DNA:

5'-GGTGTGTTGTTCTAGATCTTGCACTAYCARTT-3' (SEQ ID NO:78)
and
5'-AGAATTGGTGGGTAAAGAATTCCARCCACCACTCRTG-3' (SEQ ID NO:79). The resulting PCR product was cloned into the pCR2.1 vector (Invitrogen, Carlsbad, CA) and sequence analysis revealed homology to known *ALG3/RHK1/NOT56* homologs (Genbank NC_001134.2, AF309689, NC_003424.1). Subsequently, 1929 bp upstream and 2738 bp downstream of the initial PCR product were amplified from a *P. pastoris* genomic DNA library (Boehm (1999) *Yeast* 15(7):563-72) using the internal oligonucleotides 5'- CCTAAGCTGGTATCGCGTTCTCTTGCCATATC-3' (SEQ ID NO:80) and 5'-GCGGCATAAACATAATAGATGCTATAAAG-3' (SEQ ID NO:81) along with T3 (5'-AATTAACCCTCACTAAAGGG-3') (SEQ ID NO:49) and T7 (5'- GTAA TACGACTCACTATAGGGC-3') (SEQ ID NO:48) (Integrated DNA Technologies, Coralville, IA) in the backbone of the library bearing plasmid lambda ZAP II (Stratagene, La Jolla, CA). The resulting fragments were cloned into the pCR2.1-TOPO vector (Invitrogen) and sequenced. From this sequence, a 1395 bp ORF was identified that encodes a protein with 35% identity and 53% similarity to the *S. cerevisiae* *ALG3* gene (using BLAST programs). The gene was named *PpALG3*.

[0379] The sequence of *PpALG3* was used to create a set of primers to generate a deletion construct of the *PpALG3* gene by PCR overlap (Davidson *et al* (2002) *Microbiol.* 148(Pt 8):2607-15). Primers below were used to amplify 1 kb regions 5' and 3' of the *PpALG3* ORF and the KAN^R gene, respectively:

30 RCD142 (5'-CCACATCATCCGTGCTACATATAG-3') (SEQ ID NO:82),
RCD144 (5'-ACGAGGCAAGCTAACACAGATCTCGAAGTATCGAGGGTTATCCAG-3') (SEQ ID NO:83),
RCD145 (5'-CCATCCAGTGTGCAAAACGAGCCAATGGTCATGTCTATA

AATC-3') (SEQ ID NO:84),
RCD147 (5'-AGCCTCAGCGCCAACAAGCGATGG-3') (SEQ ID NO:85),
RCD143 (5'-CTGGATAACCCTCGATACTTCGAGATCTGTTAGCTTGCCTCGT-3') (SEQ ID NO:86), and
5 RCD146 (5'-GATTATAGACATGAACCATTGGCTCGTTTCGACACTGGATGG-3') (SEQ ID NO:87).

Subsequently, primers RCD142 and RCD147 were used to overlap the three resulting PCR products into a single 3.6 kb *alg3::KAN^R* deletion allele.

10 **Identification, cloning and deletion of the *ALG3* gene in *K. lactis*.**

[0380] The *ALG3p* sequences from *S. cerevisiae*, *Drosophila melanogaster*, *Homo sapiens* etc were aligned with *K. lactis* sequences (PENDANT EST database). Regions of high homology that were in common homologs but distinct in exact sequence from the homologs were used to create pairs of degenerate primers that were directed against genomic DNA from the *K. lactis* strain MG1/2 (Bianchi *et al*, 1987). In the case of *ALG3*, PCR amplification with primers KAL-1 (5'-ATCCTTACCGATGCTGTAT-3') (SEQ ID NO:88) and KAL-2 (5'-ATAACAGTATGTGTTACACCGCGTAG-3') (SEQ ID NO:89) resulted in a product that was cloned and sequenced and the predicted translation 15 was shown to have a high degree of homology to Alg3p proteins (>50% to *S. cerevisiae* Alg3p).

[0381] The PCR product was used to probe a Southern blot of genomic DNA from *K. lactis* strain (MG1/2) with high stringency (Sambrook *et al.*, *Molecular Cloning: A Laboratory Manual*, 2nd ed., Cold Spring Harbor Laboratory Press, 20 Cold Spring Harbor, NY, 1989). Hybridization was observed in a pattern 25 consistent with a single gene. This Southern blot was used to map the genomic loci. Genomic fragments were cloned by digesting genomic DNA and ligating those fragments in the appropriate size-range into pUC19 to create a *K. lactis* subgenomic library. This subgenomic library was transformed into *E. coli* and 30 several hundred clones were tested by colony PCR, using primers KAL-1 and KAL-2. The clones containing the predicted *KIALG3* and *KIALG61* genes were sequenced and open reading frames identified.

[0382] Primers for construction of an *alg3::NAT^R* deletion allele, using a PCR overlap method (Davidson *et al.* (2002) *Microbiol.* 148(Pt 8):2607-15), were designed and the resulting deletion allele was transformed into two *K. lactis* strains and NAT-resistant colonies selected. These colonies were screened by PCR and 5 transformants were obtained in which the *ALG3* ORF was replaced with the *och1::NAT^R* mutant allele.

EXAMPLE 11

10 Generation of an *alg3 och1* mutant strain expressing an α -1,2-mannosidase, GnT1 and GnTII for production of a human-like glycoprotein.

[0383] A *P. pastoris* *alg3::KAN^R* deletion construct was generated as described in Example 10. Approximately 5 μ g of the resulting PCR product was transformed into strain PBP-3 (see Example 3), and colonies were selected on YPD medium containing 200 μ g/ml G418. One strain out of 20 screened by PCR was confirmed 15 to contain the correct integration of the *alg3::KAN^R* mutant allele and lack the wild-type allele. This strain was named RDP27 (Figure 36).

[0384] A library of GnTII constructs was then generated, which was comprised of in-frame fusions of the leader library with the catalytic domains of GnTII genes from human and rat sources (WO 02/00879). This library was created in a *P. pastoris* integration vector containing the *NST^R* gene conferring resistance to the drug nourseothricin. The library plasmids were linearized with *Eco*RI, 20 transformed into strain RDP27 by electroporation, and the resulting strains were screened by analysis of the released glycans from purified K3. A *P. pastoris* strain expressing the rat GnTII fused in-frame to the *S. cerevisiae MNN9* (s) construct 25 was named PBP6-5 (Figure 36).

Generation of GnTII expression constructs

[0385] The construction of a GnTI expression vector (pNA15) containing a 30 human GnTI gene fused with the N-terminal part of *S. cerevisiae MNN9* gene is described in Choi *et al.* (2003) *Proc Natl Acad Sci U S A.* 100(9):5022-27. In a similar fashion, the rat GnTII gene was cloned. The rat GnTII gene (GenBank accession number U21662) was PCR amplified using Takara EX TaqTM

polymerase (*Panvera*) from rat liver cDNA library (Clontech) with
RAT1 (5'-TTCCTCACTGCAGTCTTCTATAACT-3') (SEQ ID NO:90) and
RAT2 (5'-TGGAGACCATGAGGTTCCGCATCTAC-3') (SEQ ID NO:91)
primers. The PCR product was then cloned into pCR2.1-TOPO vector (Invitrogen)
5 and sequenced. Using this vector as a template, the *Ascl-PacI* fragment of GnTII,
encoding amino-acids 88-443, was amplified with *Pfu Turbo* polymerase
(Stratagene) and primers,
RAT44 (5'-TTGGCGGCCCTCCCTAGTGTACCAGTTGAACCTTG-3') (SEQ
ID NO:92) and
10 RAT11 (5'-GATTAATTAACTCACTGCAGTCTTCTATAACT -3') (SEQ ID
NO:93) respectively, introduced *Ascl* and *PacI* restriction sites are underlined).
Following confirmation by sequencing, the catalytic domain of rat GnTII was then
cloned downstream of the *PMA1* promoter as a *Ascl-PacI* fragment in pBP124. In
the final step, the gene fragment encoding the *S. cerevisiae Mnn2* localization
15 signal was cloned from pJN281 as a *NotI-Ascl* fragment to generate an in-frame
fusion with the catalytic domain of GnTII, to generate plasmid pTC53.

EXAMPLE 12

Cloning and Expression Of GnTIII To Produce Bisecting GlcNAcs Which 20 Boost Antibody Functionality

[0386] The addition of an *N*-acetylglucosamine to the GlcNAc₂Man₃GlcNAc₂
structure by *N*-acetylglucosaminyltransferases III yields a so-called bisected *N*-
glycan (see Figure 15). This structure has been implicated in greater antibody-
dependent cellular cytotoxicity (ADCC) (Umana *et al.* (1999) *Nat. Biotechnol.*
25 17(2):176-80).

[0387] A host cell such as a yeast strain capable of producing glycoproteins with
bisected *N*-glycans is engineered according to the invention, by introducing into
the host cell a GnTIII activity. Preferably, the host cell is transformed with a
nucleic acid that encodes GnTIII (*e.g.*, a mammalian such as the murine GnTIII
30 shown in Figure 24) or a domain thereof having enzymatic activity, optionally
fused to a heterologous cell signal targeting peptide (*e.g.*, using the libraries and
associated methods of the invention.)

[0388] IgGs consist of two heavy-chains (V_H , C_{H1} , C_{H2} and C_{H3} in Figure 22), interconnected in the hinge region through three disulfide bridges, and two light chains (V_L , C_L in Figure 22). The light chains (domains V_L and C_L) are linked by another disulfide bridge to the C_{H1} portion of the heavy chain and together with the 5 C_{H1} and V_H fragment make up the so-called Fab region. Antigens bind to the terminal portion of the Fab region. The Fc region of IgGs consists of the C_{H3} , the C_{H2} and the hinge region and is responsible for the exertion of so-called effector functions (see below).

[0389] The primary function of antibodies is binding to an antigen. However, 10 unless binding to the antigen directly inactivates the antigen (such as in the case of bacterial toxins), mere binding is meaningless unless so-called effector-functions are triggered. Antibodies of the IgG subclass exert two major effector-functions: the activation of the complement system and induction of phagocytosis. The complement system consists of a complex group of serum proteins involved in 15 controlling inflammatory events, in the activation of phagocytes and in the lytical destruction of cell membranes. Complement activation starts with binding of the C1 complex to the Fc portion of two IgGs in close proximity. C1 consists of one molecule, C1q, and two molecules, C1r and C1s. Phagocytosis is initiated through an interaction between the IgG's Fc fragment and Fc-gamma-receptors (Fc γ RI, II 20 and III in Figure 22). Fc receptors are primarily expressed on the surface of effector cells of the immune system, in particular macrophages, monocytes, myeloid cells and dendritic cells.

[0390] The C_{H2} portion harbors a conserved N-glycosylation site at asparagine 297 (Asn297). The Asn297 N-glycans are highly heterogeneous and are known to 25 affect Fc receptor binding and complement activation. Only a minority (*i.e.*, about 15-20%) of IgGs bears a disialylated, and 3-10% have a monosialylated N-glycan (reviewed in Jefferis (2001) *Biopharm.* 14:19-26). Interestingly, the minimal N-glycan structure shown to be necessary for fully functional antibodies capable of complement activation and Fc receptor binding is a pentasacharide with terminal 30 N-acetylglucosamine residues ($GlcNAc_2Man_3$) (reviewed in Jefferis, R., Glycosylation of human IgG Antibodies. BioPharm, 2001). Antibodies with less than a $GlcNAc_2Man_3$ N-glycan or no N-glycosylation at Asn297 might still be able

to bind an antigen but most likely will not activate the crucial downstream events such as phagocytosis and complement activation. In addition, antibodies with fungal-type *N*-glycans attached to Asn297 will in all likelihood solicit an immune-response in a mammalian organism which will render that antibody useless as a
5 therapeutic glycoprotein.

Cloning And Expression Of GnTIII

[0391] The DNA fragment encoding part of the mouse GnTIII protein lacking the TM domain is PCR amplified from murine (or other mammalian) genomic
10 DNA using forward (5'-TCCTGGCGCGCTCCGAGAGAACTGGCCTCCCTC-3') (SEQ ID NO:94) and reversed (5'-AATTAATTAAACCCTAGCCCTCCGCTGTATCCAACTTG-3') (SEQ ID NO:95) primers. Those primers include *AscI* and *PacI* restriction sites
15 that may be used for cloning into the vector suitable for the fusion with leader library.
[0392] The nucleic acid (SEQ ID NO:45) and amino acid (SEQ ID NO:46) sequences of murine GnTIII are shown in Figure 24.

20 **Cloning of Immunoglobulin-Encoding Sequences**

[0393] Protocols for the cloning of the variable regions of antibodies, including primer sequences, have been published previously. Sources of antibodies and encoding genes can be, among others, *in vitro* immunized human B cells (*see, e.g.*, Borreback *et al.* (1988) *Proc. Natl. Acad. Sci. USA* 85:3995-3999), peripheral
25 blood lymphocytes or single human B cells (*see, e.g.*, Lagerkvist *et al.* (1995) *Biotechniques* 18:862-869; and Terness *et al.* (1997) *Hum. Immunol.* 56:17-27) and transgenic mice containing human immunoglobulin loci, allowing the creation of hybridoma cell-lines.

[0394] Using standard recombinant DNA techniques, antibody-encoding nucleic
30 acid sequences can be cloned. Sources for the genetic information encoding immunoglobulins of interest are typically total RNA preparations from cells of interest, such as blood lymphocytes or hybridoma cell lines. For example, by

employing a PCR based protocol with specific primers, variable regions can be cloned via reverse transcription initiated from a sequence-specific primer hybridizing to the IgG C_H1 domain site and a second primer encoding amino acids 111-118 of the murine kappa constant region. The V_H and V_K encoding cDNAs 5 can then be amplified as previously published (see, e.g., Graziano *et al.* (1995) *J Immunol.* 155(10):4996-5002; Welschof *et al.* (1995) *J. Immunol. Methods* 179:203-214; and Orlandi *et al.* (1988) *Proc. Natl. Acad. Sci. USA* 86:3833). Cloning procedures for whole immunoglobulins (heavy and light chains) have also 10 been published (see, e.g., Buckel *et al.* (1987) *Gene* 51:13-19; Recinos *et al.* (1994) *Gene* 149: 385-386; Recinos *et al.* (1995) *Gene* 158:311-12). Additional protocols for the cloning and generation of antibody fragment and antibody expression constructs have been described in *Antibody Engineering*, Kontermann and Dübel (2001), Eds., Springer Verlag: Berlin Heidelberg New York.

[0395] Fungal expression plasmids encoding heavy and light chain of 15 immunoglobulins have been described (see, e.g., Abdel-Salam *et al.* (2001) *Appl. Microbiol. Biotechnol.* 56:157-164; and Ogunjimi *et al.* (1999) *Biotechnology Letters* 21:561-567). One can thus generate expression plasmids harboring the constant regions of immunoglobulins. To facilitate the cloning of variable regions into these expression vectors, suitable restriction sites can be placed in close 20 proximity to the termini of the variable regions. The constant regions can be constructed in such a way that the variable regions can be easily in-frame fused to them by a simple restriction-digest / ligation experiment. Figure 23 shows a schematic overview of such an expression construct, designed in a very modular way, allowing easy exchange of promoters, transcriptional terminators, integration 25 targeting domains and even selection markers.

[0396] As shown in Figure 23, V_L as well as V_H domains of choice can be easily cloned in-frame with C_L and the C_H regions, respectively. Initial integration is targeted to the *P. pastoris* AOX locus (or homologous locus in another fungal cell) and the methanol-inducible AOX promoter will drive expression. Alternatively, 30 any other desired constitutive or inducible promoter cassette may be used. Thus, if desired, the 5'AOX and 3'AOX regions as well as transcriptional terminator (TT) fragments can be easily replaced with different TT, promoter and integration

targeting domains to optimize expression. Initially the alpha-factor secretion signal with the standard KEX protease site is employed to facilitate secretion of heavy and light chains. The properties of the expression vector may be further refined using standard techniques.

5 [0397] An Ig expression vector such as the one described above is introduced into a host cell of the invention that expresses GnTIII, preferably in the Golgi apparatus of the host cell. The Ig molecules expressed in such a host cell comprise N-glycans having bisecting GlcNAcs.

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EXAMPLE 13 Generation of Yeast Strain YSH-1 (*Och1*, α 1,2-mannosidase, GnTI)

[0398] The previously reported *P. pastoris* strain BK64 (Choi *et al.* (2003) *Proc Natl Acad Sci U S A.* 100(9):5022-7), a triple auxotroph (*ADE*, *ARG*, *HIS*) possessing the *OCH1* knock-out and expressing the kringle 3 domain (K3) of 15 human plasminogen, was used as the host strain. BK64 was transformed with the plasmid pPB103 linearized with the restriction enzyme *EcoNI* to introduce the *K. lactis* UDP-*N*-acetylglucosamine transporter into the host cell, thus creating the strain PBP-1. The mouse MnsI was introduced into this strain by transformation with the plasmid pFB8 linearized with the restriction enzyme *EcoNI*, generating 20 strain PBP-2. K3 glycan analysis from proteins isolated from strain PBP-2 demonstrated that the primary glycoform present was Man₅GlcNAc₂.

[0399] PBP-2 was subsequently transformed with the human GnTI plasmid pNA15 linearized with the restriction enzyme *AatII*, generating the strain PBP-3. Analysis of the K3 glycoforms produced in strain PBP-3 demonstrated that the 25 hybrid glycan GlcNAcMan₅GlcNAc₂ was the predominant structure. To recover the *URA3* marker from PBP-3, this strain was grown in YPD prior to selection on minimal media containing 5-Fluoroorotic (5-FOA, BioVectra) and uracil (Boeke *et al.* (1984) *Mol. Gen. Genet.* 197:345-346). The recovered Ura-minus strain producing GlcNAcMan₅GlcNAc₂ glycoforms was designated YSH-1 (Figure 36). 30 The *N*-glycan profile from strain YSH-1 is shown in Figure 25 (top) and displays a predominant peak at 1465 m/z corresponding to the mass of GlcNAcMan₅GlcNAc₂ [d].

EXAMPLE 14**Generation of Yeast Strain YSH-37 (*P. pastoris* expressing mannosidase II)**

[0400] YSH-1 (Example 13) was transformed with the *D. melanogaster* mannosidase II Δ 74/*S. cerevisiae* *MNN2*(s) plasmid (pKD53) linearized with the restriction enzyme *Apal*, generating strain YSH-37 (Figure 36). Analysis of the K3 glycan structures produced in strain YSH-37 (Figure 25 (bottom)) demonstrated that the predominant glycoform at 1140 m/z corresponds to the mass of GlcNAcMan₃GlcNAc₂ [b] and other glycoforms GlcNAcMan₄GlcNAc₂ [c] at 1303 m/z and GlcNAcMan₅GlcNAc₂ [d] at 1465 m/z.

EXAMPLE 15**Generation of Yeast Strain YSH-44**

[0401] Strain YSH-37 (Example 14) was transformed with a plasmid encoding a rat GnTII/*MNN2* (s) leader, pTC53, linearized with the restriction enzyme *EcoRI*. The resulting strain, YSH-44 (Figure 36), produced a K3 N-glycan having a single glycoform at 1356 m/z, corresponding to the mass of GlcNAc₂Man₃GlcNAc₂ [x], by positive mode MALDI-TOF mass spectrometry (Figure 29).

20 β -N-acetylhexosaminidase Digestion

[0402] The glycans from YSH-44 were released and separated from the glycoproteins by a modification of a previously reported method (Papac, et al. A. J. S. (1998) *Glycobiology* 8, 445-454). After the proteins were reduced and carboxymethylated and the membranes blocked, the wells were washed three time with water. The protein was deglycosylated by the addition of 30 μ l of 10 mM NH₄HCO₃ pH 8.3 containing one milliunit of N-glycanase (Glyko, Novato, CA). After a 16 hr digestion at 37°C, the solution containing the glycans was removed by centrifugation and evaporated to dryness. The glycans were then dried in aSC210A speed vac (Thermo Savant, Halbrook, NY). The dried glycans were put in 50 mM NH₄Ac pH 5.0 at 37°C overnight and 1mU of hexos (Glyko, Novato, CA) was added.

EXAMPLE 16
Construction of Plasmid pJN 348

[0403] The plasmid pBLURA-SX (from Jim Cregg) was digested with *BamHI* and *BglII* to release the *AOX* expression cassette. The *BamHI* fragment containing
5 the GAPDH/CYC1 expression cassette from pJN261 (Figure 4B) (Example 4)
was then ligated into the pBLURA-SX backbone to create pJN338. The plasmid
pJN338 was cut with *NotI* and *PacI* and the two oligonucleotides
5'-GGCCGCCTGCAGATTAAATGAATTCTGGCGCGCTTAAT-3' (SEQ ID
NO:96) and
10 5'-TAAGGCGCGCC GAATTCAATTAAATCTGCAGGGC-3' (SEQ ID NO:97)
that had been annealed *in vitro*, were ligated into the open sites, to create pJN348.

EXAMPLE 17
Construction of an Integration Plasmid pRCD259

15 [0404] The *PpURA3* containing GAPDH expression vector pJN348 was linearized with *XhoI* and blunted with T4 DNA polymerase and calf intestinal phosphatase (CIP) treated. The *HYG* resistance marker was digested from pAG32 with *BglII* and *SacI* and blunted, then ligated into pJN348 to create pRCD259 which can be used as a *HYG* expression vector that integrates at the *PpURA3* locus.
20

EXAMPLE 18
Generation of GnTIII Fusion Constructs

[0405] Fusion constructs between mammalian GnTIII and yeast targeting sequences were made using mouse *Mgat3* gene (GenBank accession number
25 L39373, Bhaumik *et al.*, 1995). Three DNA fragments corresponding to N-terminal deletions Δ32, Δ86, and Δ212 of the mouse GnTIII gene were PCR amplified using *Pfu Turbo* polymerase (Stratagene) with forward MG3-B (5'-TCCTGGCGCGCTTCCGAGAGAACTGGCCTCCCTC-3') (SEQ ID NO:98),
MG3-C (5'-CCGAGGCGCGCCACAGAGGAACTGCACC GG GTG-3') (SEQ ID NO:99),
MG3-D (5'-ACCGAGGCGCGCCATCAACGCCATCAACATCAACCAC-3') (SEQ ID NO:100),

and reverse

MG3-A (5'-AATTAATTAACCCTAGCCCTCGCTGTATCCAACTTG-3') (SEQ ID NO:101) primers. The PCR products were then cloned into pJN 348 vector as *AscI-PacI* fragments and sequenced. The resulting vectors pVA (GnTIII Δ32), pVB (GnTIII Δ86), and pVC (GnTIII Δ212) were digested with *NotI-AscI* enzymes and used for the ligation with yeast leader library (leaders 20-67). These targeting peptides are fused to the catalytic domains selected from the mouse GnTIII with 32, 86, 212 amino acid *N*-terminal deletions. For example, the *MNN2* targeting peptide from *S. cerevisiae* (long, medium and short) and *GNT1* from *K. lactis* (short, and medium) (see Example 11) are shown in Table 11.

Table 11. A representative combinatorial library of targeting peptide sequences/ catalytic domains exhibiting UDP-*N*-Acetylglucosaminyltransferase III (GnTIII) activity in *P. pastoris* YSH-1

15

Catalytic Domain		Targeting peptide			
		<i>S. cerevisiae</i> <i>MNN2(s)</i>	<i>S. cerevisiae</i> <i>MNN2(m)</i>	<i>S. cerevisiae</i> <i>MNN2(l)</i>	<i>K. lactis</i> <i>GNT1(m)</i>
Mouse GnTIII Δ32	50% (pVA53)	30-40% (pVA54)	20-30% (pVA55)	0% (pVA51)	
Mouse GnTIII Δ86	20-30% (pVB53)	30-40% (pVB54)	20-30% (pVB55)	0% (pVB51)	
Mouse GnTIII Δ212	0% (pVC53)	0% (pVC54)	0% (pVC55)	0% (pVC51)	

EXAMPLE 19

Engineering of *P. pastoris* to produce bisected GlcNAc₂Man₅GlcNAc₂

[0406] The *P. pastoris* strain producing GlcNAcMan₅GlcNAc₂ (PBP-3) (see Example 8) was counterselected on 5-FOA, thereby selecting for loss of the URA3+ marker and a ura3- phenotype. This strain, designated YSH-1 (Figure 36), was transformed with the library of *N*-acetylglucosaminyltransferase III (GnTIII) catalytic domains (vectors pVA, pVB, and pVC) and leaders. Transformants were grown at 30°C in BMGY to an OD600 of about 10, harvested by centrifugation and transferred to BMMY to induce the production of K3

(kringle 3 from human plasminogen) under control of an *AOX1* promoter. K3 was purified from the medium by Ni-affinity chromatography utilizing a 96-well format on a Beckman BioMek 2000 laboratory robot. The robotic purification is an adaptation of the protocol provided by Novagen for their HisBind resin

5 (Example 3). The *N*-glycans were released by PNGase digestion (Example 3). The *N*-glycans were analyzed with a MALDI-TOF MS (Example 3). The GnTIII activities are shown in Table 11. The number of (+)s, as used herein, indicates the relative levels of bisected *N*-glycan production of % neutral glycans. Targeting peptide sequences were selected from selected from the group consisting of:

10 10 *Saccharomyces* GLS1, *Saccharomyces* MNS1, *Saccharomyces* SEC12, *Pichia* SEC, *Pichia* OCH1, *Saccharomyces* MNN9, *Saccharomyces* VAN1, *Saccharomyces* ANP1, *Saccharomyces* HOC1, *Saccharomyces* MNN10, *Saccharomyces* MNN11, *Saccharomyces* MNT1, *Pichia* D2, *Pichia* D9, *Pichia* J3, *Saccharomyces* KTR1, *Saccharomyces* KTR2, *Kluyveromyces* GnTI,

15 15 *Saccharomyces* MNN2, *Saccharomyces* MNN5, *Saccharomyces* YUR1, *Saccharomyces* MNN1, and *Saccharomyces* MNN6. The pVA53 transformants exhibiting the bisecting GlcNAc (e.g. GlcNAc₂Man₅GlcNAc₂) were designated PBP26 (Figure 36).

20

EXAMPLE 20

Engineering of *P. pastoris* YSH-44 to produce bisected GlcNAc₃Man₃GlcNAc₂

[0407] For the expression of GnTIII in the strain YSH-44 (Figure 36), GnTIII constructs from vectors pVA53, pVB53, pVA54, and pVB54 were transferred as *NotI-PacI* fragments into pRCD259 to generate vectors pPB135, pPB137, pPB136, and pPB138. The vectors contain HYG resistance marker and *P. pastoris* *URA3* gene as targeting sequence for genomic integration. Plasmids are linearized with *Sall*, transformed into strain YSH-44 by electroporation, selected on medium containing hygromycin and the resulting strains are screened by analysis of the released glycans from purified K3. Transformants were grown at 24°C in BMGY to an OD600 of about 10, harvested by centrifugation and transferred to BMMY to induce the production of K3 (kringle 3 from human plasminogen) under control of an *AOX1* promoter. K3 was purified from the medium by Ni-affinity

chromatography utilizing a 96-well format on a Beckman BioMek 2000 laboratory robot (**Example 3**). The robotic purification is an adaptation of the protocol provided by Novagen for their HisBind resin (**Example 3**). The *N*-glycans were released by PNGase digestion. The *N*-glycans were analyzed with a MALDI-TOF MS (**Example 3**). The pPB135 transformants exhibiting the bisecting GlcNAc (*e.g.* GlcNAc₂Man₅GlcNAc₂) were designated **YSH-57** (**Figure 36**). **Table 11** depicts the activity of the mouse GnTIII.

EXAMPLE 21

10 **Engineering of *P. pastoris* PBP6-5 to produce bisected GlcNAc₃Man₃GlcNAc₂**
[0408] The *P. pastoris* **PBP6-5** (**Example 11**) was transformed with the plasmid pPB135 (**Table 11**) encoding a mouse GnTIII catalytic domain ($\Delta 32$) ligated in frame to a targeting peptide derived from *S. cerevisiae* *MNN2*. Transformants were grown at 30°C in BMGY to an OD600 of about 10, harvested by
15 centrifugation and transferred to BMMY to induce the production of K3 (kringle 3 from human plasminogen) under control of an *AOX1* promoter. K3 was purified from the medium by Ni-affinity chromatography utilizing a 96-well format on a Beckman BioMek 2000 laboratory robot. The robotic purification is an adaptation of the protocol provided by Novagen for their HisBind resin (**Example 3**). The *N*-
20 glycans were released by PNGase digestion (**Example 3**). The *N*-glycans were analyzed with a MALDI-TOF MS (**Example 3**). Transformants exhibiting the bisecting GlcNAc (*e.g.* GlcNAc₂Man₃GlcNAc₂) were designated **PBP-38** (**Figure 36**). **Table 11** depicts the activity of the mouse GnTIII.

EXAMPLE 22

In vitro GnTIII activity assay using substrate GlcNAcMan₅GlcNAc₂ in engineered *P. pastoris* strain YSH-57

25 [0409] To test any potential *ex vivo* GnTIII activity in the *P. pastoris* strain, YSH-57 cell culture supernatants were tested for GnTIII activity. *P. pastoris* YSH-
30 57 cells were grown at 24°C in BMGY to an OD600 of about 10. Cells were harvested by centrifugation and transferred to BMMY to induce the production of K3 (kringle 3 from human plasminogen) under control of an *AOX1* promoter. After 24 hours of induction, cells were removed by centrifugation to yield an

essentially clear supernatant. An aliquot of the supernatant was removed for GnTIII assays and the remainder was used for the recovery of secreted soluble K3. K3 was purified from the medium by Ni-affinity chromatography utilizing a 96-well format on a Beckman BioMek 2000 laboratory robot. The robotic purification 5 is an adaptation of the protocol provided by Novagen for their HisBind resin (**Example 3**). The N-glycans were released by PNGase digestion (**Example 3**). The earlier removed aliquot of the supernatant was further tested for the presence of secreted GnTIII activity. GlcNAcMan₅GlcNAc₂ purified from K3 expressed in PBP-3 strain was added to: BMMY (A) 1 mM UDP-GlcNAc (Sigma Chemical 10 Co., St. Louis, MO)) in BMMY (B); the supernatant of YSH-44 transformed with pVA53 [YSH-57] (C); the supernatant of YSH-57 + 1 mM UDP-GlcNAc (D). After incubation for 8 hours at room temperature, samples were analyzed by amino silica HPLC to determine the extent of GnTIII activity.

15

EXAMPLE 23

In vitro GnTIII activity assay using substrate GlcNAc₂Man₃GlcNAc₂ in engineered *P. pastoris* strain YSH-57

[0410] To test any potential *ex vivo* GnTIII activity in the *P. pastoris* strain YSH-57 cell culture supernatants were tested for GnTIII activity. *P. pastoris* YSH-20 57 cells were grown at 24°C in BMGY to an OD600 of about 10. Cells were harvested by centrifugation and transferred to BMMY to induce the production of K3 (kringle 3 from human plasminogen) under control of an *AOX1* promoter. After 24 hours of induction, cells were removed by centrifugation to yield an essentially clear supernatant. An aliquot of the supernatant was removed for GnTIII assays 25 and the remainder was used for the recovery of secreted soluble K3. K3 was purified from the medium by Ni-affinity chromatography utilizing a 96-well format on a Beckman BioMek 2000 laboratory robot. The robotic purification is an adaptation of the protocol provided by Novagen for their HisBind resin (**Example 3**). The N-glycans were released by PNGase digestion (**Example 3**). The earlier removed aliquot of the supernatant was further tested for the presence 30 of secreted GnTIII activity. GlcNAc₂Man₃GlcNAc₂ purified from K3 expressed in YSH-44 strain was added to: BMMY (A) 1 mM UDP-GlcNAc (Sigma Chemical Co., St. Louis, MO)) in BMMY (B); the supernatant of YSH-44 transformed with

pVA53 [YSH-57] (C). After incubation for 8 hours at room temperature, samples were analyzed by amino silica HPLC to determine the extent of GnTIII activity.

EXAMPLE 24

5 Cloning and Expression of GnTIV in *P.pastoris*

[0411] The DNA fragment encoding part of the human GnTIV protein isoenzyme A (MGAT4A) lacking the TM domain was PCR amplified from human cDNA using forward HGIV-2 (5'-CTGATTGCTTATCAACGAGAATTCCCTG-3') and 10 reversed HGIV-3 (5'-TGTGGTTCTCAGATGATCAGTTGGT-3') primers, cloned into the pCR2.1-TOPO vector (Invitrogen), and sequenced. For cloning into a vector suitable for the fusion with a leader library, DNA fragments were PCR amplified with primers that include *AscI* and *PacI* restriction sites. *AscI* forward primers: HGIV-ASC1 (5'-
15 TGGCGCGCCAAACTGATTGCTTATCAACGAGAA-3'), HGIV-ASC2 (5'-AGTGGCGCGCCTGAATAAGTTTCAGATAATACC-3'), HGIV-ASC3 (5'-AAGGGCGCGCCAAGTGCCAAGTATTATTATC-3'). *PacI* reverse primer HGIV-PAC (5'-
20 GTTTAATTAAGATCAGTTGGCTTTTAATATG-3'). The nucleic acid and amino acid sequences of the human GnTIV A are shown in Figure 41.
[0412] Similarly, the DNA fragment encoding part of the human GnTIV protein isoenzyme B (MGAT4B) lacking the TM domain was PCR amplified from human cDNA using forward HGIVB-2 (5'-
25 AGCGGCCAGAAAGGCGACGTTGTGGAC-3') and reverse HGIVB-3 (5'-TACCCTCAGAACGCCGCAGCTTAGTC-3') primers, cloned into the pCR2.1-TOPO vector (Invitrogen), and sequenced. For cloning into a vector suitable for the fusion with a leader library, DNA fragments were PCR amplified with primers that include *AscI* and *PacI* restriction sites. *AscI* forward primers: HGIVB-A1 (5'-
30 AGCGGGCGCGCCGGCGACGTTGTGGACGTTAC-3'), HGIVB-A2 (5'-CCGTGGCGCGCCTCACACCGGCACGTGCTGCAC-3'). *PacI* reverse primer HGIVB-P (5'-TGTAAATTAAGCTTAGTCGGCCTTTCAGGAAG-3'). The

nucleic acid and amino acid sequences of human GnTIV B are shown in Figure 42.

5 **Table 12: Plasmids containing GnTV or GnTIV Fusion Constructs For Expression of Multiantennary Structures**

Catalytic domain/leader	<i>HYG</i> vector	<i>KAN</i> vector
Mouse GnT V		
Δ95-53		pPB146
Δ145-53	pPB125	pPB140
Δ145-54	pPB130	
Δ209-53	pPB126	
Δ209-54	pPB131	
Human GnT IVA		
Δ32-53	pPB127	
Δ82-53	pPB128	pPB141
Δ103-53	pPB129	
Δ32-54	pPB132	
Δ82-54	pPB133	
Δ103-54	pPB134	
Human GnT IVB		
Δ32-53	pPB143	
Δ104-53	pPB144	
Human GnT IX		
Δ43-53		pPB176

EXAMPLE 25

10 ***P. pastoris* Strain Producing Triantennary Glycan Structures**

[0413] *P. pastoris* YSH-44 strain (Example 15) producing complex glycan structures was transformed with the plamid pPB144 (Table 12) containing a gene fragment encoding the human GnTIVB catalytic domain (Δ104) ligated in frame to a *S. cerevisiae* *MNN2(s)* [nucleotides 1-108] targeting peptide. The plasmid pPB144 also contains a *HYG* resistance marker and the *P. pastoris* *URA3* gene as targeting sequence for genomic integration. 1 μg plasmids were linearized with *SalI*, transformed into strain YSH-44 by electroporation, selected on medium containing hygromycin and the resulting strains were screened by analysis of the released glycans from purified K3. Transformants were grown at 30°C in BMGY to an OD600 of about 100, harvested by centrifugation and transferred to BMMY to induce the production of K3 (kringle 3 from human plasminogen) under control

of an *AOX1* promoter. K3 was purified from the medium by Ni-affinity chromatography utilizing a 96-well format on a Beckman BioMek 2000 laboratory robot. The robotic purification is an adaptation of the protocol provided by Novagen for their HisBind resin (**Example 3**). The N-glycans were released by 5 PNGase digestion (**Example 3**). The N-glycans were analyzed with a MALDI-TOF MS (**Example 3**). Transformants exhibiting the transfer of GlcNAc residues onto the Man α 1,3 arm of the oligosaccharide structure (e.g. GlcNAc₂Man₃GlcNAc₂) were designated PBP43 (**Figure 47**). Analysis of N-glycans provides a predominant peak at 1543 m/z [y] is consistent with the mass of 10 the glycan GlcNAc₃Man₃GlcNAc₂.

EXAMPLE 26
Cloning and Expression of GnTV in *P.pastoris*

15 [0414] The DNA fragment encoding part of the mouse GnTV protein (MGAT45) lacking the TM domain was PCR amplified from murine cDNA using forward MGV-2 (5'-AAATCAAGTGGATGAAGGACATGTGGC3') and reverse MGV-3 (5'-AGCGATGCTATAAGGCAGTCTTGCAGAG-3') primers, cloned into the pCR2.1-TOPO vector (Invitrogen), and sequenced. For cloning into 20 the vector suitable for the fusion with leader library, DNA fragments were PCR amplified with primers that include *AscI* and *PacI* restriction sites. *AscI* forward primers: MGV-ASC1 (5'- TATGGCGCGCCGATCATAACTCATTGGCGGAAATC-3'), MGV-ASC2 (5'-GAAGGGCGCGCCTGCCTCATGGATGGCTACCCCCAC-3'), MGV- 25 ASC3 (5'-TGGGGCGCGCCGGCAAGCTCGAGTCAAAGGTGGACAAT-3'). *PacI* reverse primer MGV-PAC (5'- AGTTAATTAATGCTATAAGGCAGTCTTGCAGAG-3'). The nucleic acid and amino acid sequences of mouse GnTV are shown in **Figure 43**.

30
EXAMPLE 27
***P. pastoris* Strain Expressing GnTV**

[0415] *P. pastoris* YSH-44 strain (**Example 15**) producing complex glycan structures was transformed with the plamid pPB140 (**Table 12**) containing a gene

fragment encoding the mouse GnTV catalytic domain ($\Delta 45$) ligated in frame to a targeting peptide derived from *S. cerevisiae MNN2(s)*. Culture conditions were same as in **Example 25**. The K3 reporter protein from two transformants were analyzed using MALDI-TOF. A peak at 1559 m/z [y] is consistent with the mass of the glycan GlcNAc₃Man₃GlcNAc₂ (**Figure 48**). A secondary peak at 1355 m/z [u] is consistent with the mass of GlcNAc₂Man₃GlcNAc₂.

EXAMPLE 28

P. pastoris Strain Producing Tetraantennary Structures on Glycoproteins

10 [0416] *P. pastoris* PBP43 strain (**Example 25**) producing triantennary glycan structures (e.g., GlcNAc₃Man₃GlcNAc₂) was transformed with the plasmid pPB140 (**Figure 40B**) encoding the mouse GnTV (**Example 27**). The vector pPB140 contains *KAN* resistance marker and *P. pastoris HIS3* gene as targeting sequence for genomic integration. 1 μ g plasmids were linearized with *KpnI*,
15 transformed into strain PBP43 by electroporation, selected on medium containing kanamycin and the resulting strains were screened by analysis of the released glycans from purified K3. Culture conditions were same as in **Example 25**. Analysis of the K3 reporter protein by MALDI-TOF showed a predominant peak at 1747 m/z [z], which is consistent with the mass of the tetraantennary glycan
20 GlcNAc₄Man₃GlcNAc₂ (**Figure 49**). Hexosaminidase digest (See **Example 15**) of the resulting glycans showed mass of the peak corresponding to Man₃GlcNAc₂ (data not shown).

[0417] In a second experiment, *P. pastoris* YSH-44 was transformed with pPB128 and pPB140 (**Table 12**). Analysis of the transformants producing the K3 reporter protein by MALDI-TOF showed a predominant peak at 1743 m/z [z],
25 which is consistent with the mass of the tetraantennary glycan GlcNAc₄Man₃GlcNAc₂ (**Figure 50**).

EXAMPLE 29

Cloning and Expression of GnTIX in *P. pastoris*

30 [0418] The nucleic acid and amino acid sequences of the human GnTIX (AB109185.1) are shown in **Figure 45**. The codon optimized DNA fragment encoding part of the human GnTIX lacking the TM domain ($\Delta 43$) was synthesized

from oligonucleotides using PCR (Figure 46). The DNA fragment encoding the GnTIX catalytic domain was ligated in frame to a targeting peptide derived from *S. cerevisiae MNN2(s)*. The resulting plasmid pPB176 (Figures 40C) was linearized with *KpnI* and transformed in *P. pastoris* YSH-44 strain (Example 15) producing complex tetraantennary glycan structures. Culture conditions were same as in Example 25. The K3 reporter protein from a transformant was analyzed using MALDI-TOF MS.

We claim:

1. A process for making a glycoprotein in a lower eukaryotic host cell comprising the step of introducing into the cell an *N*-acetylglucosaminyltransferase activity selected from the group consisting of: *N*-acetylglucosaminyltransferase IV activity, an *N*-acetylglucosaminyltransferase V activity, an *N*-acetylglucosaminyltransferase VI activity; and an *N*-acetylglucosaminyltransferase IX activity wherein the glycoprotein comprises at least three antennae on the trimannose core.

2. A process for making a glycoprotein in a lower eukaryotic host cell comprising the step of expressing in the cell one or more enzymatic activities that produce multiple antennary *N*-glycans comprising GlcNAc β 1,2 - Man α 1,6 (GlcNAc β 1,4 GlcNAc β 1,2 Man α 1,3) Man β 1,4 - GlcNAc β 1,4 - GlcNAc β 1,4 - Asn; GlcNAc β 1,4 GlcNAc β 1,2 - Man α 1,6 (GlcNAc β 1,4 GlcNAc β 1,2 Man α 1,3) Man β 1,4 - GlcNAc β 1,4 - GlcNAc β 1,4 - Asn; and GlcNAc β 1,6 GlcNAc β 1,4 GlcNAc β 1,2 - Man α 1,6 (GlcNAc β 1,4 GlcNAc β 1,2 Man α 1,3) Man β 1,4 - GlcNAc β 1,4 - GlcNAc β 1,4 - Asn structures.

3. The process of claims 1 or 2, wherein the activity is substantially intracellular.

4. The process of claims 1 or 2, further comprising the step of isolating the glycoprotein from the host cell.

5. The process of claims 1 or 2, wherein the host cell is selected from the group consisting of *Pichia pastoris*, *Pichia finlandica*, *Pichia trehalophila*, *Pichia kochiae*, *Pichia membranaefaciens*, *Pichia minutia*, *Ogataea minuta*, *Pichia lindneri*, *Pichia opuntiae*, *Pichia thermotolerans*, *Pichia salictaria*, *Pichia guercuum*, *Pichia piperi*, *Pichia stipitis*, *Pichia methanolica*, *Pichia* sp., *Saccharomyces cerevisiae*, *Saccharomyces* sp., *Hansenula polymorpha*, *Kluyveromyces* sp., *Kluyveromyces lactis*, *Candida albicans*, *Aspergillus nidulans*, *Aspergillus niger*, *Aspergillus oryzae*, *Trichoderma reesei*, *Chrysosporium*

lucknowense, *Fusarium* sp., *Fusarium gramineum*, *Fusarium venenatum*, and *Neurospora crassa*.

6. The process of claim 5, wherein the host cell is selected from the group consisting of *Pichia pastoris*, *Pichia finlandica*, *Pichia trehalophila*, *Pichia koclamae*, *Pichia membranaefaciens*, *Pichia minutia*, *Ogataea minuta*, *Pichia lindneri*, *Pichia opuntiae*, *Pichia thermotolerans*, *Pichia salictaria*, *Pichia guercuum*, *Pichia pijperi*, *Pichia stipitis*, *Pichia methanolica*, and *Pichia* sp.

7. The process of claim 6, wherein the host cell is *Pichia pastoris*.

8. The process of claims 1 or 2 wherein the glycoprotein is a therapeutic protein.

9. The process of claim 8, wherein the therapeutic protein is selected from the group consisting of kringle domains of human plasminogen, erythropoietin, cytokines, coagulation factors, soluble IgE receptor α -chain, IgG, IgG fragments, IgM, interleukins, urokinase, chymase, urea trypsin inhibitor, IGF-binding protein, epidermal growth factor, growth hormone-releasing factor, annexin V fusion protein, angiostatin, vascular endothelial growth factor-2, myeloid progenitor inhibitory factor-1, osteoprotegerin, α -1 antitrypsin, DNase II, α -feto proteins, FSH and peptide hormones.

10. A lower eukaryotic host cell comprising an *N*-acetylglucosaminyltransferase IV, V, VI, or IX activity.

11. A lower eukaryotic host cell comprising an *N*-acetylglucosaminyltransferase IV activity and an *N*-acetylglucosaminyltransferase V activity.

12. The host cell of claim 10 or 11, wherein the activity is substantially intracellular.

13. The host cell of claim 10 or 11, wherein the cell produces N-glycans comprising GlcNAc₂Man₃GlcNAc₂ structures that are capable of reacting with GnTIV, GnTV or GnT VI activity.

14. A lower eukaryotic host cell comprising an N-glycan that comprises at least three GlcNAcs on a Man₃GlcNAc₂ oligosaccharide.

15. The host cell of claim 14, wherein the N-glycan contains at least 50, 60, 70, 80, 90 mole % or greater triantennary glycans.

16. The host cell of claim 14, wherein the N-glycan contains at least 70, 80, 90 mole % or greater tetraantennary glycans.

17. A lower eukaryotic host cell comprising multiple antennary N-glycans including GlcNAc β 1,2 - Man α 1,6 (GlcNAc β 1,4 GlcNAc β 1,2 Man α 1,3) Man β 1,4 - GlcNAc β 1,4 - GlcNAc β 1,4 – Asn; GlcNAc β 1,4 GlcNAc β 1,2 - Man α 1,6 (GlcNAc β 1,4 GlcNAc β 1,2 Man α 1,3) Man β 1,4 - GlcNAc β 1,4 - GlcNAc β 1,4 – Asn; and GlcNAc β 1,6 GlcNAc β 1,4 GlcNAc β 1,2 - Man α 1,6 (GlcNAc β 1,4 GlcNAc β 1,2 Man α 1,3) Man β 1,4 - GlcNAc β 1,4 - GlcNAc β 1,4 – Asn structures.

18. The host cell of claims 10, 11, 14, or 17, wherein the host cell is selected from the group consisting of *Pichia pastoris*, *Pichia finlandica*, *Pichia trehalophila*, *Pichia koclamae*, *Pichia membranaefaciens*, *Pichia minutia*, *Ogataea minuta*, *Pichia lindneri*, *Pichia opuntiae*, *Pichia thermotolerans*, *Pichia salictaria*, *Pichia guercuum*, *Pichia pijperi*, *Pichia stiptis*, *Pichia methanolica*, *Pichia* sp., *Saccharomyces cerevisiae*, *Saccharomyces* sp., *Hansenula polymorpha*, *Kluyveromyces* sp., *Candida albicans*, *Aspergillus nidulans*, *Aspergillus niger*, *Aspergillus oryzae*, *Trichoderma reesei*, *Chrysosporium lucknowense*, *Fusarium* sp., *Fusarium gramineum*, *Fusarium venenatum*, and *Neurospora crassa*.

19. The host cell of claim 18, wherein the host cell is selected from the group consisting of *Pichia pastoris*, *Pichia finlandica*, *Pichia trehalophila*, *Pichia koclamae*, *Pichia membranaefaciens*, *Pichia minutia*, *Ogataea*

minuta, *Pichia lindneri*, *Pichia opuntiae*, *Pichia thermotolerans*, *Pichia salictaria*, *Pichia guercuum*, *Pichia pijperi*, *Pichia stiptis*, *Pichia methanolica*, and *Pichia* sp..

20. The host cell of claim 19, wherein the host cell is *Pichia pastoris*.

21. A lower eukaryotic host cell comprising a trimannose core ($\text{GlcNAc}_2\text{Man}_3\text{GlcNAc}_2$) structure or a Man₅ core structure ($\text{GlcNAc}\text{Man}_5\text{GlcNAc}_2$) modified by a GlcNAc β 1,4 or a GlcNAc β 1,6 transferase.

22. The host cell of claim 21, wherein the cell produces greater than 70 mole % of the modified structure.

23. The host cell of claim 21, wherein the cell produces greater than 90 mole % of the modified structure.

24. A lower eukaryotic host cell comprising an *N*-acetylglucosaminyltransferase I activity and an *N*-acetylglucosaminyltransferase IV activity.

25. The host cell of claim 24, wherein the activities are substantially intracellular.

26. The host cell of claim 24, wherein the cell produces *N*-glycans comprising $\text{GlcNAc}_2\text{Man}_3\text{GlcNAc}_2$ that are capable of reacting with GnTV, GnTVI or GnTIX activity.

27. The host cell of claim 24, wherein the *N*-acetylglucosaminyltransferase activities produces a multiple antennary glycan.

28. A lower eukaryotic host cell comprising a GnTIV, V, VI or IX activity and a mannosidase II activity.

29. The host cell of claim 28, further comprising an *N*-acetylglucosaminyltransferase I activity.

30. The host cell of claim 29, further comprising an *N*-acetylglucosaminyltransferase II activity.
31. The host cell of claim 28, further comprising an *N*-acetylglucosaminyltransferase I activity and an *N*-acetylglucosaminyltransferase II activity.
32. The host cell of any one of claims 10, 11, 24, or 28 that is deficient in an 1,6 mannosyltransferase activity.
33. The host cell of any one of claims 10, 11, 24, or 28 that is deficient in a Dol-P-Man:Man₅GlcNAc₂-PP-Dol mannosyltransferase activity.
34. The host cell of any one of claims 10, 11, 24, or 28 further comprising an α -1,2-mannosidase I activity.
35. The host cell of any one of claims 10, 11, 24, or 28 further comprising a UDP-GlcNAc transporter.
36. The glycoprotein made by the process of claim 1.
37. The glycoprotein made by the process of claim 2.
38. The glycoprotein made by the process of claim 5.
39. The glycoprotein made by the process of claim 6.
40. The glycoprotein made by the process of claim 7.
41. The glycoprotein made by the process of claim 8.
42. The glycoprotein made by the process of claim 9.
43. A glycoprotein comprising at least three GlcNAc β 1,4 residues or at least three GlcNAc β 1,6 residues on a GlcNAc₂Man₃GlcNAc₂ core structure produced in a lower eukaryotic host cell.

44. A glycoprotein comprising a GlcNAc β 1,4 residue attached to either Man α 1,3 or Man α 1,6 arm of the substrates GlcNAc₂Man₃GlcNAc₂, GlcNAc₃Man₃GlcNAc₂, GlcNAcMan₃GlcNAc₂, Man₃GlcNAc₂, GlcNAcMan₅GlcNAc₂ or a Man₅GlcNAc₂ core structure produced in a lower eukaryotic host cell.

45. The glycoprotein of claim 44, wherein greater than 70 mole % of the core structures are modified by GnTIV.

46. The glycoprotein of claim 44, wherein greater than 50 mole % of the core structures are modified by GnTV.

47. A pharmaceutical composition comprising the glycoprotein of any one of claims 36 - 46.

48. A vector capable of expressing GnTIV, GnTV, GnTVI, GnTIX activity in a lower eukaryotic host.

49. A lower eukaryotic host cell comprising *N*-glycans having at least two GlcNAc residues on either the Man α 1,3 or Man α 1,6 arm of the trimannose core oligosaccharide intermediate.

50. The lower eukaryotic host cell of claim 49, further comprising two GlcNAc β 1,4 residues on the Man α 1,3 and Man α 1,6 arm of the trimannose core oligosaccharide intermediate.

51. The lower eukaryotic host cell of claim 49, further comprising two GlcNAc β 1,6 residues on the Man α 1,3 and Man α 1,6 arm of the trimannose core oligosaccharide intermediate.

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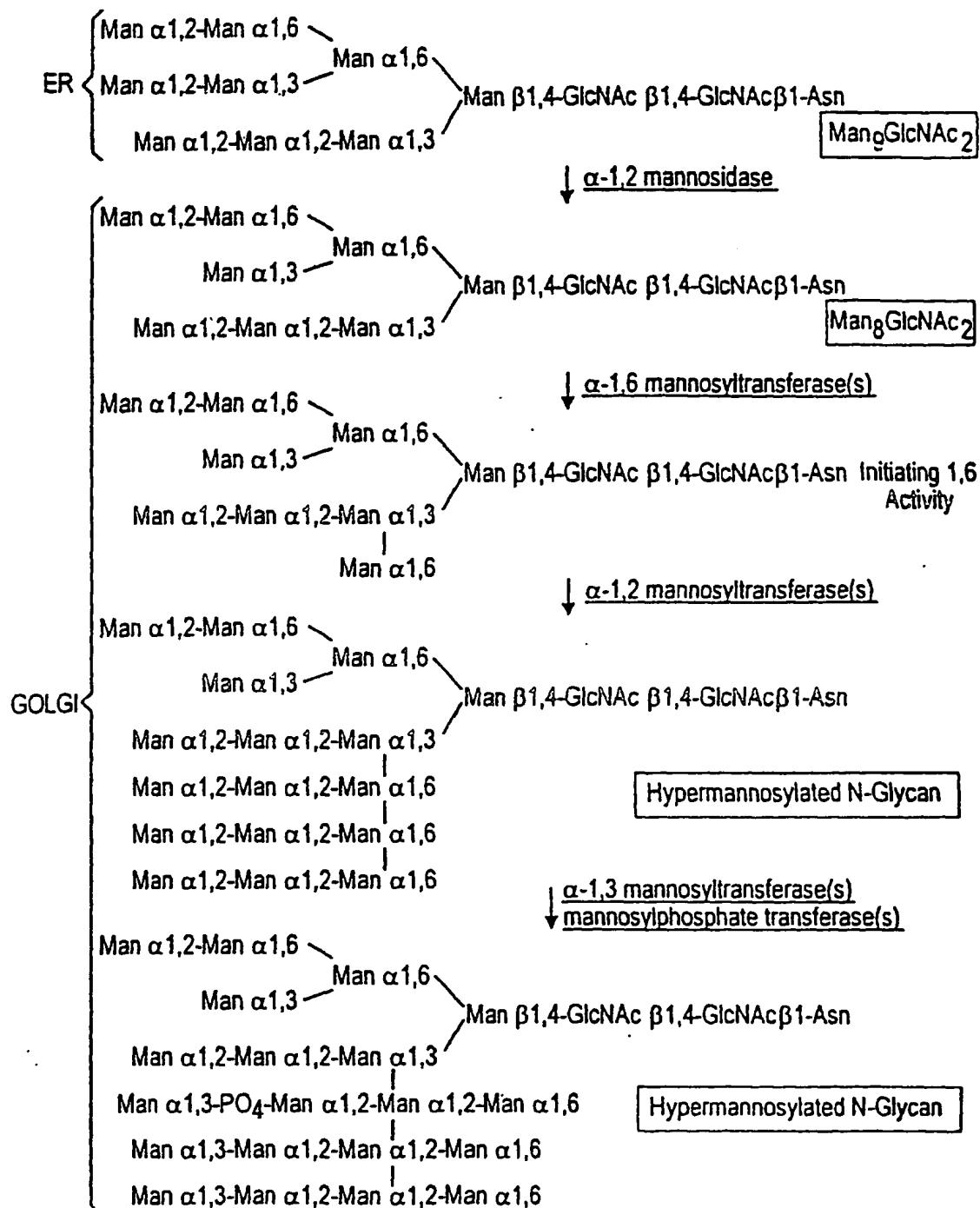


FIG. 1A

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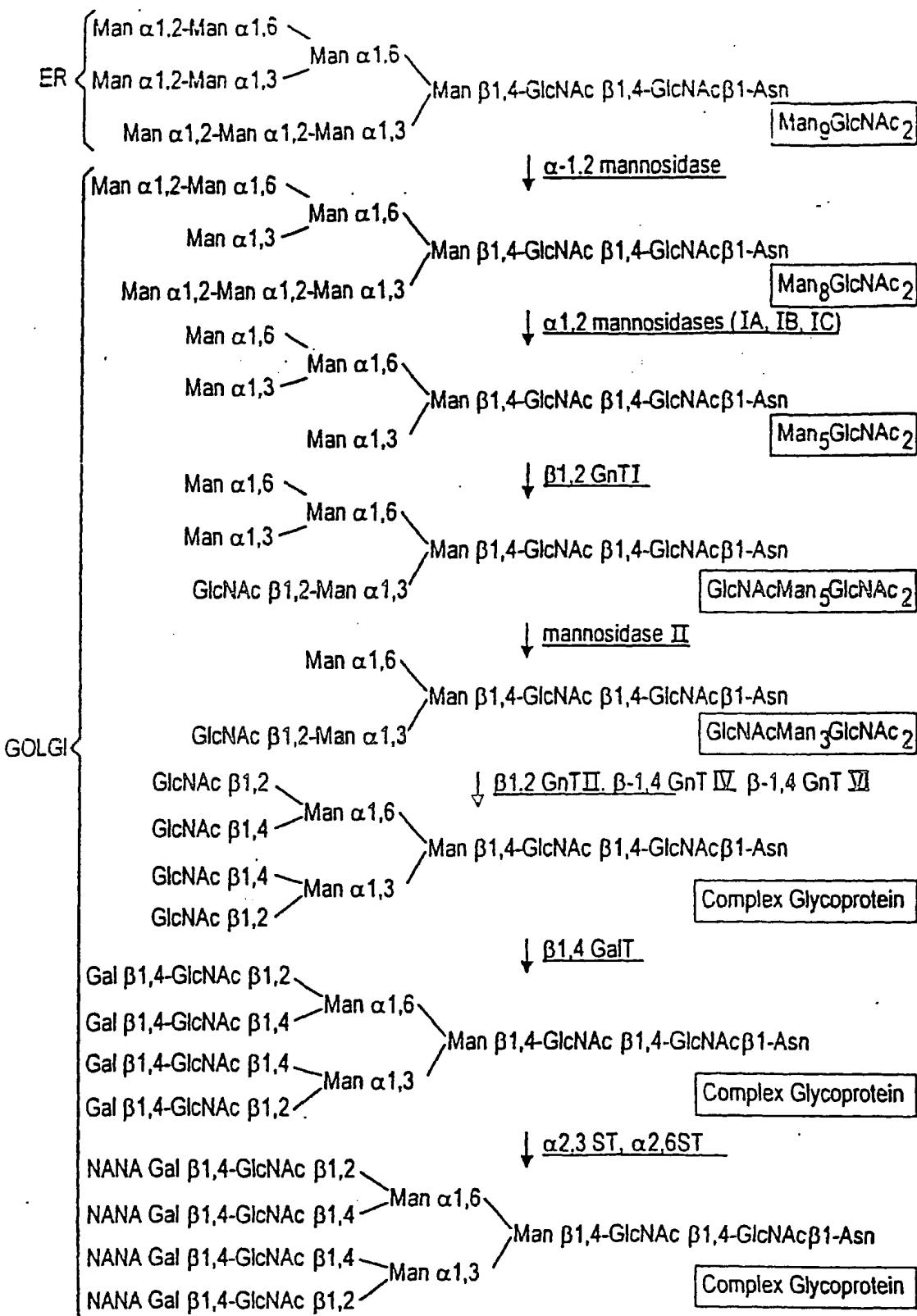
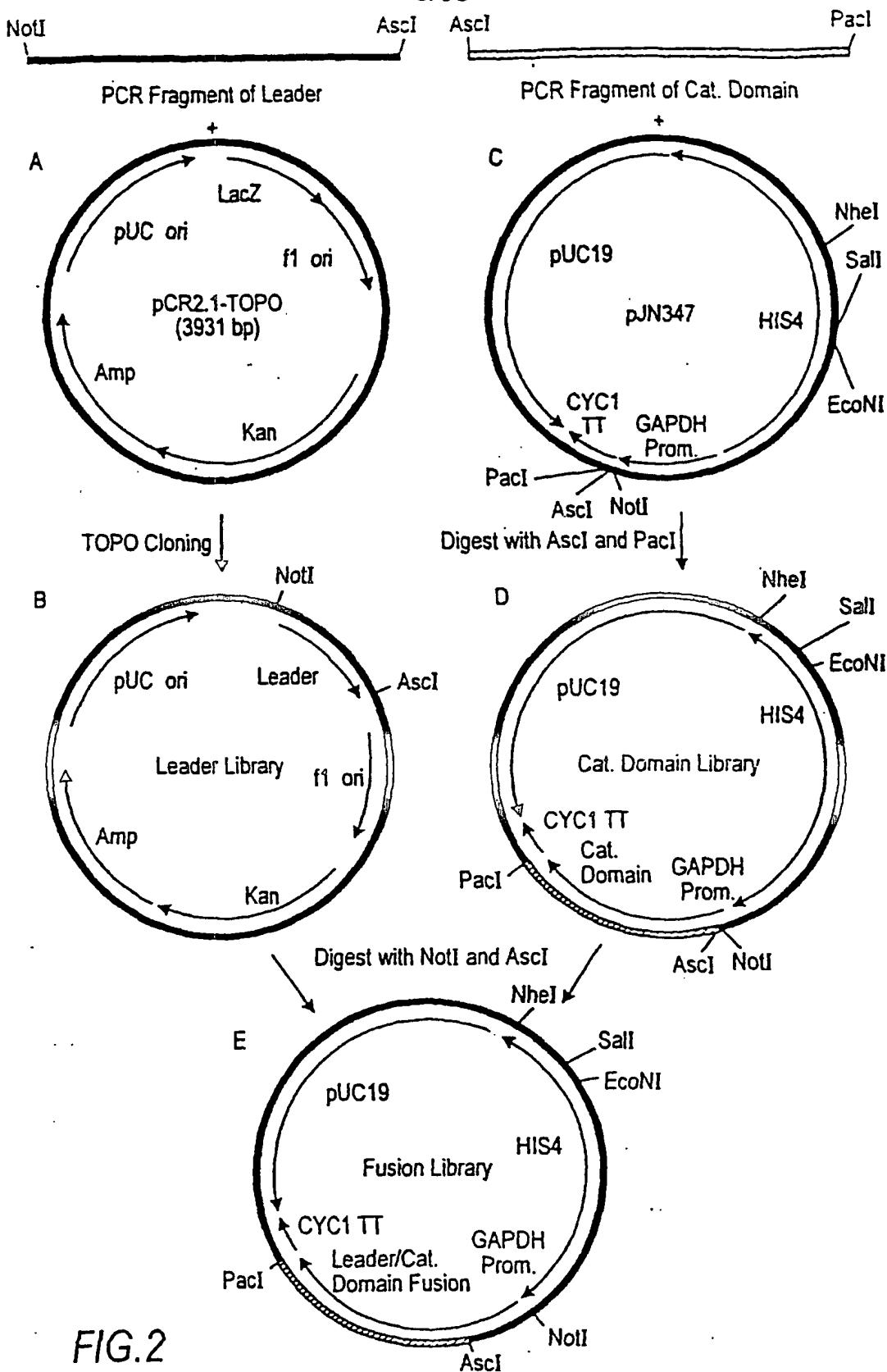


FIG. 1B

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M. musculus alpha-1,2-mannosidase I A open reading frame. The transmembrane and catalytic domains are highlighted in bold respectively. The sequence of the primers used to generate the N-terminal truncations are highlighted by underlining and the start of each respective protein fragment indicated by an arrow.

FIG. 3

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FIG. 3 CON'T

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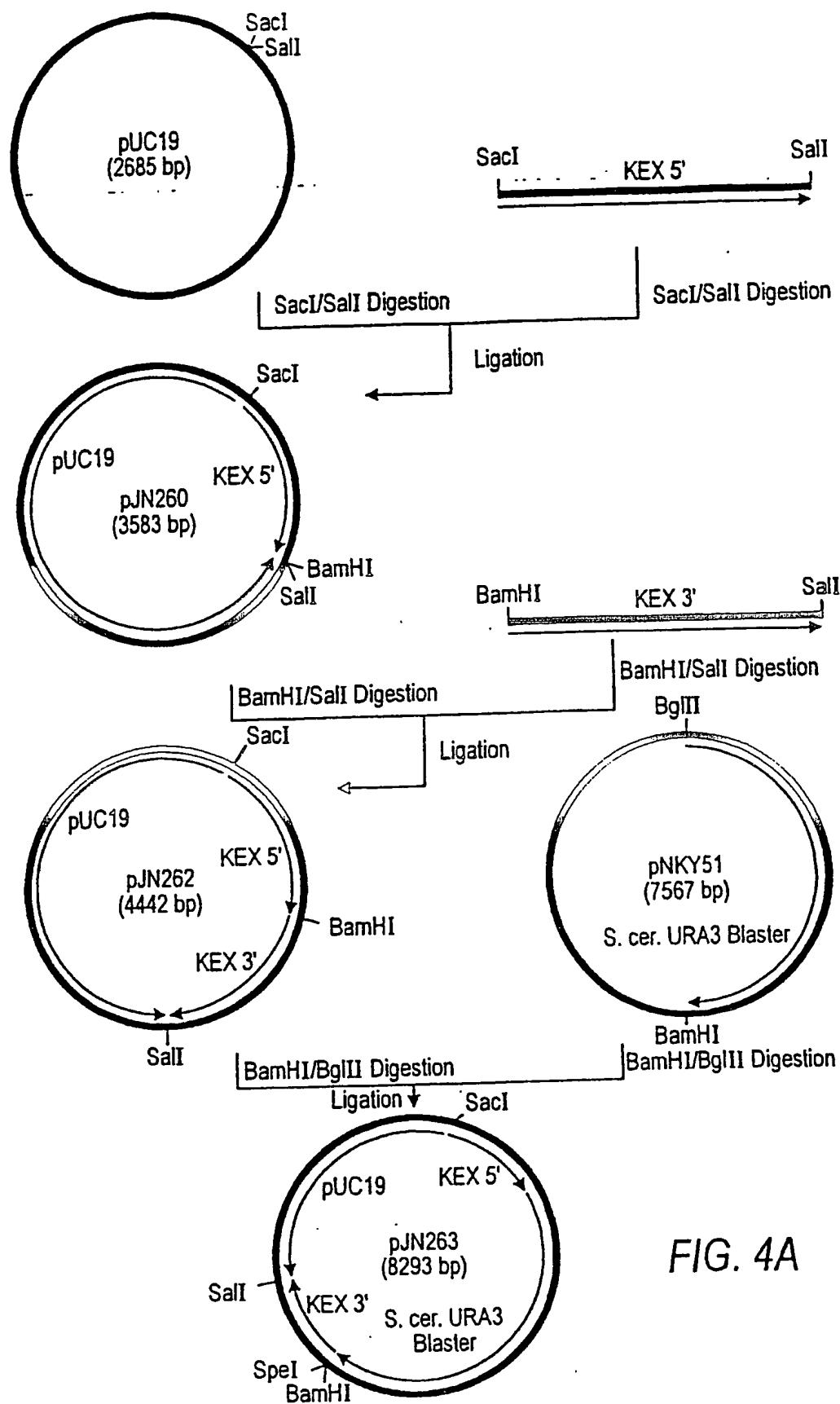


FIG. 4A

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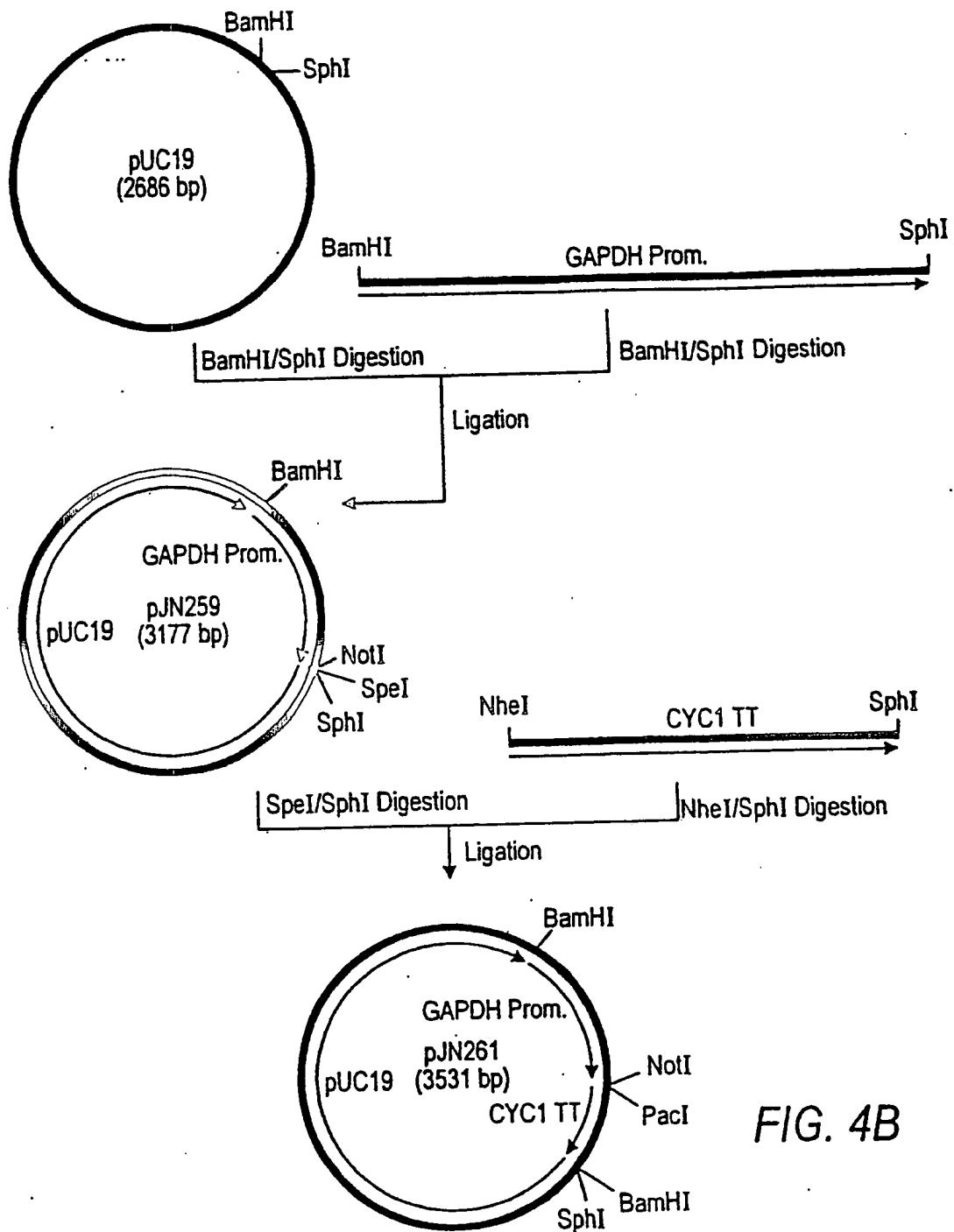
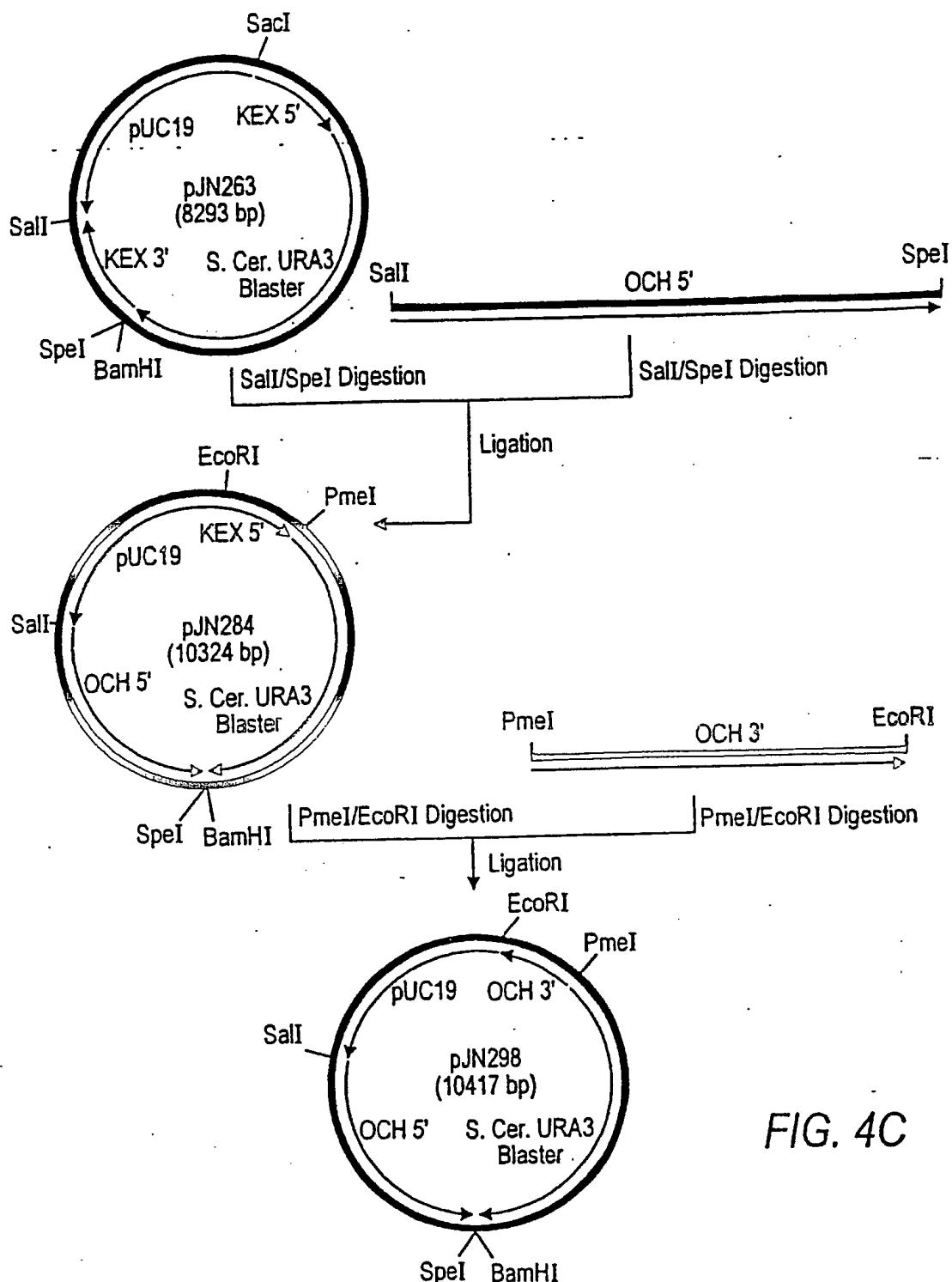


FIG. 4B

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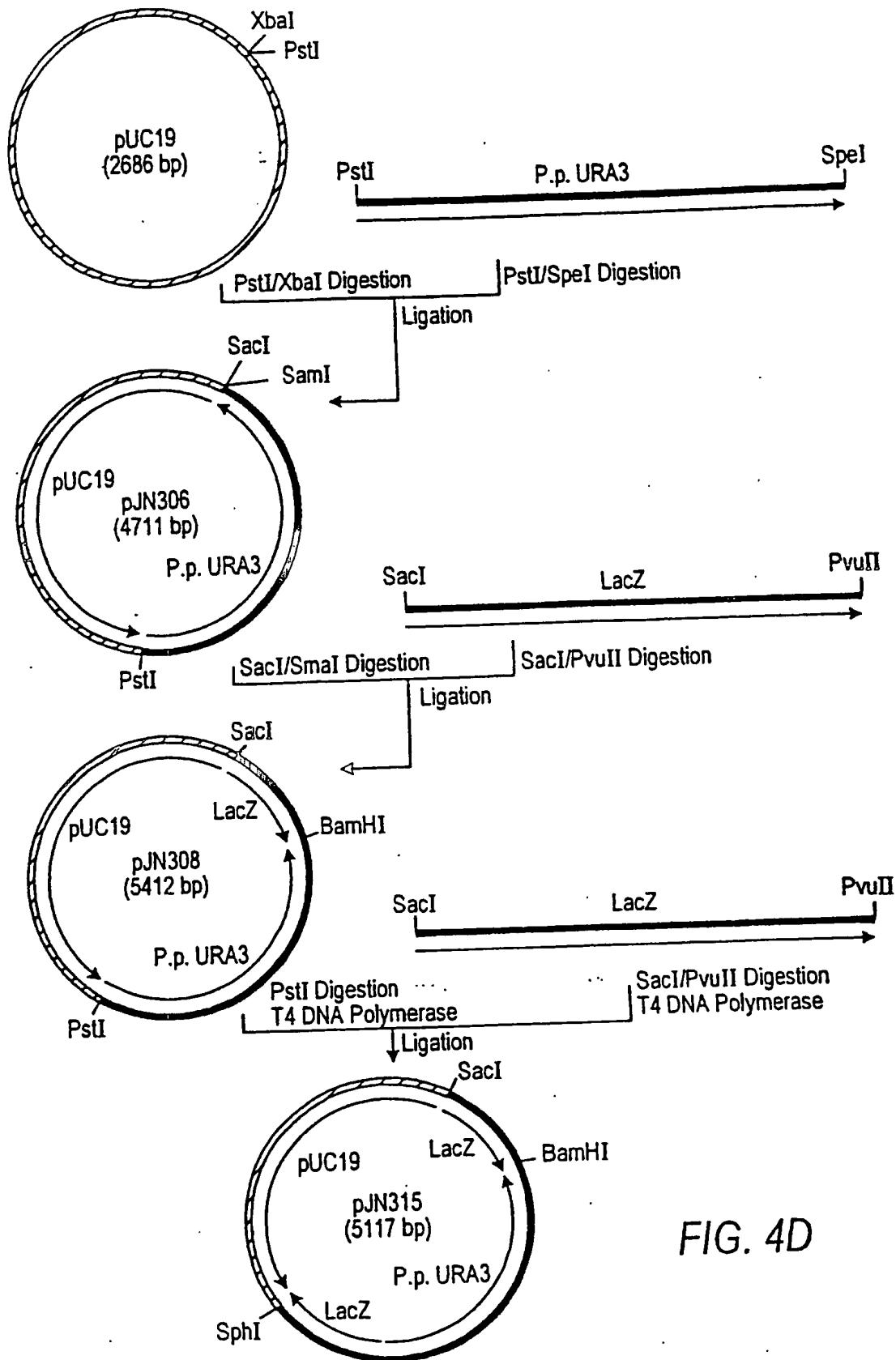


FIG. 4D

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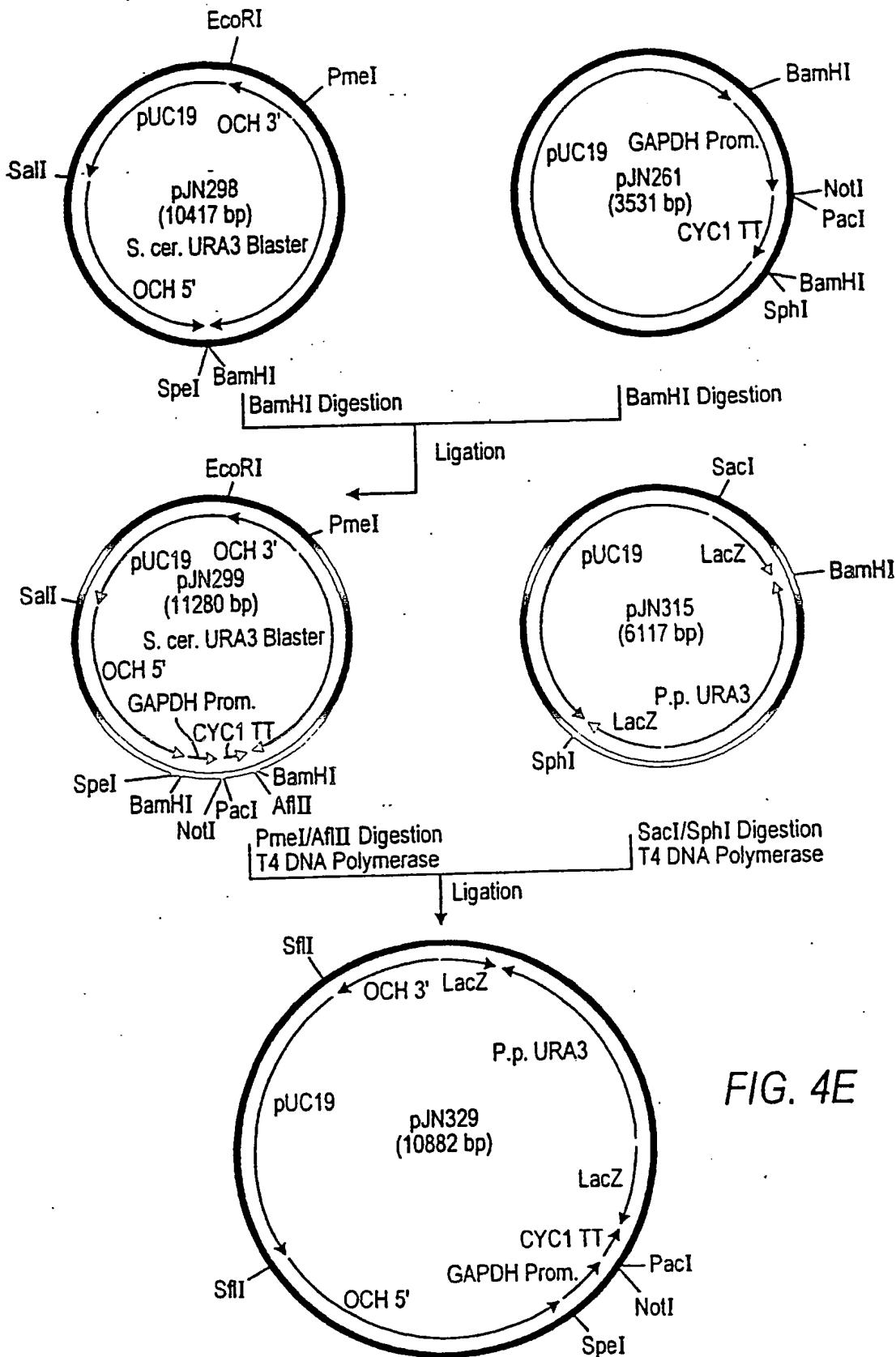
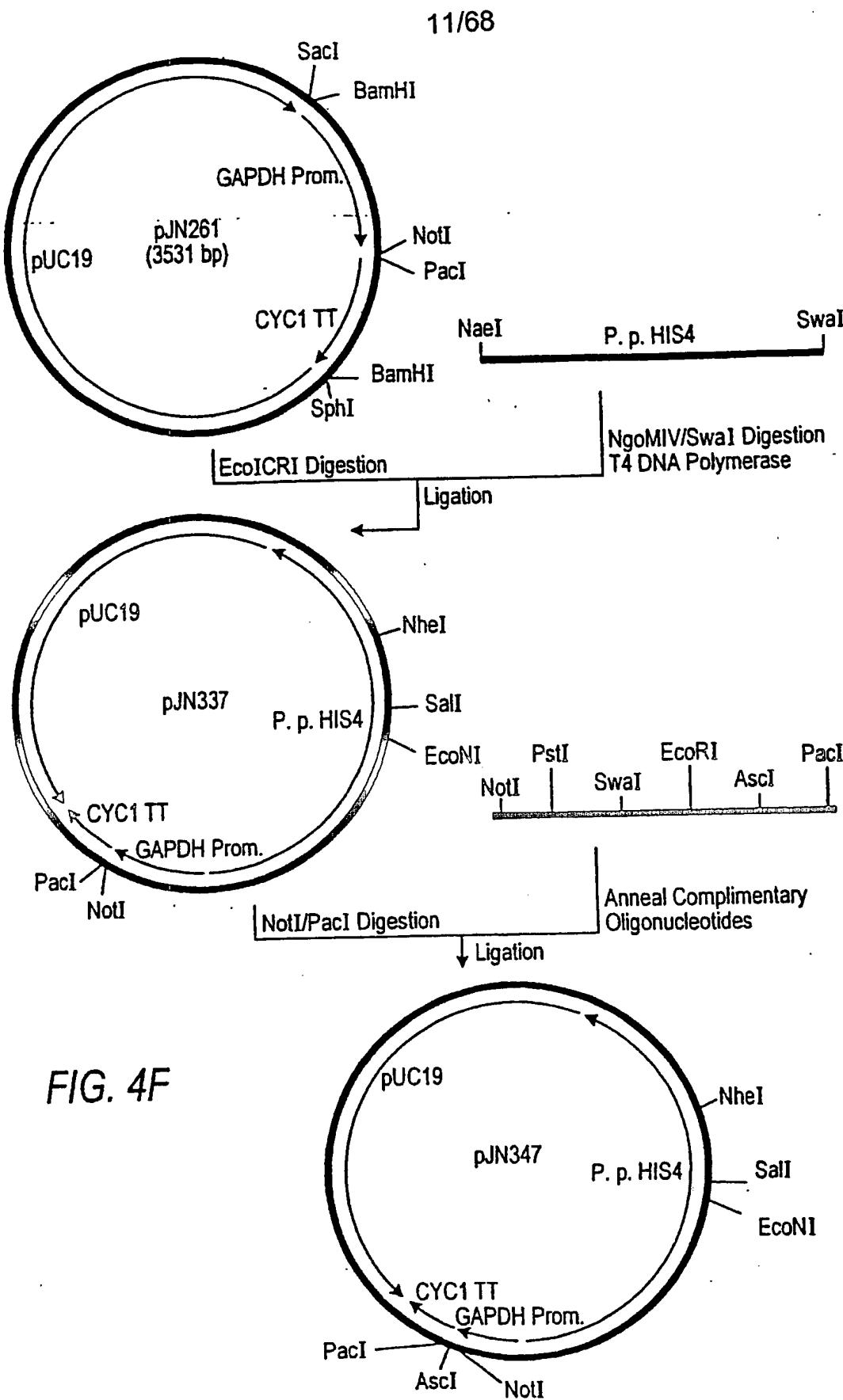


FIG. 4E



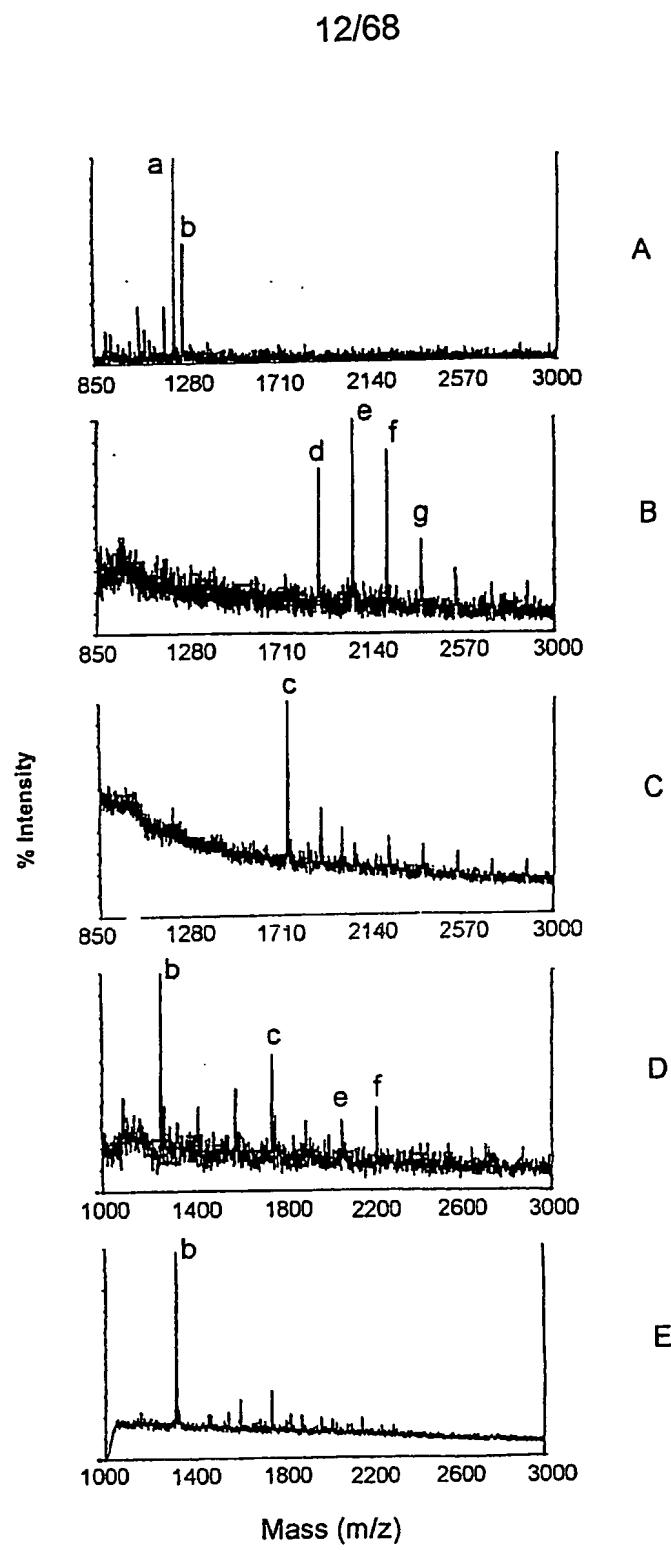
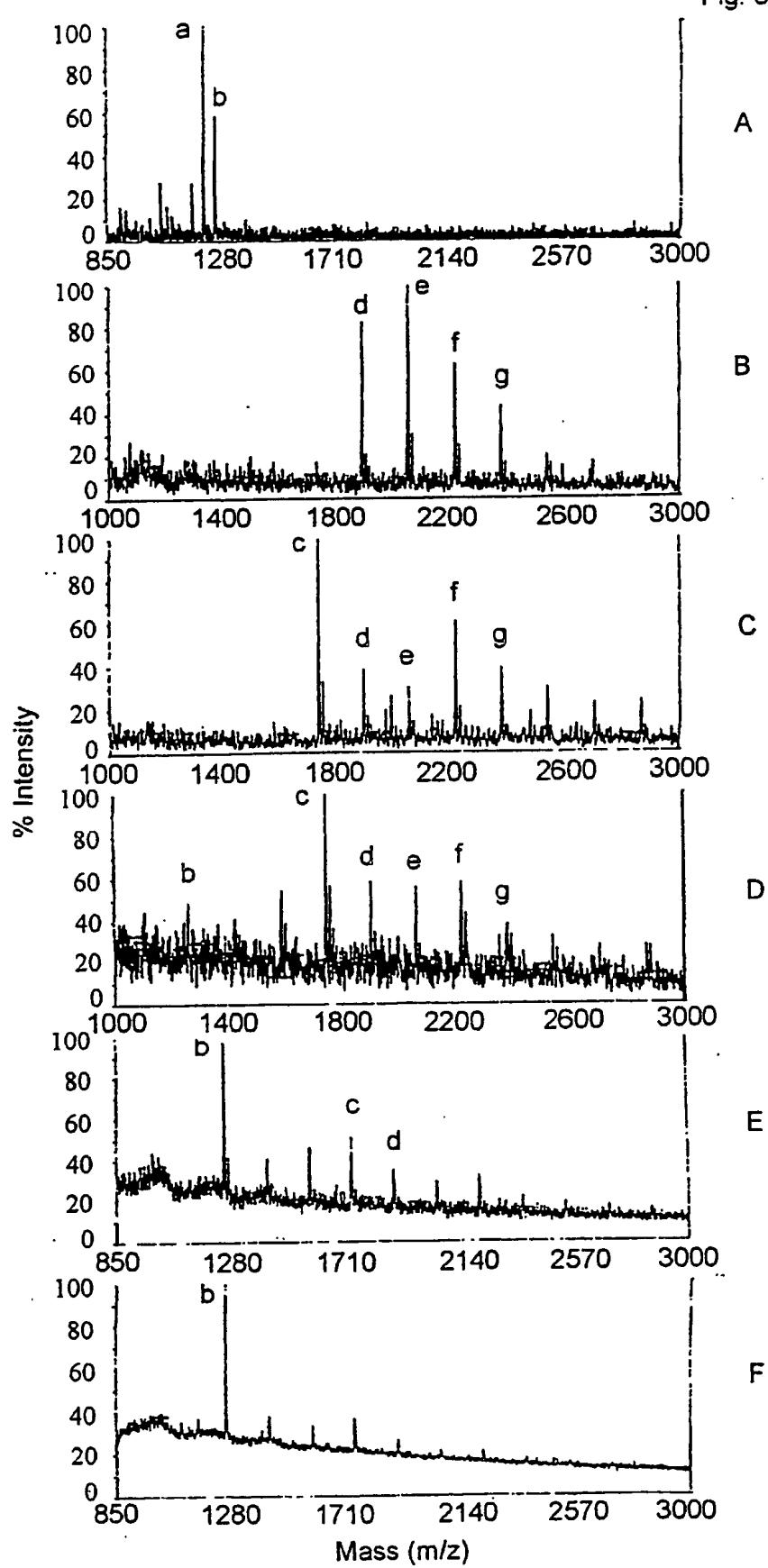


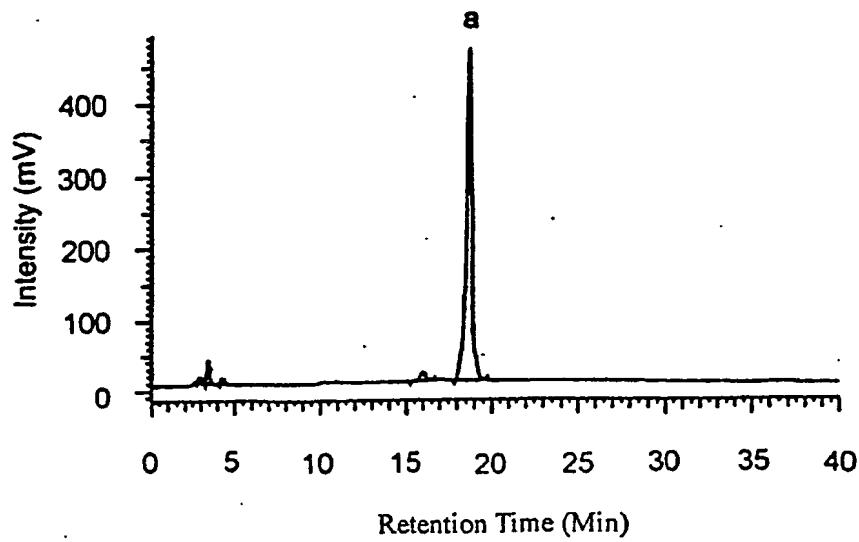
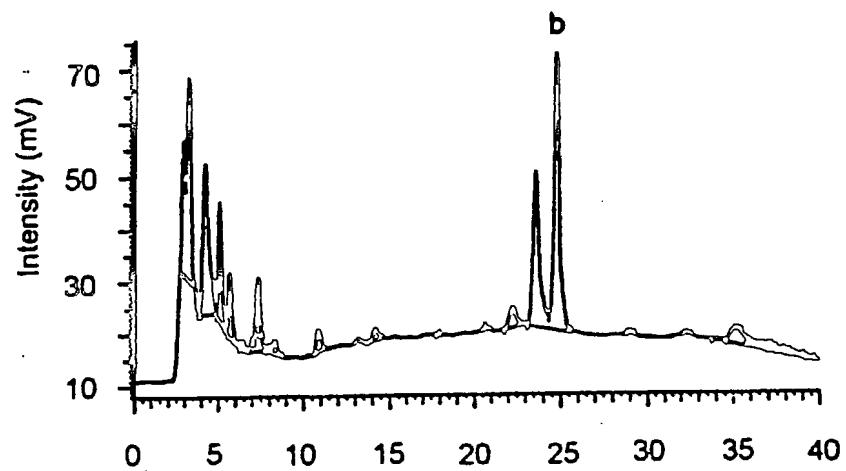
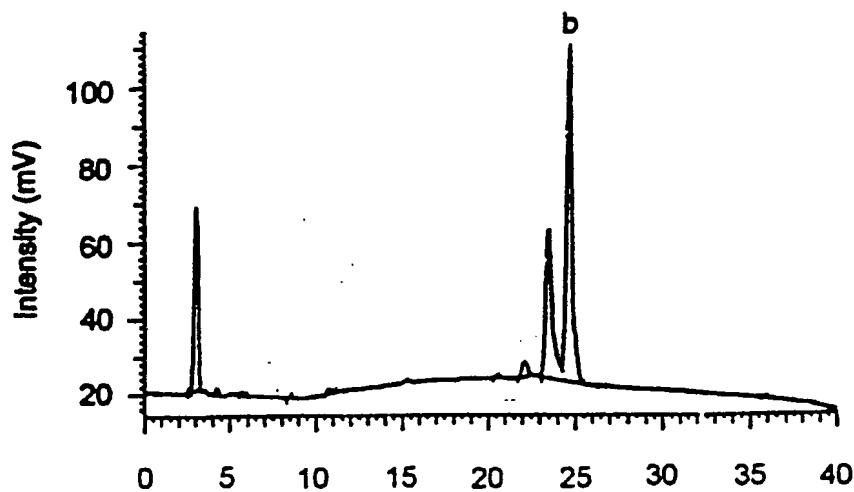
Fig. 5

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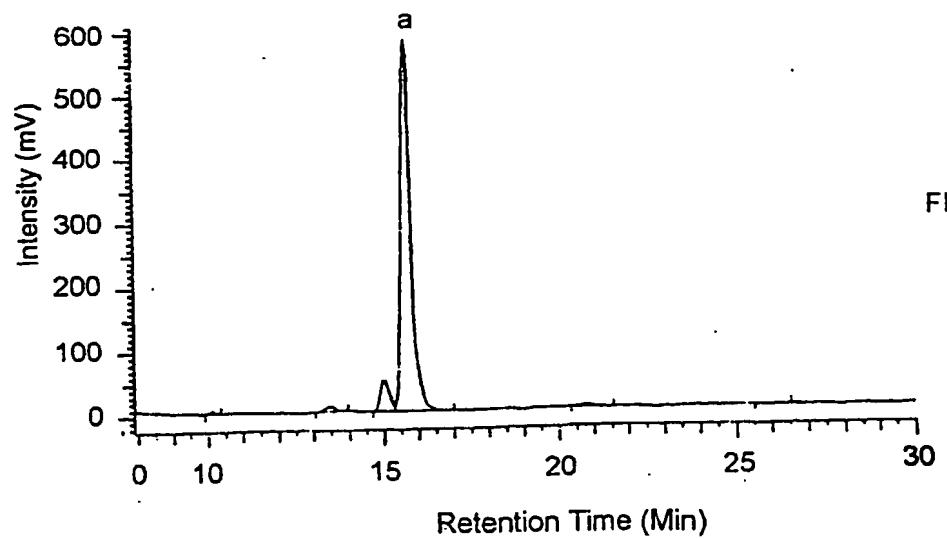
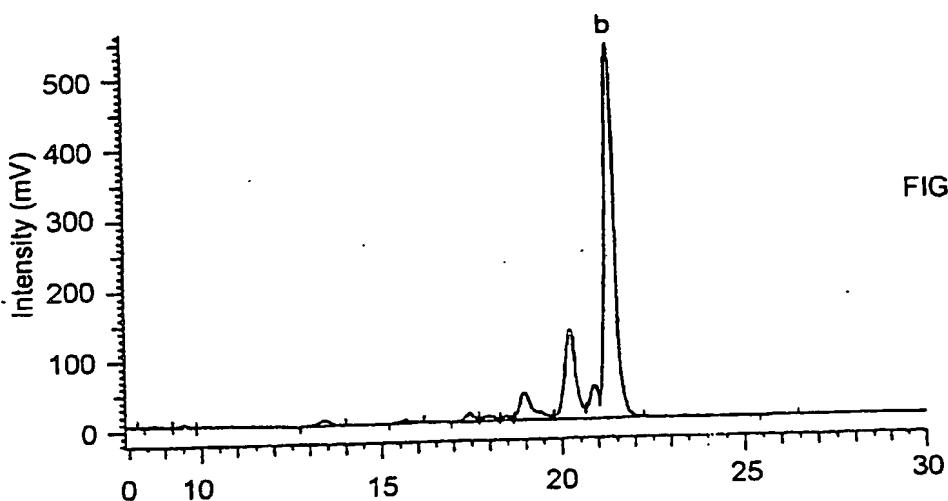
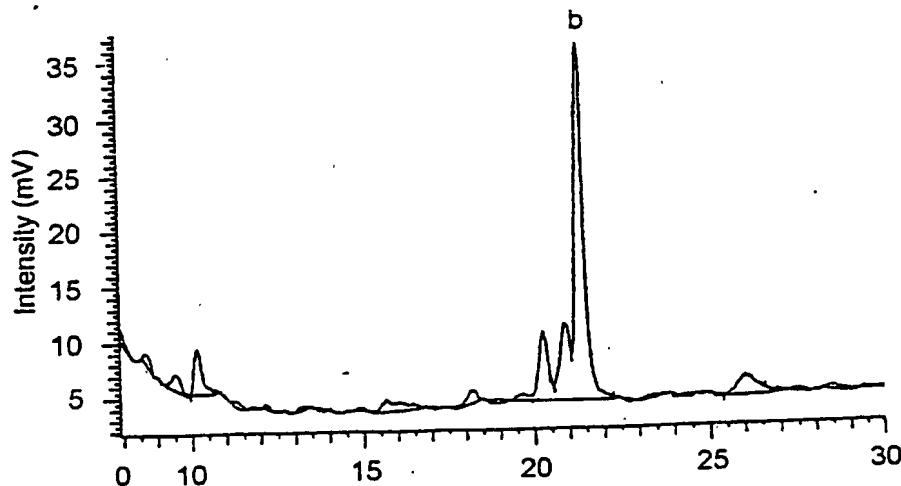
Fig. 6



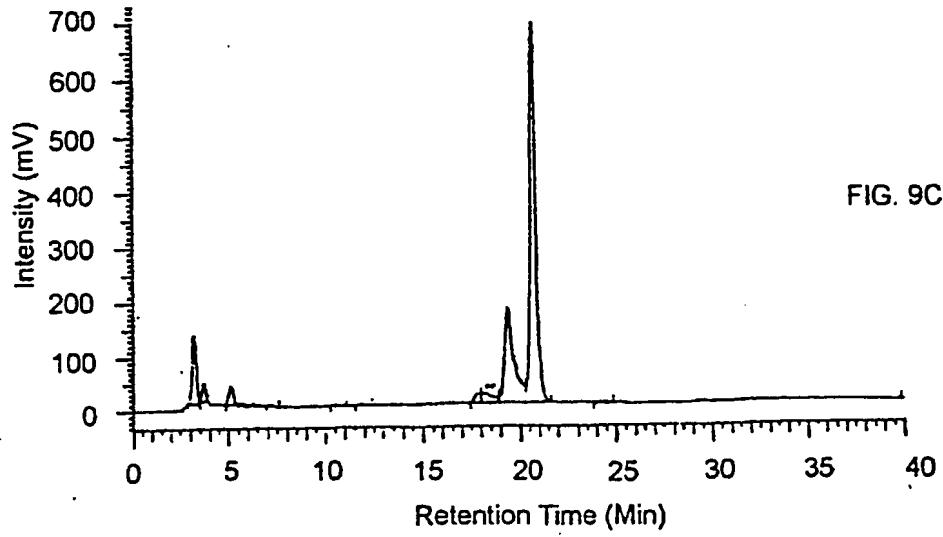
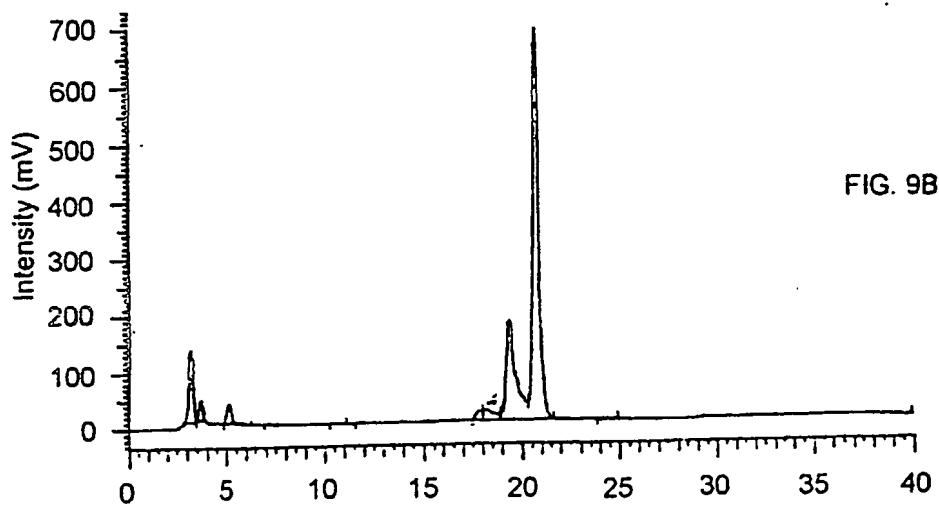
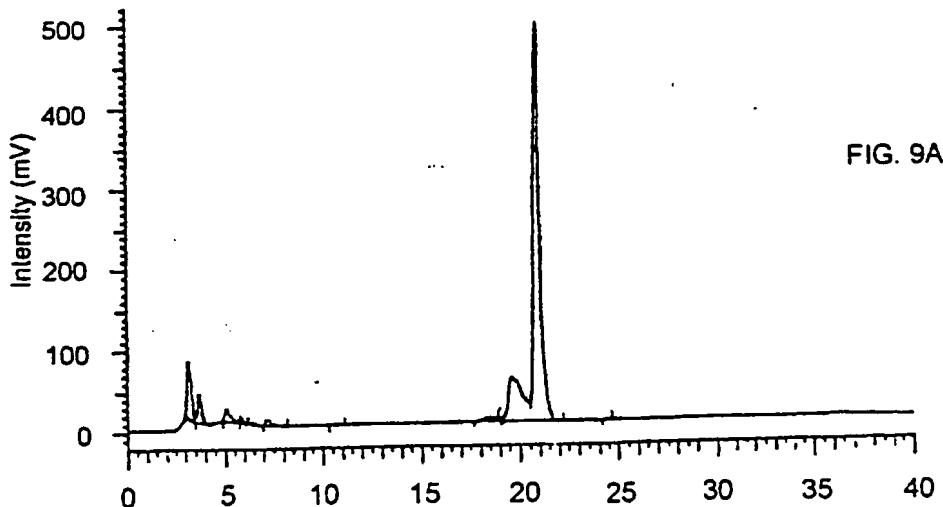
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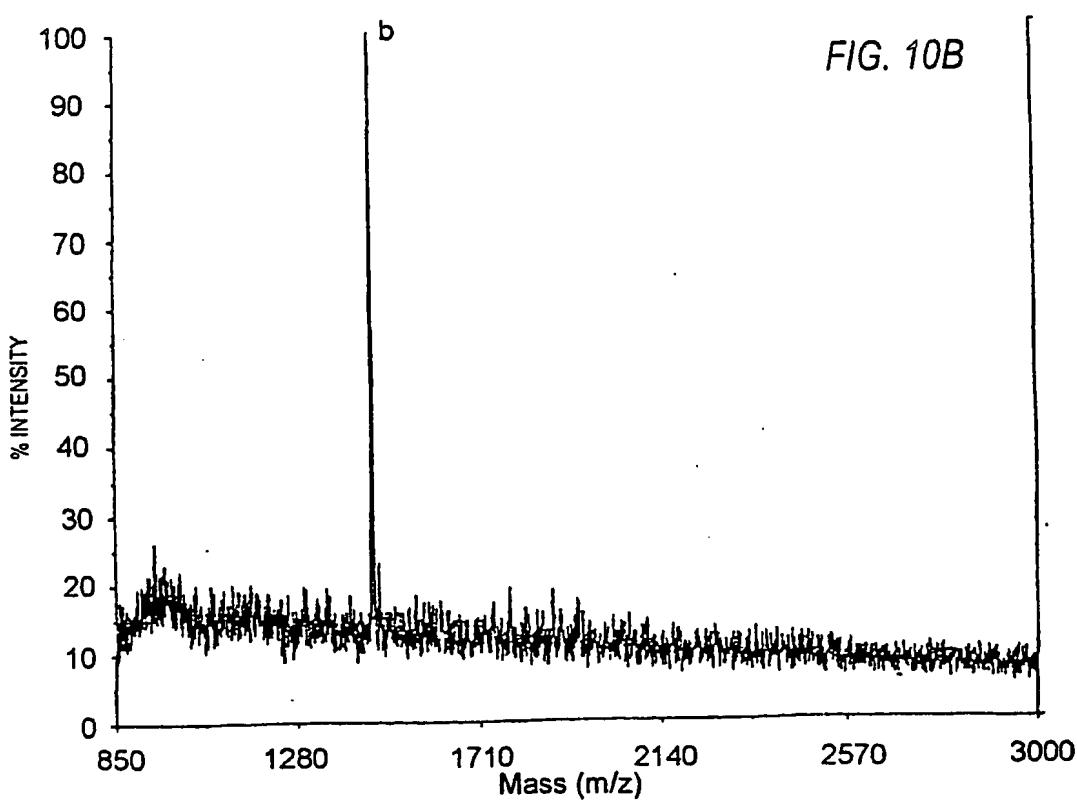
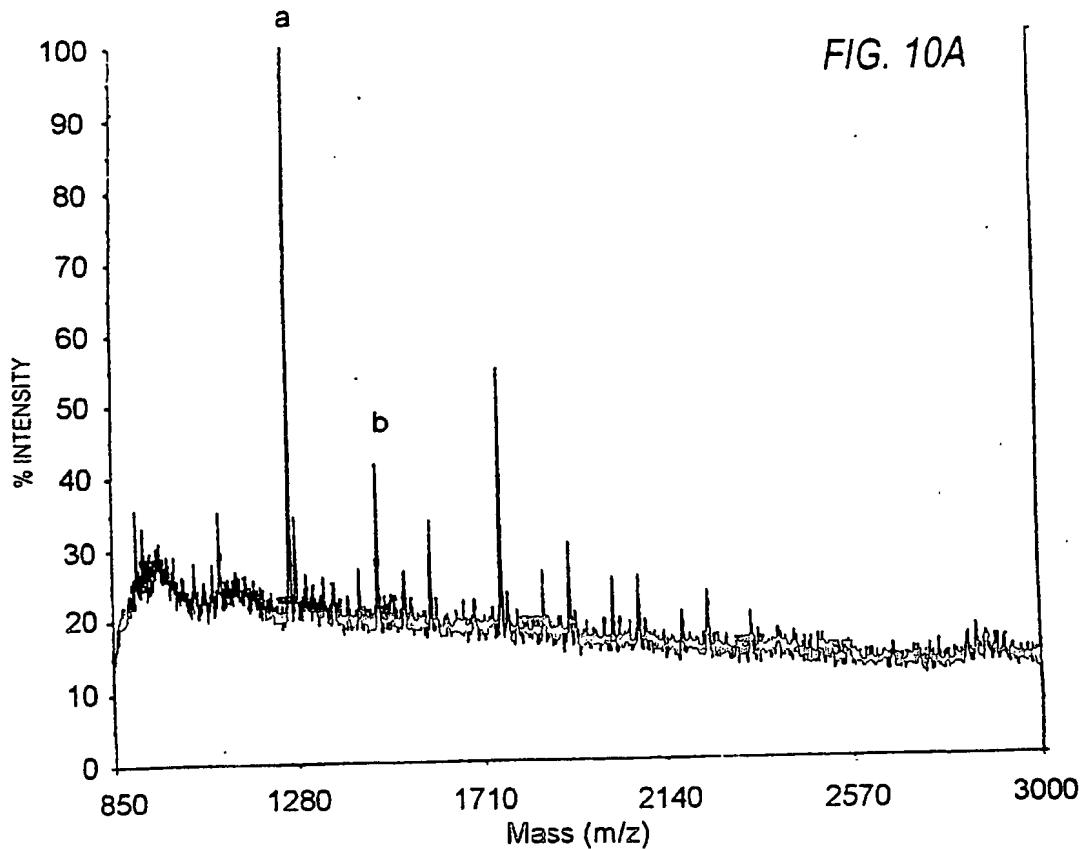
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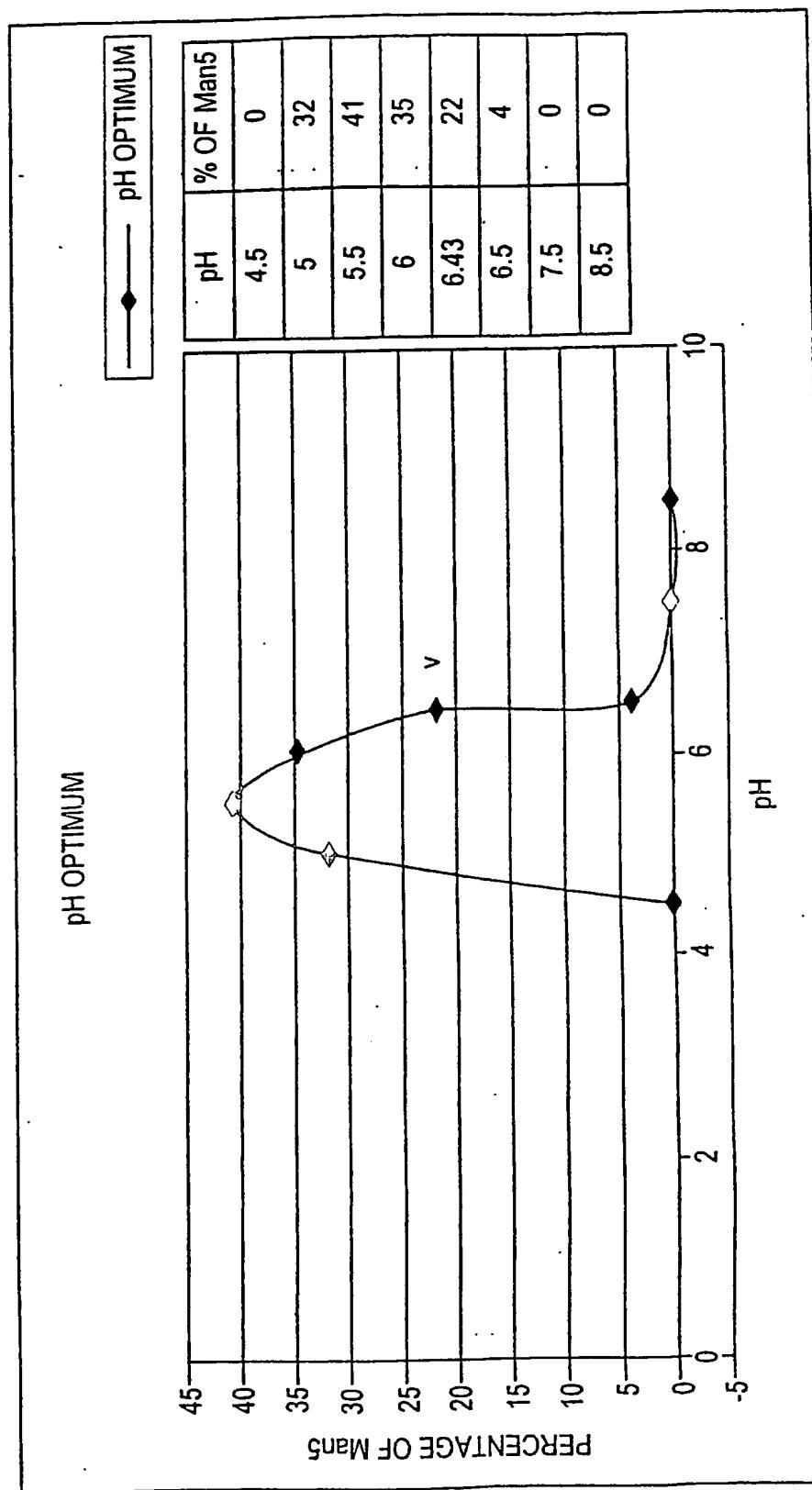
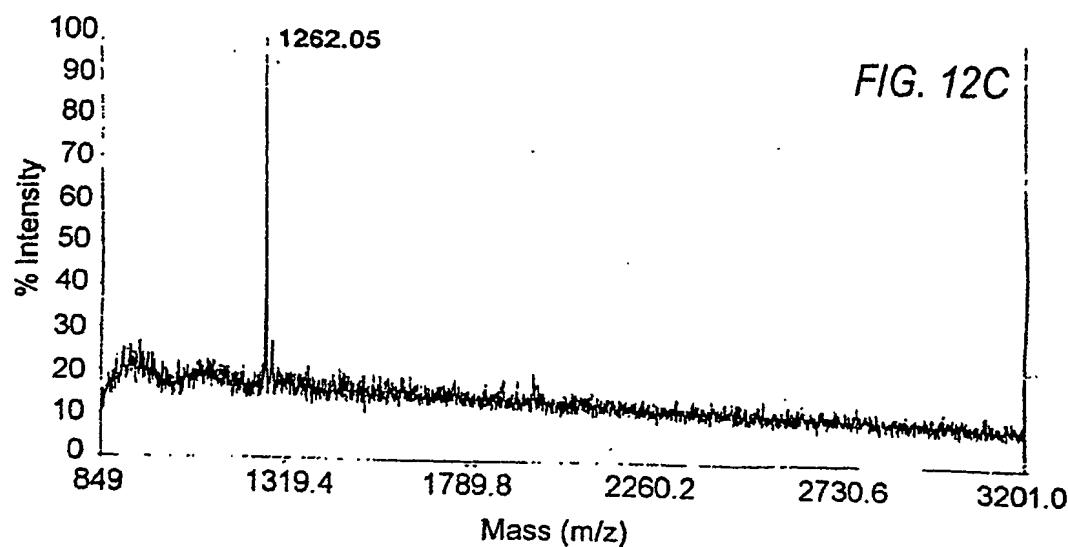
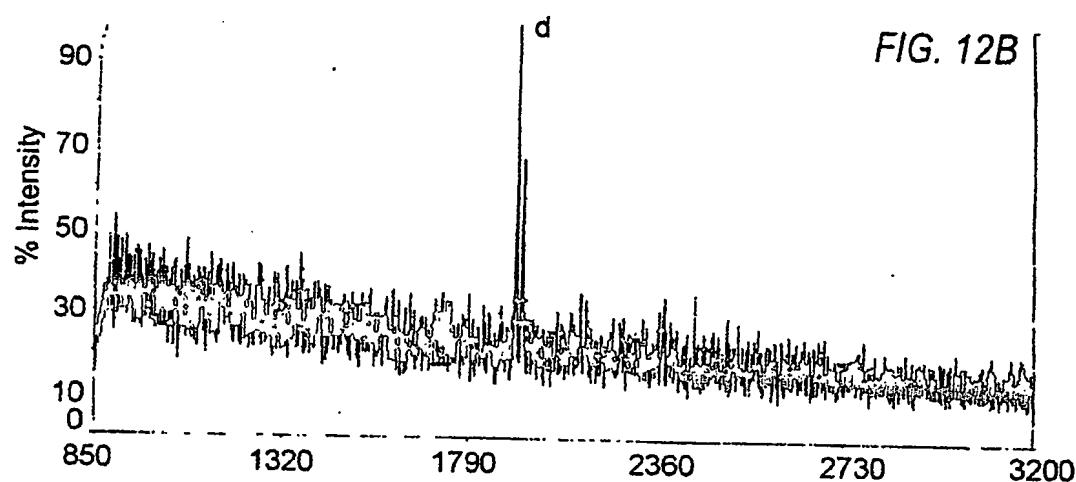
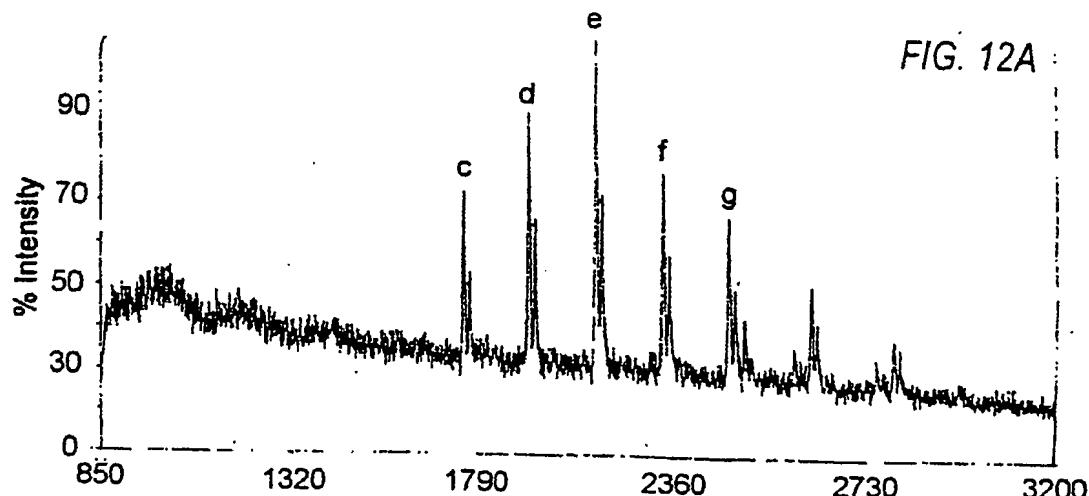


FIG. 11

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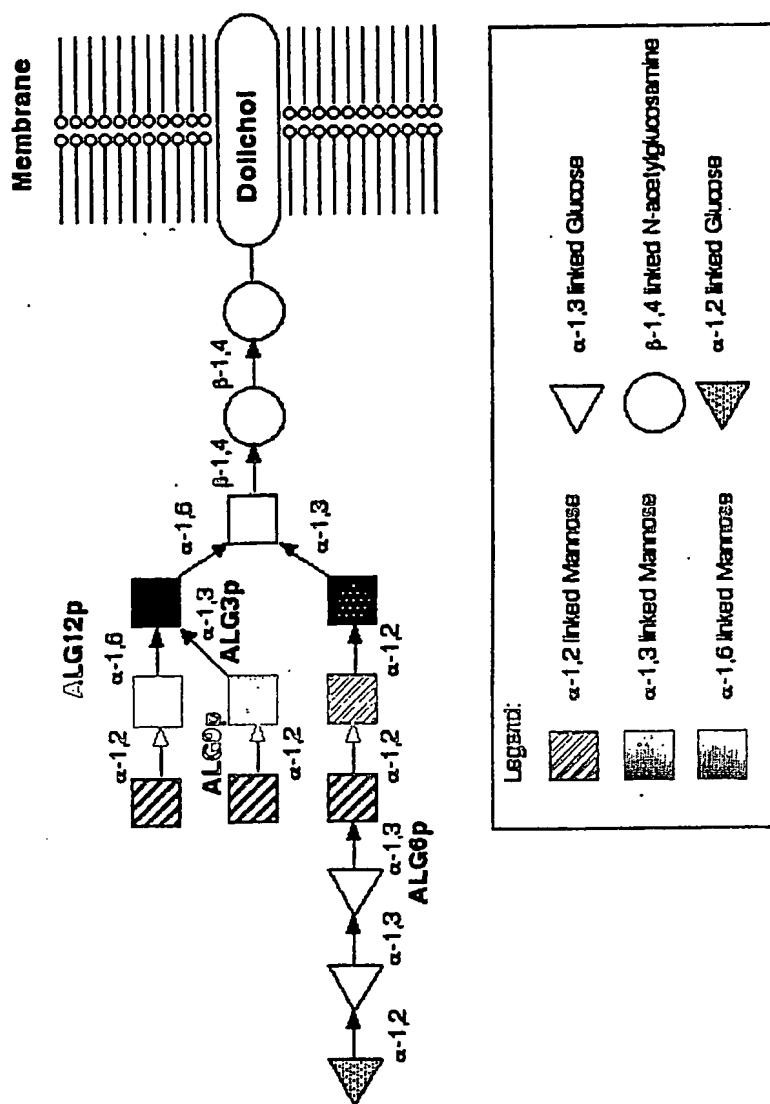


FIG. 13

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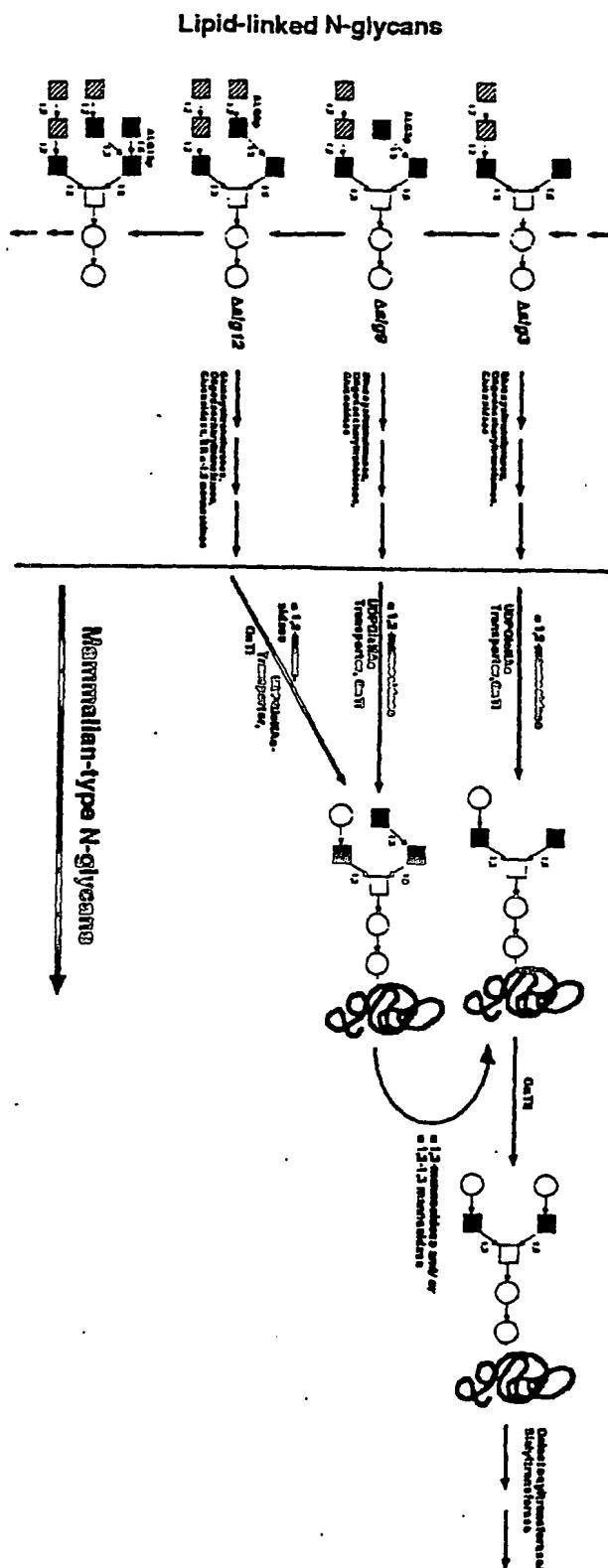


FIG. 14

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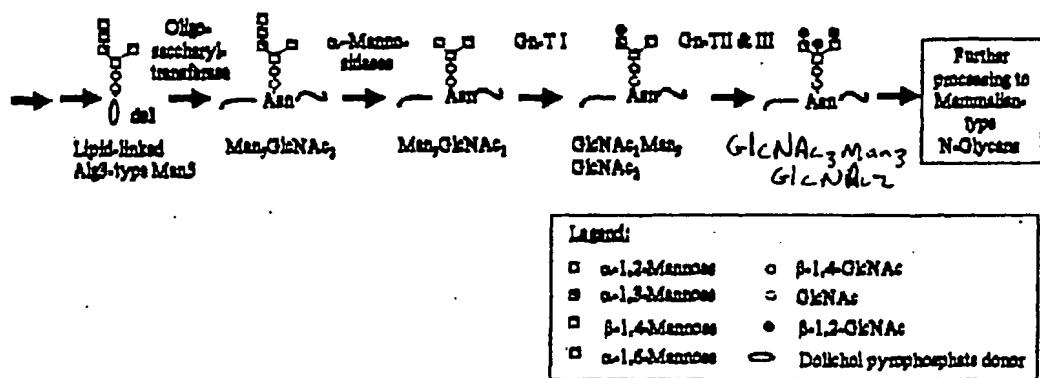


FIG. 15

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ALG3 Blast 05-22-01

Sequences producing significant alignments: (bits) Value

gi 586444 sp P38179 ALG3_YEAST	DOLICHYL-P-MAN:MAN(5)GLCNAC(...797	0.0
gi 3024226 sp Q92685 ALG3_HUMAN	DOLICHYL-P-MAN:MAN(5)GLCNAC...173	7e-43
gi 3024221 sp Q24332 NT56_DROVI	LETHAL(2)NEIGHBOUR OF TID P...145	3e-34
gi 3024222 sp Q27333 NT56_DROME	LETHAL(2)NEIGHBOUR OF TID P...121	3e-27
gi 10720153 sp P82149 NT52_DROME	LETHAL(2)NEIGHBOUR OF TID ...121	5e-27
gi 1707982 sp P40989 GLS2_YEAST	1,3-BETA-GLUCAN SYNTHASE CO... 32	2.8
gi 1346146 sp P38631 GLS1_YEAST	1,3-BETA-GLUCAN SYNTHASE CO... 31	6.6

Alignments

Yeast

>gi|586444|sp|P38179|ALG3_YEAST DOLICHYL-P-MAN:MAN(5)GLCNAC(2)-PP-DOLICHYL MANNOSYLTRANSFERASE
(DOL-P-MAN DEPENDENT ALPHA(1-3)-MANNOSYLTRANSFERASE)
(HM-1 KILLER TOXIN RESISTANCE PROTEIN)

Length = 458

Score = 797 bits (2059), Expect = 0.0
Identities = 422/458 (92%), Positives = 422/458 (92%)

Query: 1 MEGEQSPQGEKSLQRKOFVRPPLDLWQDLKDGVRYVIFDCRANLIVMPLLILFESMLCKI 60
MEGEQSPQGEKSLQRKOFVRPPLDLWQDLKDGVRYVIFDCRANLIVMPLLILFESMLCKI
Sbjct: 1 MEGEQSPQGEKSLQRKOFVRPPLDLWQDLKDGVRYVIFDCRANLIVMPLLILFESMLCKI 60

Query: 61 IIKKVAYTEIDYKAYMEQIEMIQLDGMLDYSQVSGGTGPLVYPAGHVLIYKMMYWLTEGM 120
IIKKVAYTEIDYKAYMEQIEMIQLDGMLDYSQVSGGTGPLVYPAGHVLIYKMMYWLTEGM
Sbjct: 61 IIKKVAYTEIDYKAYMEQIEMIQLDGMLDYSQVSGGTGPLVYPAGHVLIYKMMYWLTEGM 120

Query: 121 DHVERGVFFRYLYLLTLALQMACYYLLHPPWCVVLACLSKRLHSIYVRLFNDCFTTL 180
DHVERGOVFFRYLYLLTLALQMACYYLLHPPWCVVLACLSKRLHSIYVRLFNDCFTTL
Sbjct: 121 DHVERGVFFRYLYLLTLALQMACYYLLHPPWCVVLACLSKRLHSIYVRLFNDCFTTL 180

Query: 181 FMVVTVLGAIVASRCHQRPKLKSLALVISATYSMAVSIKMNALLYFPAMMISLFILNDA 240
FMVVTVLGAIVASRCHQRPKLKSLALVISATYSMAVSIKMNALLYFPAMMISLFILNDA
Sbjct: 181 FMVVTVLGAIVASRCHQRPKLKSLALVISATYSMAVSIKMNALLYFPAMMISLFILNDA 240

Query: 241 NVILTLLDLVAMIAWQVAVAPFLRSFPQQYLHCAFNFGRKFMYQWSINWQMMDEEAEND 300
NVILTLLDLVAMIAWQVAVAPFLRSFPQQYLHCAFNFGRKFMYQWSINWQMMDEEAEND
Sbjct: 241 NVILTLLDLVAMIAWQVAVAPFLRSFPQQYLHCAFNFGRKFMYQWSINWQMMDEEAEND 300

Query: 301 KRFXXXXXXFVTRYPRILPDWLSSLCHPLRKNAVLNANPAKTIFFVLIASN 360
KRF FVTRYPRILPDWLSSLCHPLRKNAVLNANPAKTIFFVLIASN
Sbjct: 301 KRFHLALLISHLIALTTLFVTRYPRILPDWLSSLCHPLRKNAVLNANPAKTIFFVLIASN 360

Query: 361 FIGVLFSLRSLHYQFLSWYHWTLPILIFWSGMPFFVGPIIWYVLHEWCWNSYPPNSQXXXX 420
FIGVLFSLRSLHYQFLSWYHWTLPILIFWSGMPFFVGPIIWYVLHEWCWNSYPPNSQ
Sbjct: 361 FIGVLFSLRSLHYQFLSWYHWTLPILIFWSGMPFFVGPIIWYVLHEWCWNSYPPNSQASTLL 420

Query: 421 XXXXXXXXXXXXXXXXSGSVALAKSHLRTTSSMEKKLN 458
SGSVALAKSHLRTTSSMEKKLN
Sbjct: 421 LALNTVLLLLALTQLSGSVALAKSHLRTTSSMEKKLN 458

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Human

>gi|3024226|sp|Q92685|ALG3_HUMAN DOLICHYL-P-MAN:MAN(5)GLCNAC(2)-PP-DOLICHYL
 MANNOSYLTRANSFERASE
 (DOL-P-MAN DEPENDENT ALPHA(1-3)-MANNOSYLTRANSFERASE)
 (NOT56-LIKE PROTEIN)
 Length = 438

Score = 173 bits (439), Expect = 7e-43
 Identities = 133/396 (33%), Positives = 195/396 (48%), Gaps = 28/396 (7%)

Query: 26 WQDLKDGVRVIFDCRANLIVMPLLLILFESMLCKIIKKVAYTEIDYKAYMEQIEMIQLD 85
 WQ+ R ++ + R L+V L L E + +I +VAYTEID+KAYM ++E + ++
 Sbjct: 29 WQER----RLLIREPRYTLVVAACLCLAEVGITFWVIHRVAYTEIDWKAYMAEVEGV-IN 83

Query: 86 GMLDYSQVSQGGTGPLVYPAGHVLIYKMMYWLTGMDHVERGQVFFRYLYLLTLALOMACY 145
 G DY+Q+ G TGPLVYPAG V I+ +Y+ T + Q F LYL TL L Y
 Sbjct: 84 GTYDYTQLQGDTGPLVYPAGFVYIFMGLYYATSRGTDIRMAQNIPIAVLYLATLLLVFLIY 143

Query: 146 Y-LHLPPWC-VVIACLSKRLHSIYVRLFLNDCFTTLFMVVTVLGAIVASRCHORPKLKK 203
 + +PP+ + C S R+HSI+VRLFLND + + +L + OR
 Sbjct: 144 HQTCKVPPVFVFFFMCCASYRVHSIFVRLFLNDP-----VAMVLLFLSINLLLAQRNGWG- 197

Query: 204 SLALVISATYSMAVSIKMNALLYFPAMMISLFILNDANVILTLDDLVAMIAWQAVAVPF 263
 +S+AVS+KMN LL+ P ++ L L L + A + QV + +PF
 Sbjct: 198 -----CCFFPSLAVSVKMNVVLFAPGPLLFLLTQFGFRGALPKLGICAGL--QVVLGLPF 249

Query: 264 LRSFPQQYLHCAFNFGRKFCMYQWSINWQMMDEEEAFNDIKRXXXXXXXXXXXXXXFVTRY 323
 L P YL +F+ GR+F++ W++NW+ + E F + F + R+
 Sbjct: 250 LLENPSGYLSRSFDLGRQFLFHWTVNWRFLPEALFLHRAFHALLTAHLTLLLFAFCRW 309

Query: 324 PRILPDLWSSLCHPLRKNAVILNANPAKTIPFVLIASNFIGVLFSSRLHYQFLSWYHWTL 383
 R + S L P ++ I L SNEFIG+ FSRSRLHYQF WY TLP
 Sbjct: 310 HRTGESILSLLRDPSKRKVPPQPLTPNQIVSTLFTSNFIGICFSRSRLHYQFYVWYFHTLP 369

Query: 384 ILIF-----WSGMPFFVGPINWYVLHEWCWNNSYPPNS 414
 L++ W + + + E WN+YP S
 Sbjct: 370 YLLWAMPARWLTHLLRLVLGLI--ELSNNTYPSTS 403

Drosophila Vi

>gi|3024221|sp|Q24332|NT56_DROVI LETHAL(2) NEIGHBOUR OF TID PROTEIN (NOT58)
 Length = 526

Score = 145 bits (366), Expect = 3e-34
 Identities = 103/273 (37%), Positives = 157/273 (56%), Gaps = 17/273 (6%)

Query: 33 VRVYIFDCRANLIVMPLLLILFESMLCKIIKKVAYTEIDYKAYMEQIEMIQLDGMLDYSQ 92
 ++Y+ F+ A IV L++L E+++ ++I++V YTEID+KAYM++ E L+G +YS
 Sbjct: 34 IKYLAPEPAALPIVSVLIVLAEAVINVVLVIQRVPYTEIDWKAYMQECCEGF-LNGTTNYS 92

Query: 93 VSGGTGPLVYPAGHVLIYKMMYWLTGMDHVERGQVFFRYLYLLTLALOMACYLLH-LP 151
 + G TGPLVYPAG V IY +Y+LT +V Q F +YLL + L + Y +P
 Sbjct: 93 LRGDTGPLVYPAAFVYIYSGLYYLTGQGTNVRLAQYIFACIYLLQMCVLRLYTKSRKVP 152

Query: 152 PWCVLACL-SKRLHSIYVRLFLNDCFTTLFMVVTVLGAIVASRCHORPKLKSALVIS 210
 P+ +VL+ S R+HSIYVRLFLND L +L A + QR L S
 Sbjct: 153 PYVLVLSAFTSYRIHSIYVRLFLNDPVAIL-----LLYAAALNLFLDQRWTLG-----S 200

Query: 211 ATYSMAVSIKMNALLYFPAMMISLFILNDANVILTLDDLVAMIAWQAVAVPFLRSFPQQ 270
 YS+AV +KMN + A + LF L + V+ TL+ L Q+ + PFLR+ P +
 Sbjct: 201 ICYSLAVGVKMN-ILLFAPALLLFYLANGLVRLTVQLTICAVLQLFIGAPFLRTHPM 258

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Query: 271 YLHCAFNFGRKFMYQWSINWQMMDEEAFNDKRF 303
 YL +F+ GR F ++W++N++ + +E F + F
 Sbjct: 259 YLRGSFDLGRIFEHKWTVNYRFLSKELFEQREF 291

Score = 53.3 bits (127), Expect = 1e-06
 Identities = 31/62 (50%), Positives = 41/62 (66%), Gaps = 6/62 (9%)

Query: 352 IPFVLIASNFIGVLFSRSLHYQFLSWYHWTLPILIFWSGMPFFVGPIWYVLH--EWCWNS 409
 +PF L NFIGV +RSLHYQF WY +LP L+ WS P+ +G + +L E+CWN+
 Sbjct: 412 LPFFL--CNFIGVACARSILHYQFYIWYFHSLPYLV-WS-TPYSLGVRYLILGIIEYCWN 467

Query: 410 YP 411
 YP
 Sbjct: 468 YP 469

Drosophila melanogaster

>gi|3024222|sp|Q27333|NT56_DROME LETHAL(2)NEIGHBOUR OF TID PROTEIN (NOT56)
 (NOT45)
 Length = 510

Score = 121 bits (305), Expect = 3e-27
 Identities = 96/272 (35%), Positives = 154/272 (56%), Gaps = 17/272 (6%)

Query: 34 RYVIFDCRANLIVMPLLILFESMLCKIIKKVAYTEIDYKAYMEQIEMIQLDGMDYSQV 93
 +Y++ + A IV ++L E ++ ++I++V YTEID+ AYM++ E L+G +YS +
 Sbjct: 36 KYLLEPAALPIVGLFVLLAELVINVVVIQRVFVTEIDWVAYMQECEGF-LNGTTNYSLL 94

Query: 94 SGGTGPLVYPAGHVLIYKMMYWLTEGMHDVERGOVFRYLYLLTLALQMACYLLH-LPP 152
 G TGPLVYPA V IY +Y++T +V Q F +YLL LAL + Y +PP
 Sbjct: 95 RGDGTGPLVYPAAFVYIYSALLYVTSHGTNVRLAQQYIFAGIYLLQLALVRLYSKSRKVPP 154

Query: 153 WCVLJACL-SKRLHSIYVRLFNDCCPTILFMVVTVLGAIVASRCNQRPHLKKSIALVISA 211
 + +VL+ S R+HSIYVRLFND + V +L A + +R L S
 Sbjct: 155 YVLVLSAFTSYRIHSIYVRLFNDP-----VAVLLLAAALNLFLDRRWTLG-----ST 202

Query: 212 TYSMAVSIKMNALLYFPAMMISLFLINNDANVILTLDDLVAMIAWQAVAVPFLRSFPQQY 271
 +S+AV +KMN + A + LF L + ++ T+L L O+ + PFL + P +Y
 Sbjct: 203 FFSLAVGVKMN---ILLFAPALLLFYLANLGLLRTILQLAVCGVIQLLLGAFFLLTHPVEY 260

Query: 272 LHCAFNFGRKFMYQWSINWQMMDEEAFNDKRF 303
 L +F+ GR F ++W++N++ + +F ++ F
 Sbjct: 261 LRGSFDLGRIFEHKWTVNYRFLSDVFNRTF 292

Score = 49.4 bits (117), Expect = 2e-05
 Identities = 27/60 (45%), Positives = 35/60 (58%), Gaps = 2/60 (3%)

Query: 352 IPFVLIASNFIGVLFSRSLHYQFLSWYHWTLPILIFWSGMPFFVGPIWYVLH-EWCWNS 411
 +PF L N +GV SRSIHYOF WY +LP L + + V + L E+CWN+YP
 Sbjct: 407 LPFFL--CNLVGVACRSRSLHYQFYVWYFHSLPYLAWSTPYSLGVRCILGLIEYCWN 464

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Matrix: BLOSUM62
Gap Penalties: Existence: 11, Extension: 1
Number of Hits to DB: 28883317
Number of Sequences: 95469
Number of extensions: 1107545
Number of successful extensions: 2870
Number of sequences better than 10.0: 16
Number of HSP's better than 10.0 without gapping: 5
Number of HSP's successfully gapped in prelim test: 11
Number of HSP's that attempted gapping in prelim test: 2839
Number of HSP's gapped (non-prelim): 23

length of query: 458
length of database: 35,174,128
effective HSP length: 45
effective length of query: 413
effective length of database: 30,833,023
effective search space: 12734038499
effective search space used: 12734038499
T: 11
A: 40
X1: 15 (7.1 bits)
X2: 38 (14.6 bits)
X3: 64 (24.7 bits)
S1: 40 (21.8 bits)
S2: 67 (30.4 bits)

FIG.16-4

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S. cerevisiae ALG3

ATGGAAGGTGAACAGTCTCCGCAAGGTGAAAAGTCTCTGCAAAGGAAGC
 AAITTGTCAGACCTCCGCTGGATCTGTGGCAGGATCTAAGGACGGTGTG
 CGCTACGTGATCTCGATTGTAGGGCCAATCTTATCGTTATGCCCTTTG
 ATTTGTTGAAAGCATGCTGTGCAAGATTATCATTAAAGAAGGTAGCTTAC
 ACAGAGATCGATTACAAGGCATACATGGAGCAGATCGAGATGATTAGCT
 CGATGGCATGCTGGACTACTCTCAGGTGAGTGGTGGAACGGGCCGCTGG
 TGTATCCAGCAGGCCACGTCTTGTATCTACAAGATGATGTAAGCTAACA
 GAGGGAAATGGACCACGTTGAGCGCGGGCAAGTGTTCAGATACTTGT
 TCTCCTTACACTGGCGTACAAATGGCGTGTACTACCTTTACATCTACC
 ACCGTGGTGTGTGGTCTTGGCGTGCCTCTCTAAAAGATTGCACTCTATT
 CGTGCCTACGGTATTCAATGATTGCTTCACTACTTTGTATGGTCGTACG
 GTTTGGGGCTATCGTGGCCAGCAGGTGCCATCAGCGCCCCAAATTAAA
 GAAGTCCCTTGCCTGGTGTCTCCGCAACATACAGTATGGCTGTGAGCA
 TTAAGATGAATGCGCTGTTGTATTCCCTGCAATGATGATTCTCTATT
 CCTTAATGACGCAACGTAATCCTTACTTTGTGGATCTCGTTGCGATGAT
 TGCATGGCAAGTCGAGTTGCAGTGCCTTCCCTGCGCAGCTTCCGCAACA
 GTACCTGCATTGCGTTTAAATTCCGGCAGGAAGTTATGTAACCAATGGAG
 TATCAATTGGCAAATGATGGATGAAGAGGGTTCAATGATAAGAGGTCC
 ACTTGGCCCTTTAAATCAGCCACCTGATAGCGCTCACCAACTGTTGTCA
 CAAGATACCCTCGCATCCTGCCGATTATGGCTTCCCTGCGCATCCGC
 TGAGGAAAAATGCAGTGCTCAATGCCAATCCGCCAAGACTATTCCATT
 GTTCTAATCGCATCCAACCTCATCGGCGTCTATTTCAGGTCCCTCCAC
 TACCAAGTTCTATCCTGGTATCACTGGACTTGCCTATACTGATTTGGT
 CGGGAAATGCCCTTCTCGITGGCCATTGGTACGTCTGACGAGGTGGT
 GCTGGAATTCTATCCACCAAAACTCACAAAGCAAGCACGCTATTGGCA
 TTGAATACTGTTCTGTTGCTTCTATTGGCCTGACGCCAGCTATGGTTCGG
 TCGCCCTGCCAAAAGCCATCTCGTACCAACCAGCTATGGAAAAAAAG
 CTCAACTGA

S. cerevisiae Alg3p

MEGEQSPQGEKSLQRKQFVRPLDLWQDLKDGVRYVIFDCRANLIVMPLLIL
 FESMLCKIIKKVAYTEIDYKAYMEQIEMIQLDGMLDYSQVSGGTGPLVYPAG
 HVLIYKMMYWLTGMDHVERGVFFRYLYLLTLALQMACYLLHLPPWCV
 VLAICSLKRLHSIYVRLFNDCFTTLMVVTVLGAIVASRCHQRPKLKSLALV
 ISATYSMAVSIKMNALLYFPAMMISLFILNDANVILTLLDVAMIAWQVAVA
 VPFLRSFPQQYLHCAFNFGRKFMYQWSINWQMMDEEAFNDKRFHLALLSHL
 IALTLFVTRYPRILPDLWSSLCHPLRKNAVLANPAKTIPVLIASNFIGVLFS
 RSLHYQFLSWYHWTLPIFWSGMPFFVGPIWYVLHEWCWNSYPPNSQASTL
 LLALNTVLLLLALTQLSGSVALAKSHLRTTSSMEKKLN

FIG. 17

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P. pastoris ALG3

ATGCCTCCGATAGAGCCAGCTGAAAGGCCAAGCTTACGCTGAAAAATGT
 TATCGGTGATCTAGTGGCTCTTATTCAAAACGTTTATTTAACCCAGATTT
 AGTGTCTTCGTTGCACCTCTTTATGGTTAGCTGATTCCATTGTATCAAGG
 TGATCATTGGCACTGTTCTACACAGATATTGATTTCTCATATATGCA
 ACAAAATCTTAAAATTGACAAGGAGAATTAGATTATAGAACATATTG
 GTGACACCGGTCCATTGGTTACCCAGCCGGCCATGTTATGCTTACTCAG
 TACTTCTGGTACAGTGATGGTGGAGAAGACGTCACTTCAGCTCAACAA
 GCATTGGTGGTTACCTAGGTGCTTACTATCCATCAGCTCTACT
 TTTCTCTGGCTTAGGGAAAATACCTCCGGTTATTTGTTTGTGGTAGC
 GTCCAAGAGACTGCATTCAATAATTGTATTGAGACTCTCAATGACTGTT
 AACAAACATTGATGTTGGCAACTATAATCATCCITCAACAAGCAAGTAG
 CTGGAGGAAAGATGGCACAACTATTCCATTATCTGCCCTGATGCTGCAG
 ATACGTACAGTTAGCCATCTGTAAAGATGAATGCGCTGCTACCTCC
 CAGCATTCTACTACTCATATATCTATTGTGACGAAAATTGATTAAAG
 CCTTGGCACCTGTTAGTITGATATTGGTGCAAGTAGGAGTCGGTTATT
 CGTTCAATTACCGTTGCACATGATGATCAGGCAAATGAAATTGTTCTG
 CCTACTTACAGCAGGCTTTGACTTAGTCGCCAATTCTTATAAGTGGAA
 CGGTTAATTGGCGCTTTGAGCCAAGAAACTTCAACAATGCCATTTC
 ACCAGCTCCTGTTGCTCTCCATATTACGTTAGTCTGTTATCCTCAA
 GTTCCCTCTCCTAAAAACATTGAAAACCGCTGGTAGATTGTTGGAA
 CATTTCAAATTGGAAGCCAACCTTATCTCAACCAATTATCAACGA
 CCCAGAAAGAGCCCAGATTGTTACACCGTCATGGCTACTACCAACTT
 AATAGGGGTGCTTTGCAAGATCTTACACTACAGTTCTAAGCTGGTA
 TGCCTCTCTTGCATATCTCTTACAAGGCTGCTGAACCTTATAGCA
 TCTATTATTGTTATGCCGCTCACGAGTATTGCTGGTGGTTCTCAGCTA
 CAGAACAAAGTCCCGTGTGGTATCTATCTACTACTTATCCTGATTC
 TCATTTCACCAACGAACAGTTATTCTCAATCGGCCCTGAGAAA
 AAAAGAACATACATAA

P. pastoris Alg3p

MPIEPAERPKLTKNVIGDLVALIQNVLNPDSVFVAPLLWLADSIVIKVIIG
 TVSYTDIDFSSYMQQIFKIRQGELDYSNIFGDTGPLVYPAGHVAYSVLSWYS
 DGGEDVSFVQQAFGWLYLGCLLSSIYFFSGLGKIPPVYFVLLASKRLHSIF
 VRLFNDCLTFMLATIILQQASSWRKDGTIPLSVPDAADTYSLAISVKMN
 ALLYLPALLLIYLCDENLIKALAPVLVLILVQVGVGYSFILPLHYDDQANEIR
 SAYFRQAFDFSRQFLYKWTVNWRFLSQETFNNVHFQLLFALHIITLVLFILKF
 LSPKNIGKPLGRFVLDIFKFWKPTLSPTNIINDPERSPDFVYTVMATTNLIGVLF
 ARSLHYQFLSWYAFSLPYLLYKARLNFIASIIVYAAHEYCWLVPATEQSSAL
 LVSILLILIFTNEQLFPSQSVPAEKKNT

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P. pastoris ALG3 BLAST

Sequences producing significant alignments:	(bits)	Value
gi 586444 sp P38179 ALG3 YEAST Dolichyl-P-Man:Man(5)GlcNAc{...228	2e-58	
gi 12802365 gb AAK07848.1 AF309689 10 putative NOT-56 manno...212	8e-54	
gi 984725 gb AAA75352.1 ORF 1	206	4e-52
gi 7492702 pir T39084 probable mannosyltransferase - fission...176	8e-43	
gi 16226531 gb AAL16193.1 AF428424 1 At2g47760/F17A22.15 [A...164	2e-39	
gi 25367230 pir B84919 Not56-like protein [imported] - Ara...164	3e-39	
gi 25814791 emb CAB70171.2 Hypothetical protein K09E4.2 [C...161	2e-38	
gi 17535001 ref NP_496950.1 Putative plasma membrane membr...160	3e-38	
gi 1654000 emb CAA70220.1 Not56-like protein [Homo sapiens...155	2e-36	
gi 13279206 gb AAH04313.1 AAH04313 Unknown (protein for IMA...154	2e-36	
gi 22122365 ref NP_666051.1 hypothetical protein MGC36684 ...150	3e-35	
gi 21292031 gb EAA04176.1 agCP3388 [Anopheles gambiae str...120	4e-26	
gi 1780792 emb CAA71167.1 lethal(2)neighbour of tid [Droso...114	3e-24	

Alignments

S. cerevisiae

Score = 228 bits (580), Expect = 2e-58
 Identities = 154/429 (35%), Positives = 229/429 (53%), Gaps = 37/429 (8%)

Query: 9 RPKLTALKVIGDLVALIQNVLFPNPDFSVFVAPLLWLADSVIKVIIGTVSYTDIDFSSYM 68
 RP L L DL ++ V+F+ ++ V PLL L +S++ K+II V+YT+ID+ +YM
 Sbjct: 20 RPPLDLWQ---DLKDGVRYVIFDCRANLIVMPLLILFESMLCKIIKKVAYTEIDYKAYM 76

Query: 69 QQIFKIR-QGELDYSNI F GDTGPLLVYPAGHVHAYSVLSWYS DGGEDVS FVQQAFGHLYLG 127
 +QI I+ G LDYS + G TGPLVYPAGHV Y ++ W ++G + V Q F +LYL
 Sbjct: 77 EQIEMIQLDGMLDYSQVSGGTGPLVYPAGHVLIYGMWLT EGMDHVERGQVFFRYLYLL 136

Query: 128 CLLLSSIYYFFSGLKGIPPVYFVLLVASKRLHSIFVLRFLNDCLTTFLMLATI---IILQ 184
 L L ++ Y+ L +PP VL SKRLHSI+VRLRFND C TT M+ T+ I+
 Sbjct: 137 TLALOMACYY---LLHLPWCVVLACLSKRLHSIVVRLRFNDCTTLMVVTVLGAIVAS 193

Query: 185 QASSWRKDGTITPLSVPDAADTYSLAISVKMNXXXXXXXXXXXXCDENLIKALAPXX 244
 + K ++ L + + TYS+A+S+KMN D N+I L
 Sbjct: 194 RCHQRPKLKKSLALVI---SATYSMAVSIKMNALLYFPAMMISLFILNDANVILTLDDLV 250

Query: 245 XXXXXXXXXXXXYSFILPLHYDDQANEIRSAYFRQAFDFSRQFLYKWTVNWRFLSQETFNNV 304
 F+ Y AF+F R+F+Y+W++NW+ + +E FN+
 Sbjct: 251 AMIAWQVAVAVPFL-----RSFPQQYLHCAFNFGRKEMYQWSINWQMMDEEAFTNDK 301

Query: 305 HFHQOLLFALHIITL-VLFILKFSLPKNIGKPLGRFVLDIFKFWKPTLSPTNIIN-DPERS 362
 FH L H+I L LF+ ++ R + D++ L ++N +P ++
 Sbjct: 302 RFHLALLLISHLIALTTLFVTRY-----PRILPDLWSSLCHPLRKNAVLANPAKT 351

Query: 363 PDFVYTVMATTNLIGVLFARSLHYQFLSWYAFSLPYLLYKARLNFIASIIVYAAHEYCWL 422
 F V+ +N IGVLF+RSLHYQFLSWY ++LP L++ + + F I Y HE+CW
 Sbjct: 352 IPF---VLIASNFIGVLFSRSLHYQFLSWYHWTL PILIFWWSGMPFFVGPIWYVLHEWCWN 408

Query: 423 VFPATEOSS 431
 +P Q+S
 Sbjct: 409 SYPPNSQAS 417

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Neurospora crassa

Score = 212 bits (540), Expect = 8e-54
 Identities = 140/400 (35%), Positives = 212/400 (53%), Gaps = 29/400 (7%)

Query: 35 SVFVAPLLWLADSTIVKVIIGTVSYTDIDFSSYMMQOIFKIROGELDYSNIFGDTGPLVYP 94
 S + P L+L D+++ II V YT+ID+++YM+Q+ +I GE DY+ + G TGPLVYP
 Sbjct: 33 SKLIPPALFLVDALLCGLIWIWKVPYTEIDWAAYMEQVSQILSGERDYTKVRGGTGPLVYP 92

Query: 95 AGHVVHAYSVLSWYSDGGEDVSFVQQAFGWLYLGCLLLSISYYFFSGLGKIPPVYFVLLVA 154
 A HV+ Y+ L +D G ++ QQ F LY+ L + + Y+ K PP F LL
 Sbjct: 93 AAHVVIYTGLYHLITDEGRNILLACQLPAGLYMVTIAVMGCYW---QAKAPPYLFPPLLTL 149

Query: 155 SKRLHSIFVRLRFNDCLTTFLMLATIIILQQASSWRKDGTIPLSVPDAADTYSLAISVK 214
 SKRLHSIFVLR FNDC + I Q+ +W+ A Y+L + VK
 Sbjct: 150 SKRLHSIFVRLCFNDCAFVLFLWLAIFFQR-RNWQA-----GALLYTLGLGVK 197

Query: 215 MNDXXXXXXCDENLIKALAPXXXXXXXXXXXXYFSILPLHYDDQANEIRSAY 274
 M + + L F+ HY + Y
 Sbjct: 198 MTLLLSSLPAVGIVLFLGSG-SFVTTLQLVATMGLVQILIGVPFL--AHYPTE-----Y 247

Query: 275 FRQAFDFSROFLYKWTVNWRFLSQETFNNVHFQOLLFAHLHIITLVLFI-LKFSPKNIGK 333
 +AF+ SRQF +KWTVNWRF+ +E F + F L ALH++ L +FI ++++ P K
 Sbjct: 248 LSRAFELSROFFFKWTVNWRFGEEIFLSKGFBATLLALRVLVLGIFITTRWIKPAR--K 305

Query: 336 PLGRFVLDIFPKFWKPTLS-PTNIINDPERSPDFVYTVMAATTNLIGVLFARSLSHYQFLSWY 392
 L + + + KP L+ P + + +P ++ T + + N +G+LFARSLSHYQF ++
 Sbjct: 306 SLVQLISFVLLAGKPPPLTVPEHRAARDVTPRYIMTTILSANAVGLLFARSLSHYQFYAYV 365

Query: 393 AFSLPYLLYKARLNFIASIIVYAAHEYCWLVPATEQSSA 432
 A+S P+LL++A L+ + +--+A HE+ W VFP+T SSA
 Sbjct: 366 AWSTPFLLWRAGLHPVLVYLLWAVHEAWNVPSTPASSA 405

Schizosaccharomyces pombe

Score = 176 bits (445), Expect = 8e-43
 Identities = 132/390 (33%), Positives = 194/390 (49%), Gaps = 35/390 (8%)

Query: 42 LWLADSTIVKVIIGTVSYTDIDFSSYMMQOIFKIROGELDYSNIFGDTGPLVYPAGHVHAY 101
 L L + + II V YT+ID+ +YM+Q+ GE DY ++ G TGPLVYP GHV Y
 Sbjct: 30 LLLLIEIPFVFAIISKVPYTEIDWIAYMEQVNSFLLGERDYKSLVGCTGPLVYPGGHVFLY 89

Query: 102 SVLSWYSDGGEDVSFVQQAFGWLYLGCLLLSISYYFFSGLGKIPPVYFVLLVASKRLHSI 161
 ++L + +DGG ++ Q F ++Y + +I Y F + + P +VLL+ SKRLHSI
 Sbjct: 90 TLLYYLTGGTNIVRAQYIFAFVYW--ITTAIVGYLFK-IVRAPFYIYVLLILSKRLHSI 146

Query: 162 FVLRLFNDCLTTFLMLATIIILQQASSWRKDGTIPLSVPDAADTYSLAISVKMNXXXXX 221
 F+LRLFND + L + I+ W + A+ S+A SVKM+
 Sbjct: 147 FILRLFNDGFNS-LFSSLFILSSCKKKWNR-----ASILLSVACSVKMSSLVY 194

Query: 222 XXXXXXXXXCDENLIKALAPXXXXXXXXXXXXYFSILPLHYDDQANEIRSAYFRQAFDF 281
 L++ L P + + + +Y+ QAFDF
 Sbjct: 195 PAYLVL-----LHQILGPKKTWMHIFVIIIVQILFSIPF---LAYFWSYWTQAFDF 242

Query: 282 SRQFLYKWTVNWRFLSQETFNNVHFQOLLFAHLHIITLVLFLKFLSPKNIGKPLGRFVLD 341
 R F YKWTVNWRF+ + F + F + LH+ LV F K + + P
 Sbjct: 243 GRAFDYKWTVNWRFIPRSIFESTSFSTSILFLHVALLVAFTCKHWNKLSRATP----- 295

Query: 342 IFKFWKPTLSPTNIINDPERSPDFVYTVMAATTNLIGVLFARSLSHYQFLSWYAFSLPYLLY 401
 F L+ + +P+F++T +AT+NLIG+L ARSLHYQF +W+A+ PYL Y

Sbjct: 296 -FAMVNNSMLTLKPLPKLQLATPNFIFTALATSNLIGILCARSLEYQFYAWFAWSPYLCY 354

Query: 402 KARLNFIASIIVYAAHEYCWLVFPATEQSS 431
+A I ++ EY W VFP+T+ SS

Sbjct: 355 QASFPAPIVIGLWMLQEYAWNVPSTKLSS 384
Arabidopsis thaliana

Score = 164 bits (415), Expect = 2e-39
Identities = 131/391 (33%), Positives = 194/391 (49%), Gaps = 29/391 (7%)

Query: 42 LWLADSVIKVIITGTVSYTDIDFSSYMMQQIFKIRQGELDSNIFGDTGPLVYPAGHVHAY 101
L LAD+I++ +II V YT ID+ +YM Q+ GE DY N+ GDTGPLVYPAG ++ Y

Sbjct: 39 LILADAILVALIIAYVPTKIDWDAYMSQVSGLGGERDYGNLKGDGPLVYPAGFLVYV 98

Query: 102 SVLSWYSDGGEDVSFVQQAFGWLYLGCLLLSISYYFFSGLGKIPPVYFVLLVASKRLHSI 161
S + + G +V Q FG LY+ L + + Y + + +P LL SKR+HSI

Sbjct: 99 SAVQNLTGG--EVYPAQILFGVLYIVNLGIVLIYVKTDV--VPWWALSLLCLSKRIHSI 154

Query: 162 FVLRLFNDCLTTFLMLATIIIILQQASSWRKDGTTIPLSVPDAADTYSLAISVKMNXXXXX 221
FVLRLFMDC L+ A++ + +RK + + + +S A+SVKMN

Sbjct: 155 FVLRLFNDCFAMTLLHASMALFL---YRJWHLGMLV-----FSGAVSVIGNVLLYA 202

Query: 222 XXXXXXXXXXXXCDENLIKALAPXXXXXXXXXXYSFILPLHYDDQANEIRSAYFRQAFDF 281
N+I ++ F++ +Y AFD

Sbjct: 203 PTLLLLLLKAM--NIIGVVVSALAGAALAQILVGLPFLITYPV-----SYIANAFDL 251

Query: 282 SRQFLYKWTVNWRFLSQETFNNVHFQOLLFALIIIITLVLFILKFLSPKNIGKPLGRFVLD 341
R F++ W+VN++F+ + F + F L H+ LV F + K+ G +G

Sbjct: 252 GRVFIHFWSVNFKFVPERVFVSKEFAVCLLIAHFLLLVAFA-NYKWCHEGGIIGFMRSR 310

Query: 342 IFKFWKP-TLSPTNIINDPERSPDFVYTVMATTNLIGVLFARSLHYQFLSWYAFSLPYILL 400
F F +LS +++ + + V T M N IG++FARSLHYQF SWY +SLPYILL

Sbjct: 311 HFFLTLPSLSSFSDVSASRIITKEHVVVTAMFVGFIGIVFARSLHYQFYSWFYSLPYILL 370

Query: 401 YKARLNFIASIIVYAAHEYCWLVFPATEQSS 431
++ +I++ E CW V+P+T SS

Sbjct: 371 WRTPFPPTWLRLIMFLGIELCWNVYPSTPSSS 401

FIG. 19-3

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K. lactis ALG3

TTTGTTCACAAGCTGATACCAACGAACATGAATAACCCGGCAGGTTACT
GAAGATTGGCAAAGCTAACCTTTACATCCTTTACCGATGCTGTATTCA
TGCATGAGAGTAAACGCAGAACAAATTGCATACATTACTTGTACCA
ATTACATTGGAGTACTATTGCTCGATCATTACACTACCAATTCTATCTT
GGTACCATGGACGGTACCACTATTGAATTGGGCCAATGTTCCGTATC
CGCTATGTGTGCTATGGTACCTAACACATGAGTGGTGCTGGAACAGCTAT
CCGCCAAACGCTACTGCATCCACACTGCTACACGCGTGTAAACACATACTG
TTATTGGCTGTATTCTTAAGAGGACCCGCAAACTCGAAAAGTGGTGATAA
CGAAACAAACACACGAGAAAGCTGAG

K. lactis Alg3p

FVYKLIPTNMNTPAGLLKIGKANLLHPFTDAVFSAMRVNAEQIAYILLVTNYI
GVLFARSLHYQFLSWYHWTLPVLLNWANVPYPLCVLWYLTHEWCWSYPP
NATASTLLHACNTYCYWLYSZEDPQTRKVVITKQHTRKL

FIG. 20

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K. lactis ALG3 BLAST

Score	E	Sequences producing significant alignments:	(bits)	Value
gi 586444 sp P38179 ALG3 YEAST	Dolichyl-P-Man:Man(5)GlcNAc(...	125	1e-28	
gi 984725 gb AAA75352.1 ORF 1		94	4e-19	
gi 16226531 gb AAL16193.1 AF428424_1	At2g47760/F17A22.15 [A...	72	1e-12	
gi 25367230 pir BB4919	Not56-like protein [imported] - Ara...	72	1e-12	
gi 21292031 gb EAA04176.1	agCP3388 [Anopheles gambiae str....	69	2e-11	
gi 20892051 ref XP_148657.1	similar to Lethal(2)neighbour ...	65	2e-10	

Alignments*S. cerevisiae*

Score = 125 bits (314), Expect = 1e-28
 Identities = 60/120 (50%), Positives = 83/120 (69%), Gaps = 1/120 (0%)
 Frame = +3

Query: 66 ANLLHPFT-DAVFSAMRVNAEQIAYILLVTNYIGVLFARSLHYQFLSWYHWTLPVLLNWA 242
 ++L HP +AV +A A+ I ++L+ +N+IGVLF+RSLHYQFLSWYHWTLP+L+ W+
 Sbjct: 332 SSLCHPLRKNAVLNANP--AKTIPFVLIASNFIGVLFSRSLHYQFLSWYHWTLPILIFWS 389

Query: 243 NVPYPLCVLWYLTHEWCWNNSYPPNATASTLLHACNTYCYWLYS*EDPQTRKVVITKQHTR 422
 +P+ + +WY+ HEWCWNNSYPPN+ ASTLL A NT .L + + V + K H R
 Sbjct: 390 GMPFFVGPIWYVLHEWCWNNSYPPNSQASTLLLALNTVLLLLLA-LTQLSGSVALAKSHLR 448

A. thaliana

Score = 72.0 bits (175), Expect = 1e-12
 Identities = 42/107 (39%), Positives = 57/107 (53%), Gaps = 3/107 (2%)
 Frame = +3

Query: 84 FTDAVFSAMRVNAEQIAYILLVTNYIGVLFARSLHYQFLSWYHWTLPVLLNWNVPYPLC 263
 F+D S + + E + + V N+IG++FARSLHYQF SWY ++LP LL P L
 Sbjct: 322 FSDVSASRI-ITKEHVVTAMFVGNNFIGIVFARSLHYQFYSLPYLLWRTPPTWLR 380

Query: 264 VLWYLTHEWCWNNSYPPNATASTL---LHACNTYCYWLYS*EDPQTRK 395
 +- +L E CWN YP +-S L LH WL DP K
 Sbjct: 381 LIMFLGIELCWNVYPSTPSSGLLLCLHLIIILVGLWLAPSVDPYQLK 427

FIG. 21

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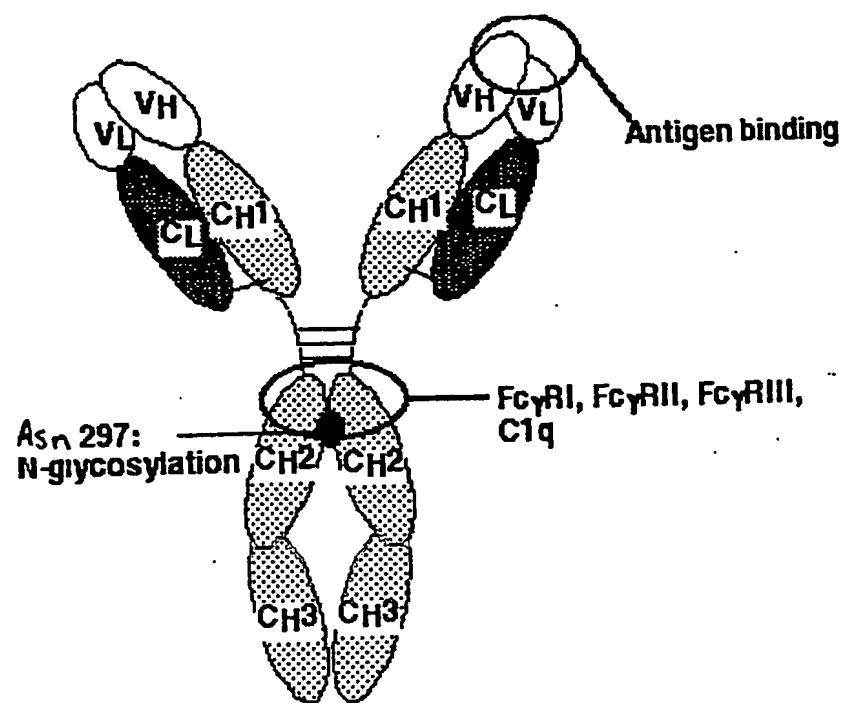


FIG. 22

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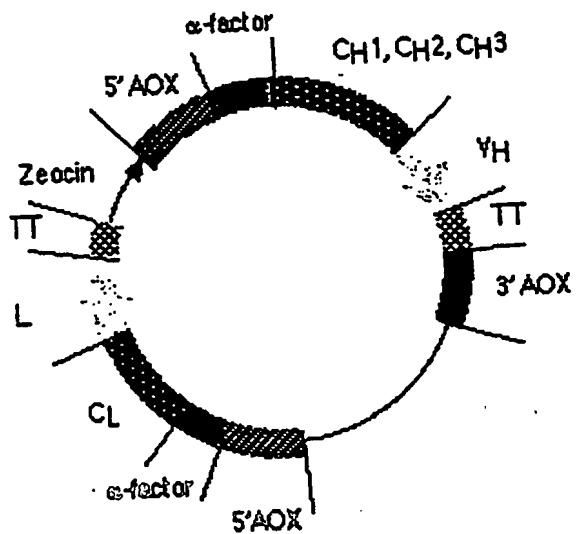


FIG. 23

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>gi|6754685|ref|NM_010795.1| Mus musculus mannoside acetyl glucosaminyltransferase 3 (Mgat3), mRNA

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ATGAAGATGAGACGCTACAAGCTTTCATGTTCTGTATGGCTGGCTGTGCCTCATATCCTTCTGC
ACTTCTTAAGACCTTATCCTATGTCACCTTCCGAGAGAACTGGCTCCCTCAGCCCTAACCTCGTATC
CAGCTTCTCTGGAACAAATGCCCTGTCACTCCCCAGGCCAGTCCGGAGCCGGGGTGCCCCGACCTATTG
CGGACACCCCTCTACTCCCACTCTCCCCGTCAGCCACTGTCCCCGAGCAAGCCACAGAGGAACCTGC
ACCGGGTGGACTTCGTGTTGCCGAGGACACCACGGAGTATTGTCGCAACCAAAGCTGGTGGTGTG
CTTCAAACAGGTACCAAGGATGCTGGAAACCTTCGCCAGGGCGAACAGAGGAAGGCCAAGTGTCT
GAGGGCTCTCAGCCCCGGGACCTGCTGGAGGCCCCTGAGGCACGTGTTGAGTACGCCGGAGCGCCTGG
GCAGCCGGGGCACTAGGCCAAGTGGGTTAGTGTGTTGCTGCCAGGCTGGCACGGGGCAAGTTCGCG
GGTGCCCCACGGTGGTGCAGTATTCAACTGCCACCAAGGAACGCCGGTACCCAGGGAGGTACCGAGG
CGGGTTATCAACGCCATCAACATCAACCAACGAGTTCGACCTGCTGGATGTCGCCCTCCATGAGCTGGGAG
ATGTTGTGGACGCCCTGCTGGTCTGTGAATCTAATTTCACCCGCTACGGGGAGCCTGGCCGCTCAAGTT
CCGAGAGATGCTGACCAATGCCACCTCGAGTACATCCGCCACAGGTGCTCTATGTCCTGGACCAT
TTCCACCTGGTGGCGTCAGGCCGGGATGGGATGGCTGAGTACCTGCGCACCTCCCTCACCAAGGATG
GCCGCTCCCGCCTGCCAACCTGCCGGGCGATGACGTCTTATCATCGACGATGCCGAGATCCCTGC
GCCGTGATGGTGTGCTGTTCTCAAACCTACGATGGCTGGACAGAGCCCTCGCCTCCACATGCCGAG
TCCCTGATGGTTCTCTGGAAAGCAGCCGGCACACTGGAGGTGGTGTGCTGAGGCTGCACCATGGACATGC
TGCAGGCCGTGATGGGCTGGGATGGCATCCGCCCTGCCGCCAGTACTACACCATGCCAACCTCCG
GCAGTATGAGAACCGCACCGGCCACATCCTAGTGCAGTGGCTCTCCGAGGCCCTGCACTTCGCCGGC
TEGCATTGCTCTGGCTTCACACCCGAGGGCATCTACTTTAAACTCGTGTGAGCCAGAATGGCAGCT
TCCCCCGCTGGGGTGAATATGAGGACAAGAGGGACCTCAATTACATCCGAGCTGATCCGACTGGGGC
ATGGTTCGACGGAACCGCAGCAGGAGTACCCCTCTGCCGGACCCAGTGAGCACATGATGCTCTAAATAC
CTGCTCAAGAACTATGACCAGTCCGCTACTTGCTGGAAATCCCTACCGGGAGCCAAAGAGCACTGTAG
AGGGTGGGGGCCAGAACCAAGGGCTCAGATGAAAGGCCATCTGCTGTCAGGGGCAAGTTGGATACAGTGG
GGGCTAG

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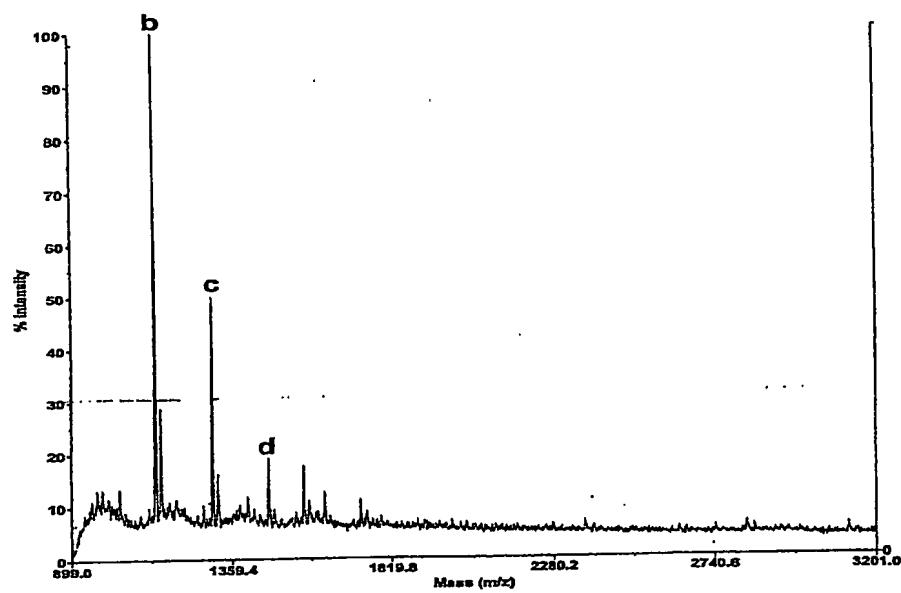
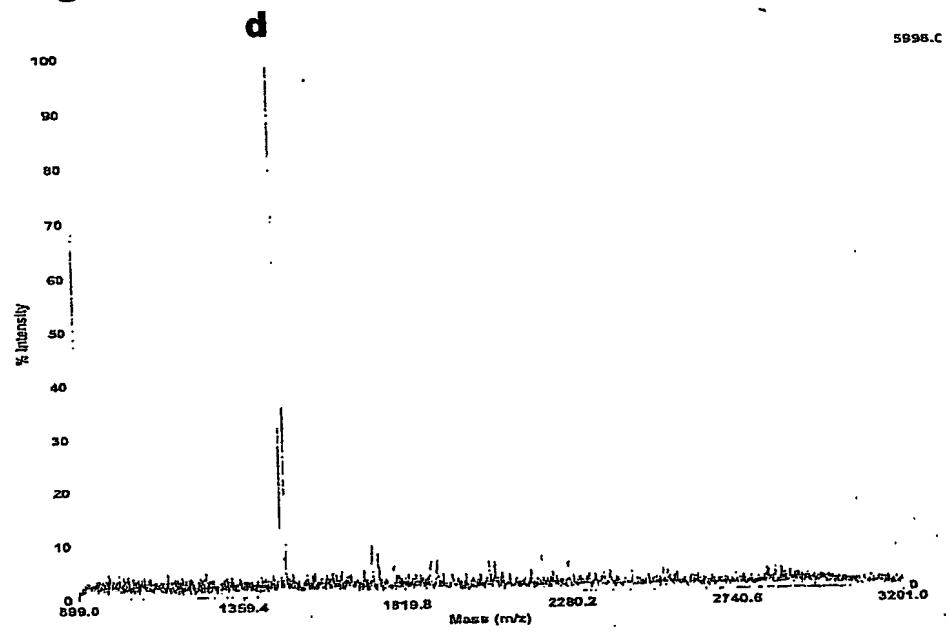
>gi|2117717|pir||JC4362 beta-1,4-mannosyl-glycoprotein 4-beta-N-acetylglucosaminyltransferase (EC 2.4.1.144) III - mouse

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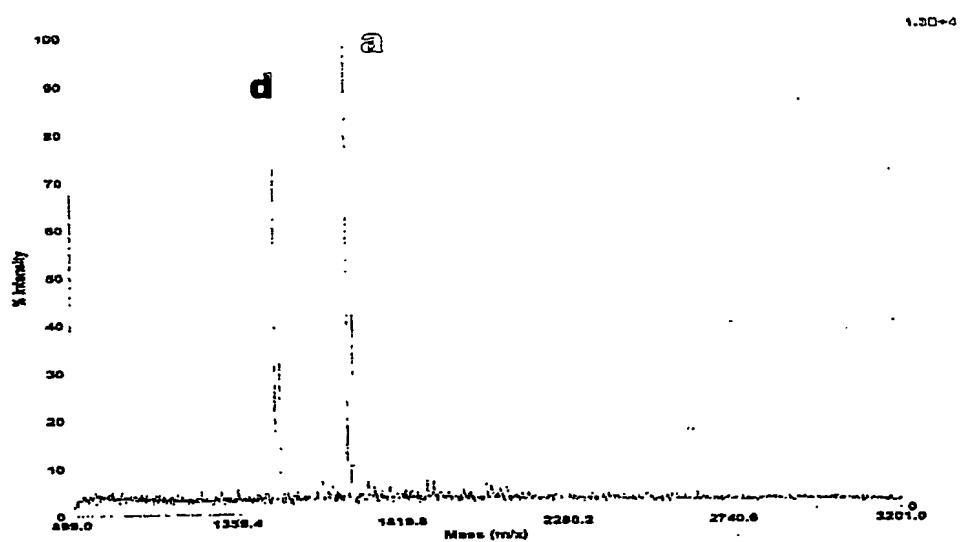
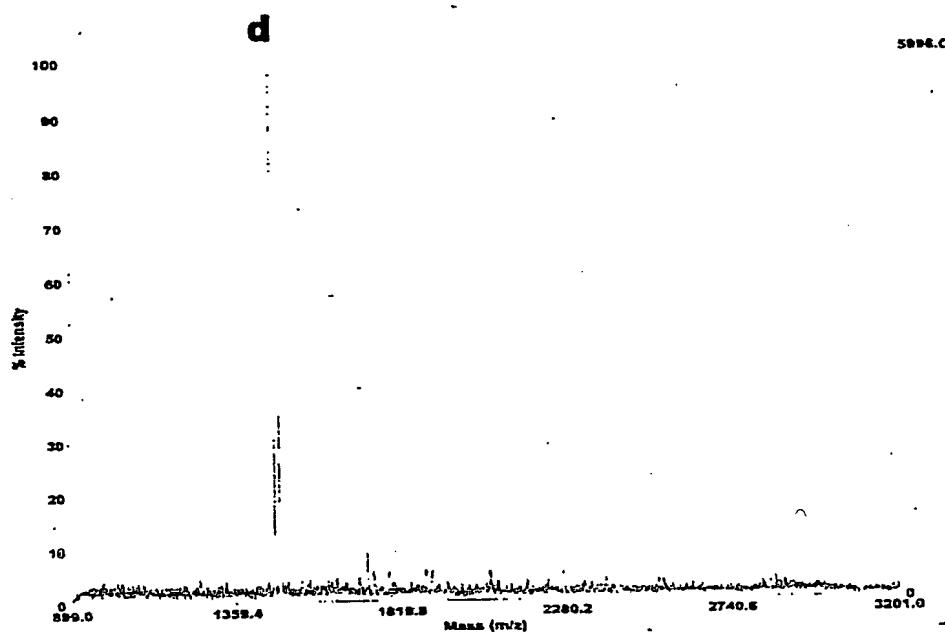
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SSARGPARRPMRHVLSSRERLGSRGTRRKWECVCLPGWHGPSCGVPTVQSNLPTKERLVPREVPRRV
INAININHEFDLLDVRFHETLGDVVDAFVVCDNSNFTAYGEPRPLKFREMLTNGTFEYIRHKVLYVFLDHFP
PGGRQDGWIADDYLRTFLTQDGVSRLRNLRPDDVFIIDDADEIPARDGVFLKLYDGWTEPPFAFHMRKSL
YFFFWKQPGTLEVVSGCTMDMLQAVYGLDGIRLRRRQYYTMPNFRQYENRTGHILVQWSLGSPLHFAGWH
CSWCFTPEGIYFKLVSAQNGDFPRWGDYEDKRDLNYSRSLIRTGGWFDTQQEYPPADPSEHMYAPKYL
KNYDQFRYLLLENPYREPKSTVEGGRQNQSGDGRSSAVRGKLDTAEG

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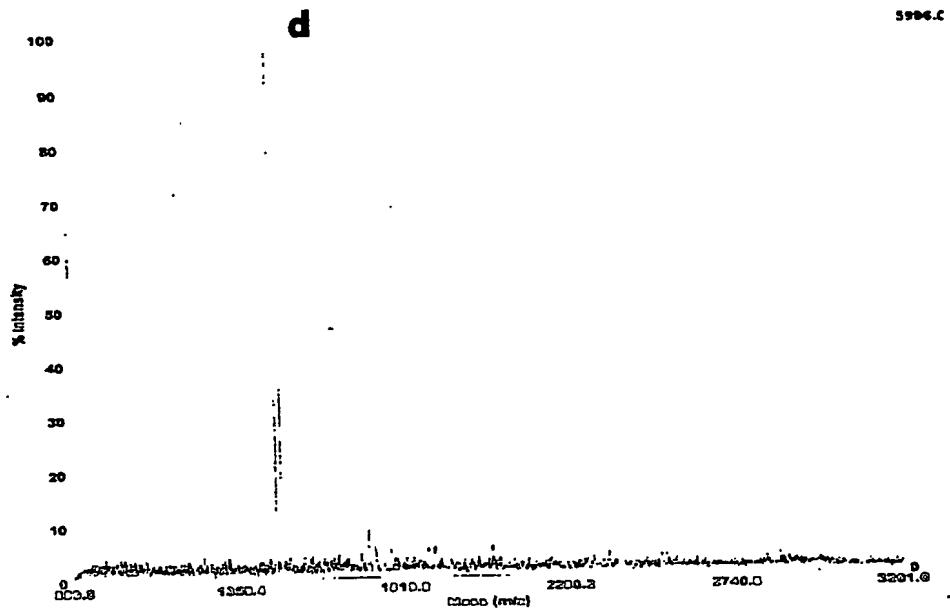
FIG. 24

Fig. 25

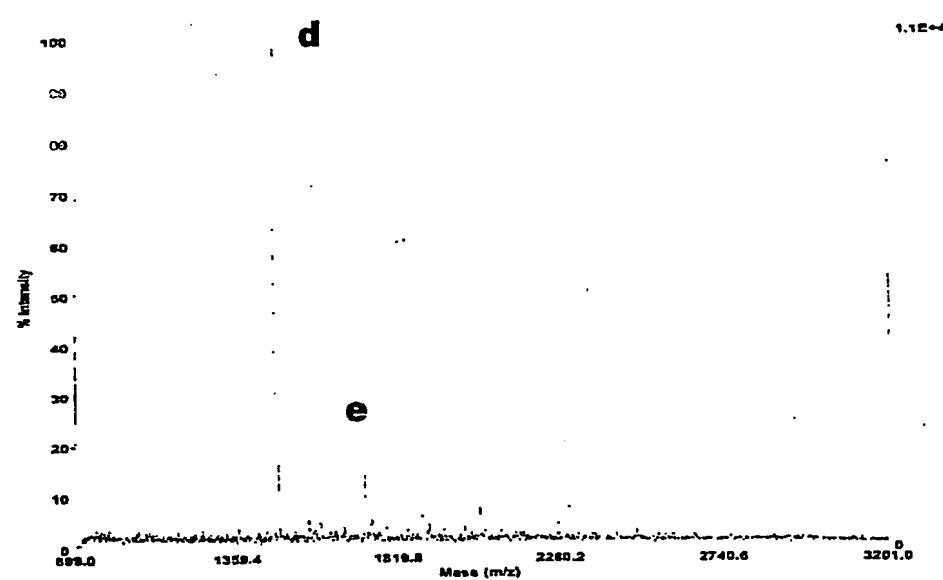
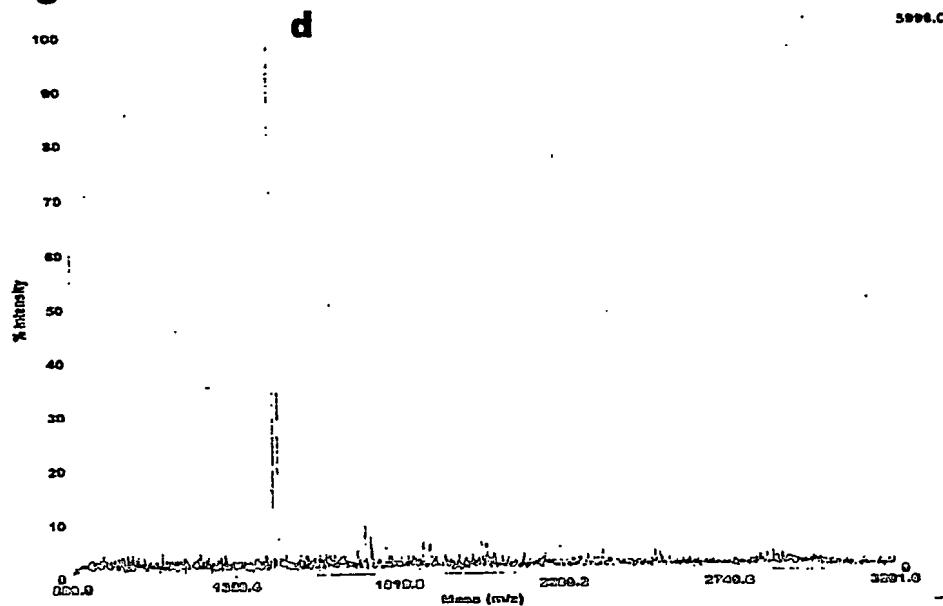
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Fig. 26

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Fig. 27

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Fig. 28

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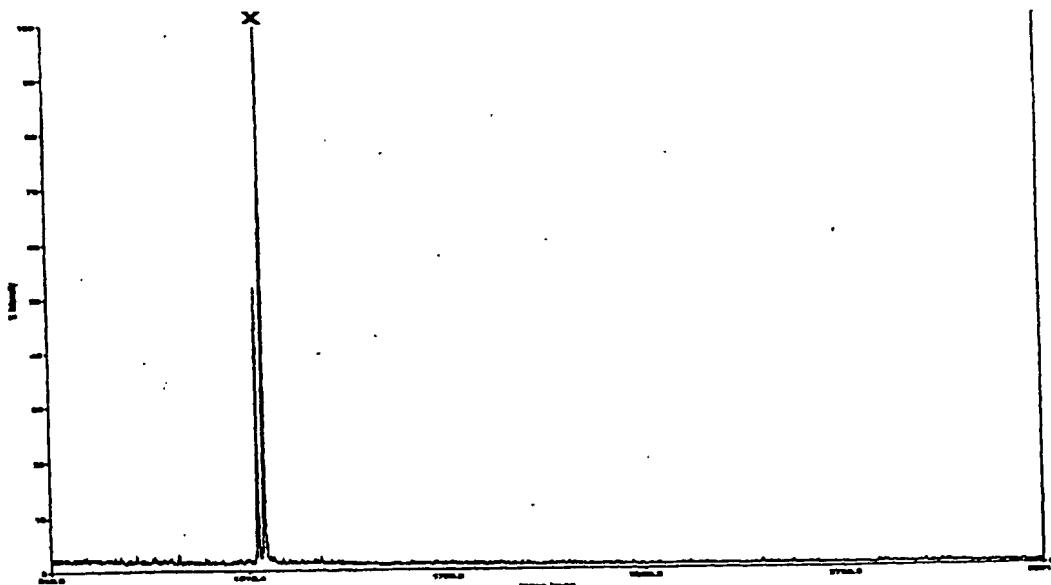
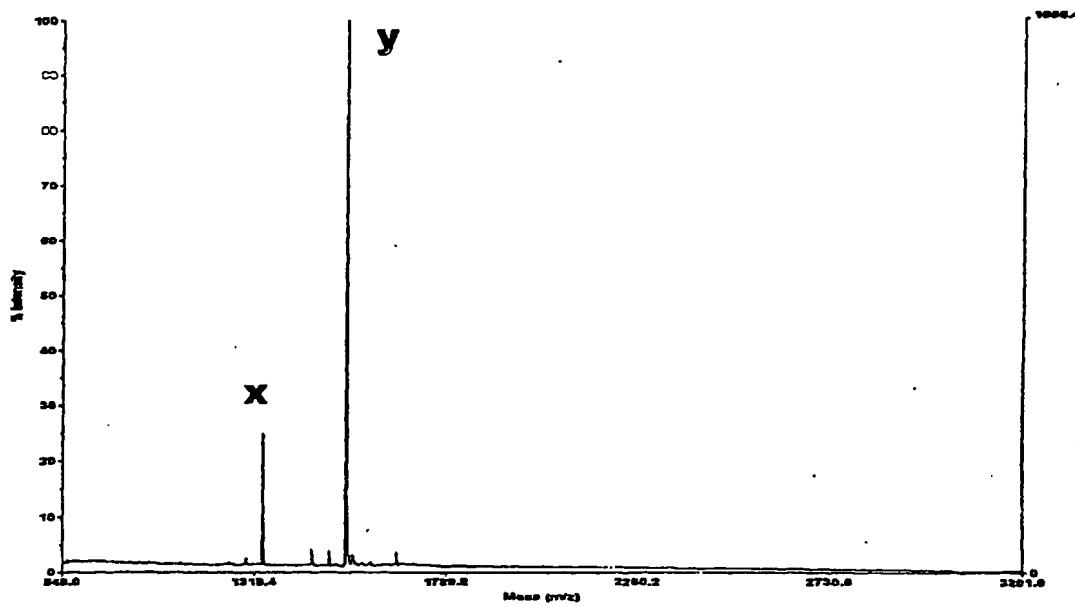
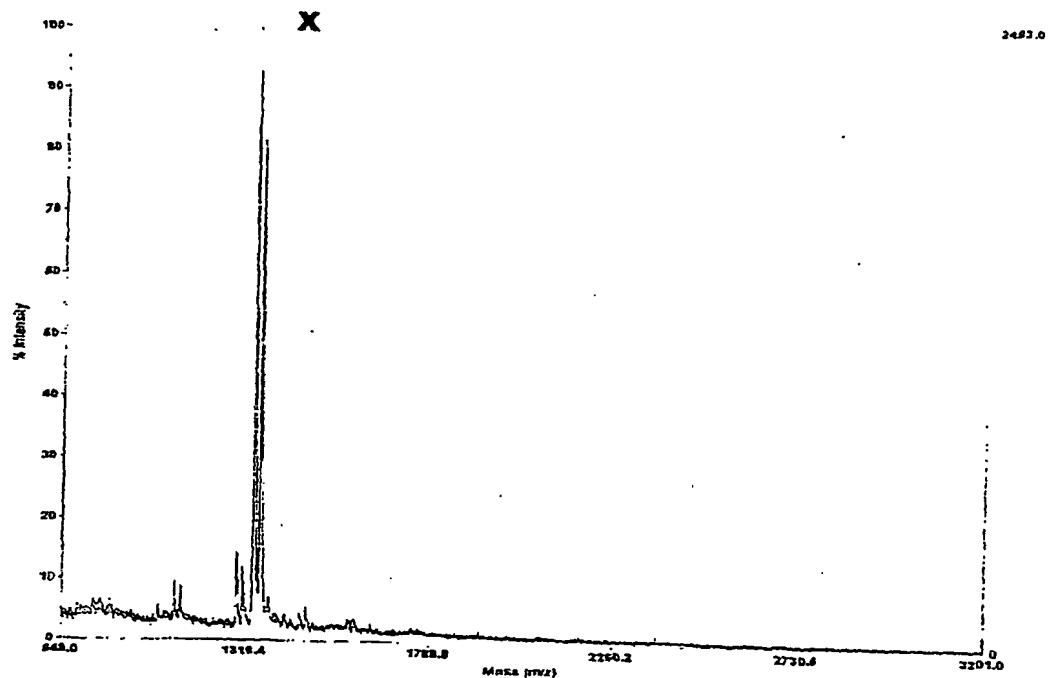
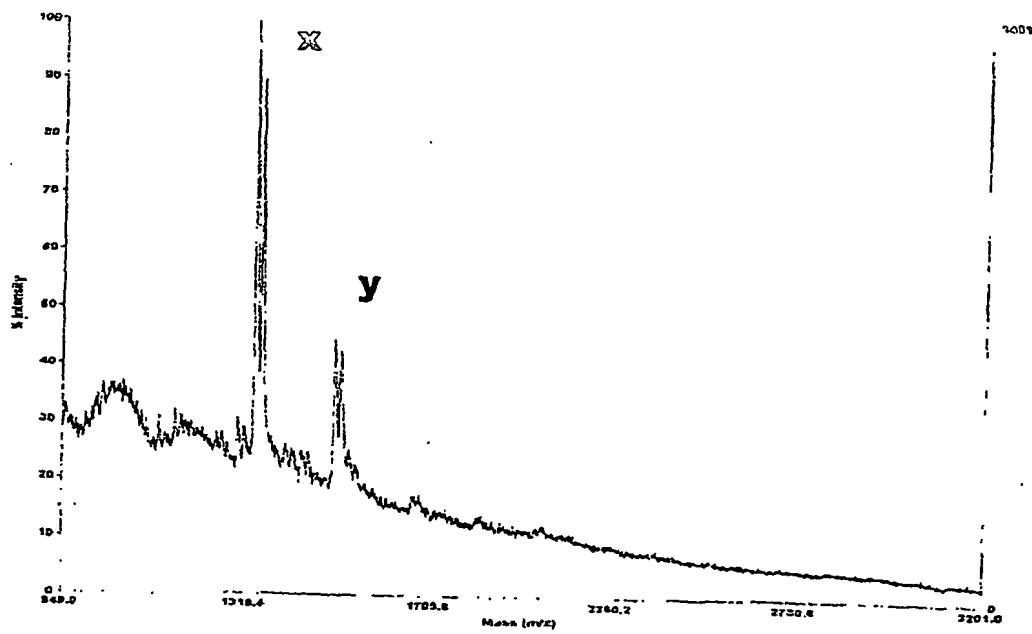
Fig. 29**Fig. 30**

Fig. 31

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**Fig. 32**

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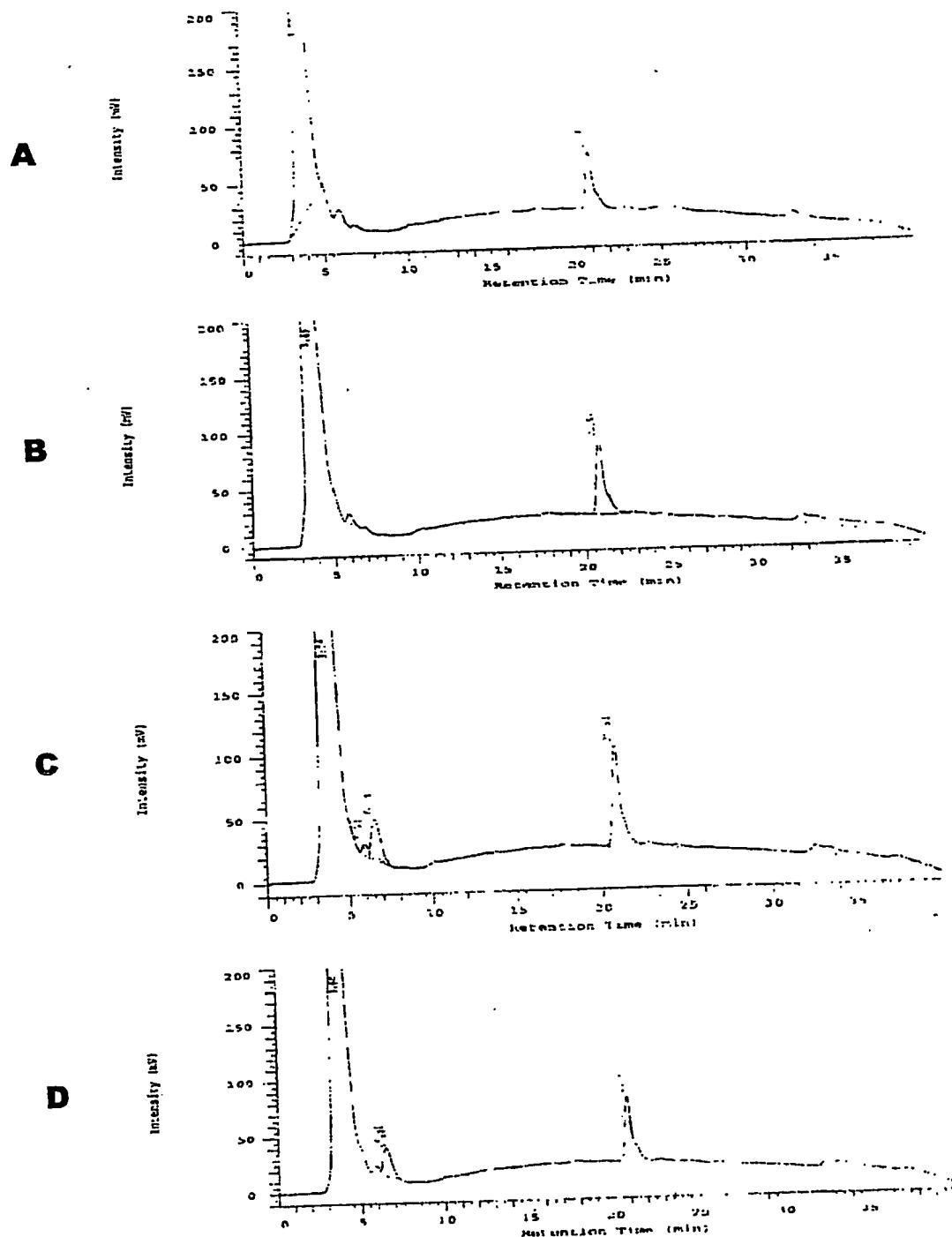


Fig. 33

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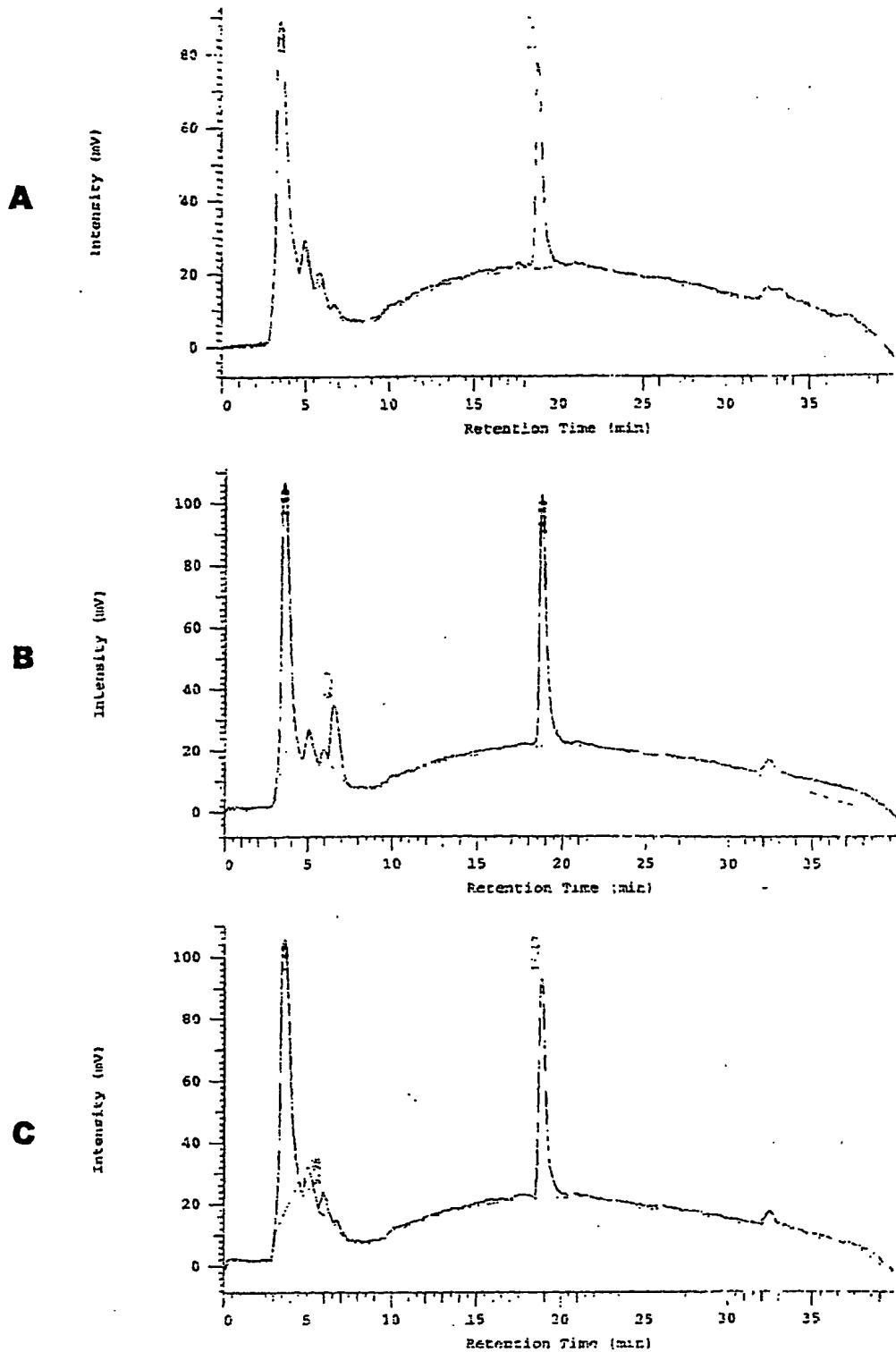


Fig. 34

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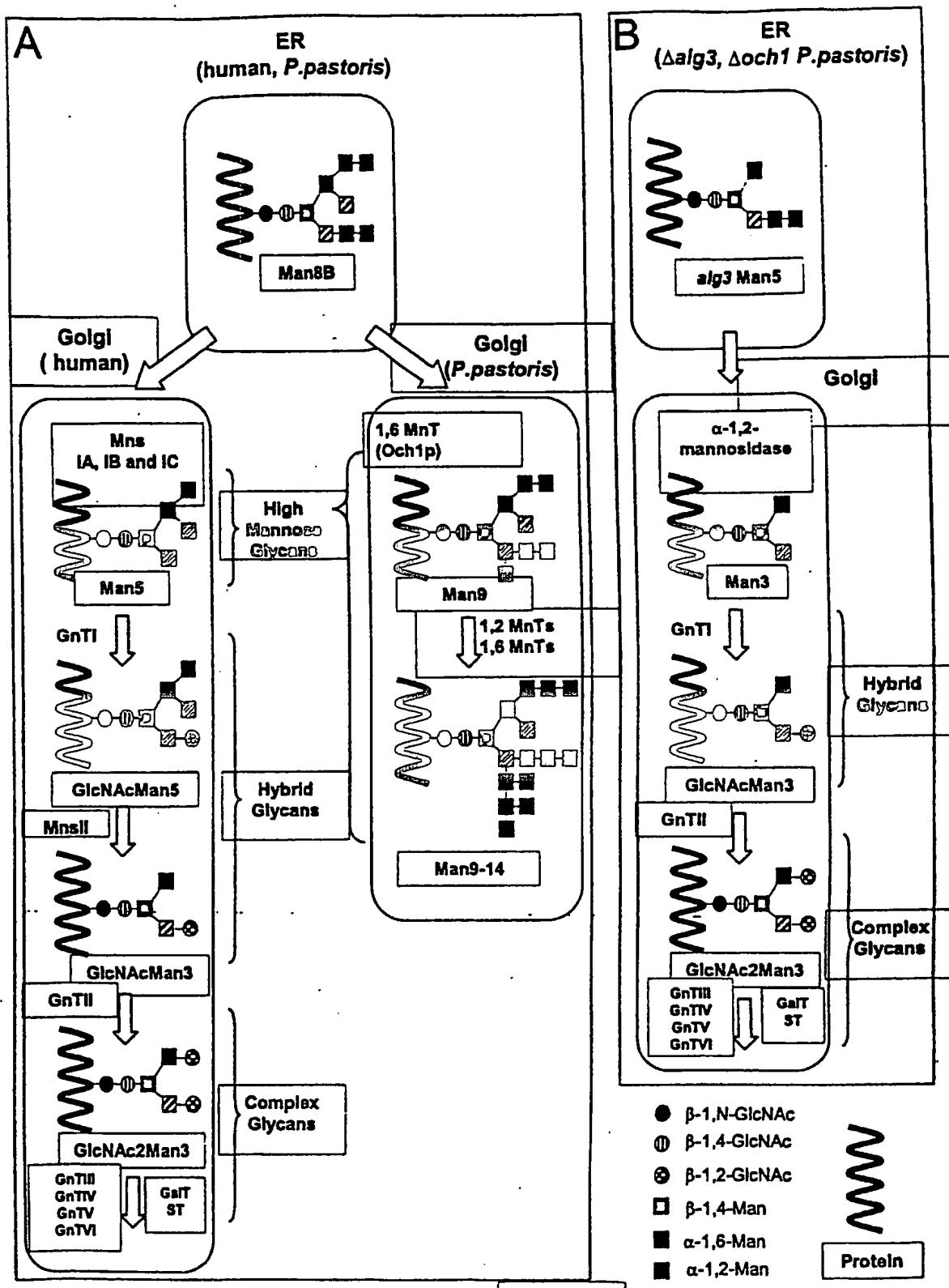
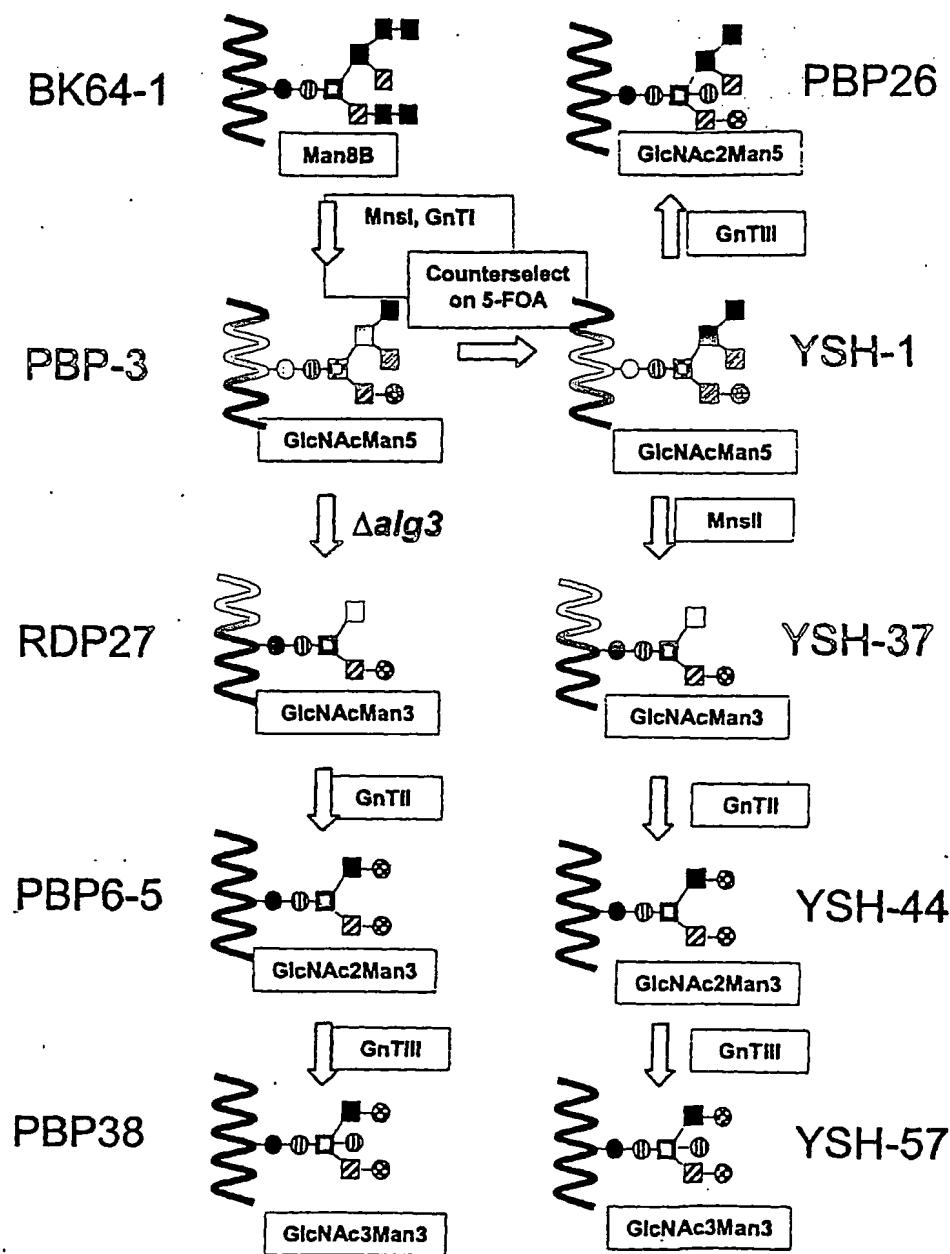
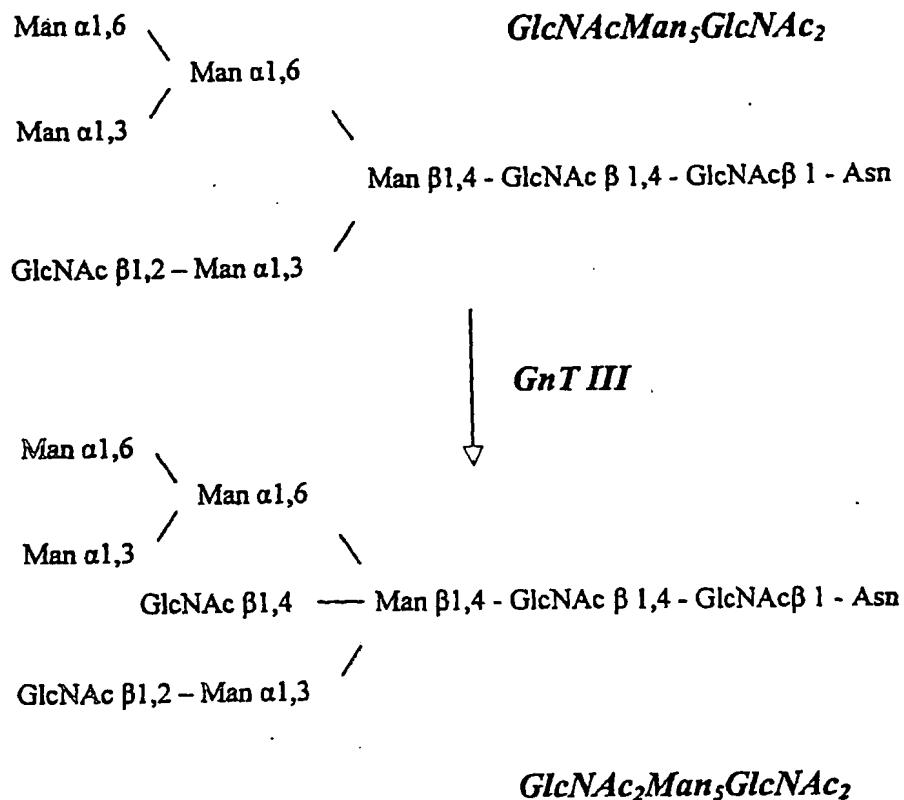


Fig. 35

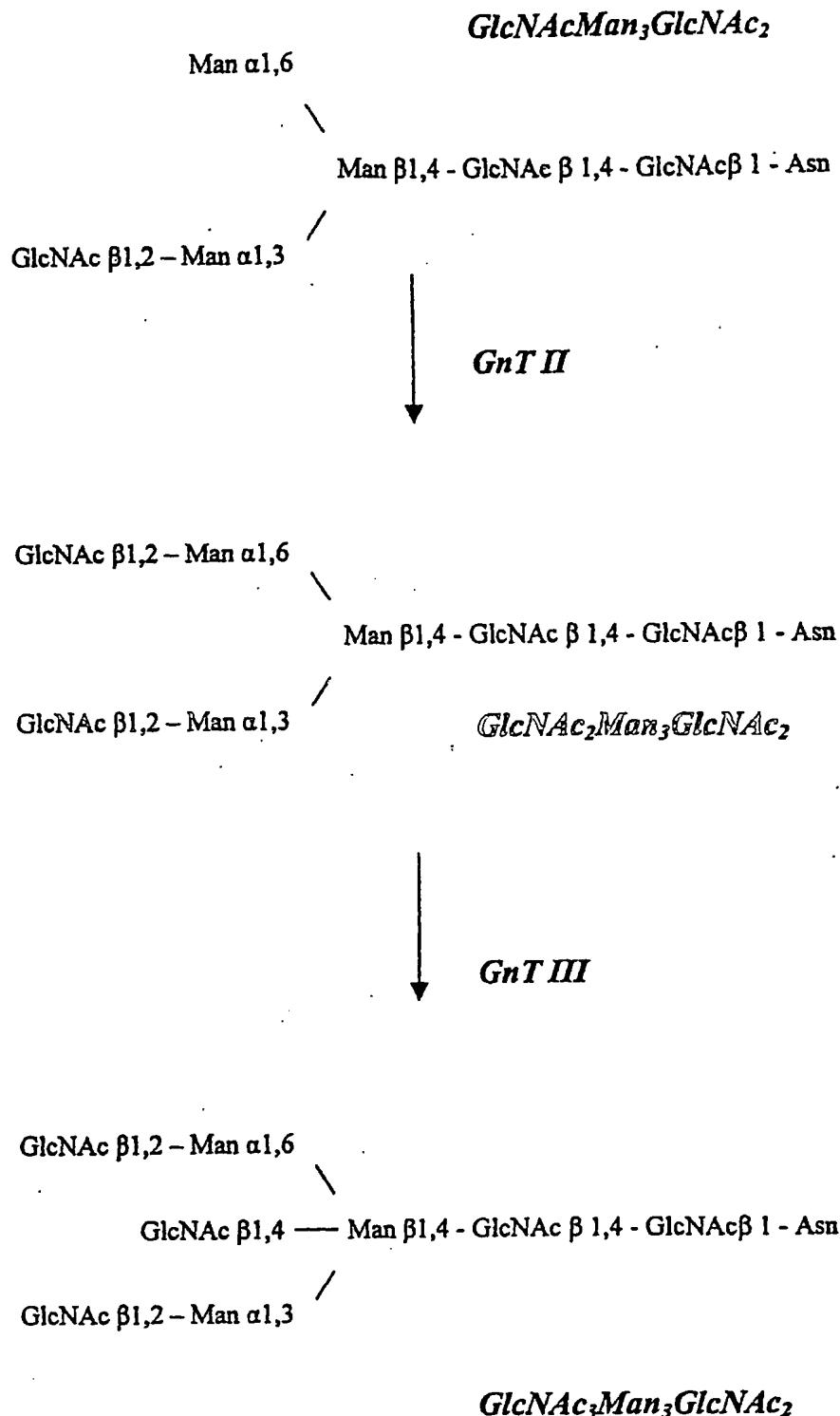
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Fig. 36

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Fig. 37

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Fig. 38

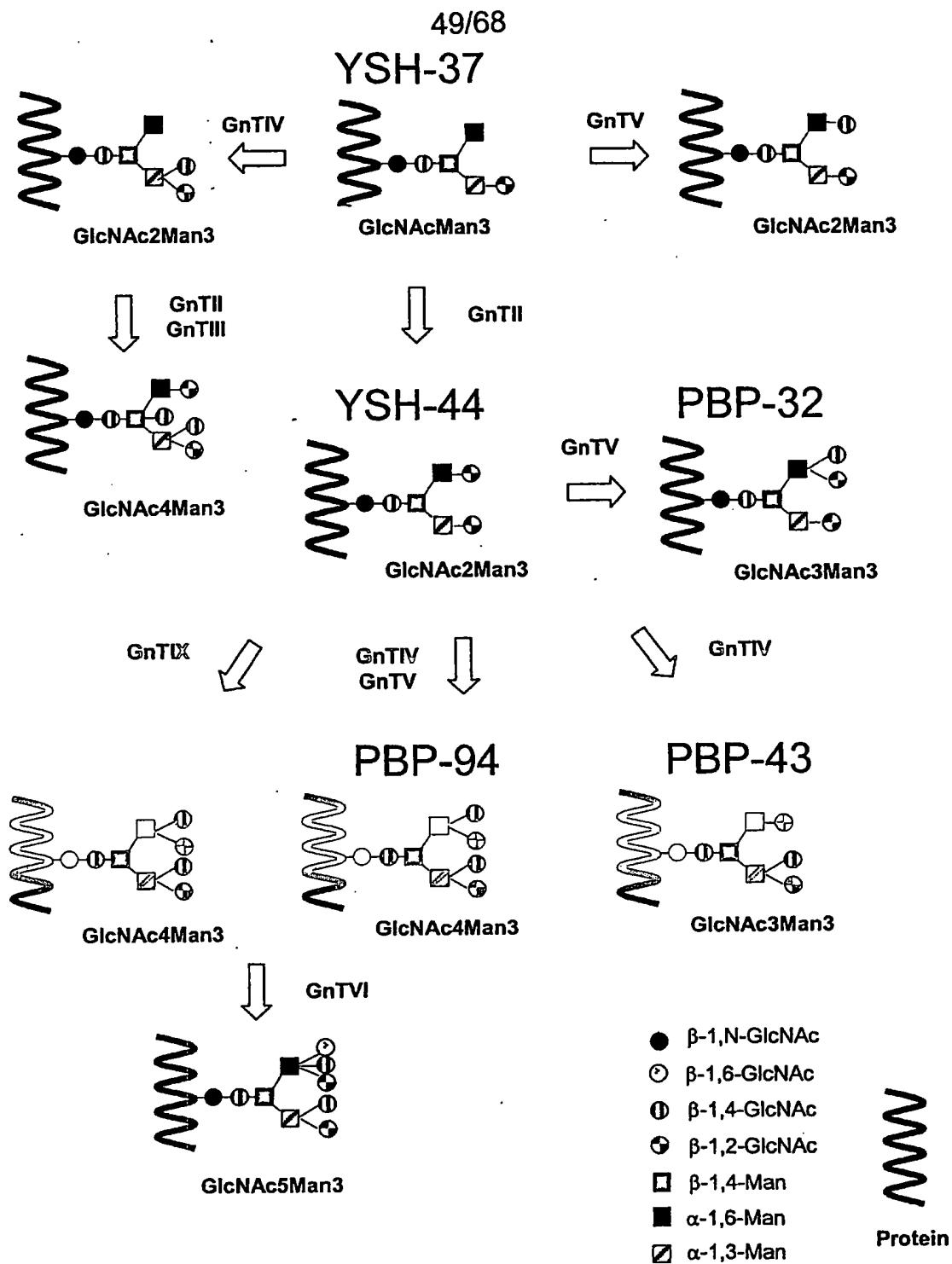


Fig. 39

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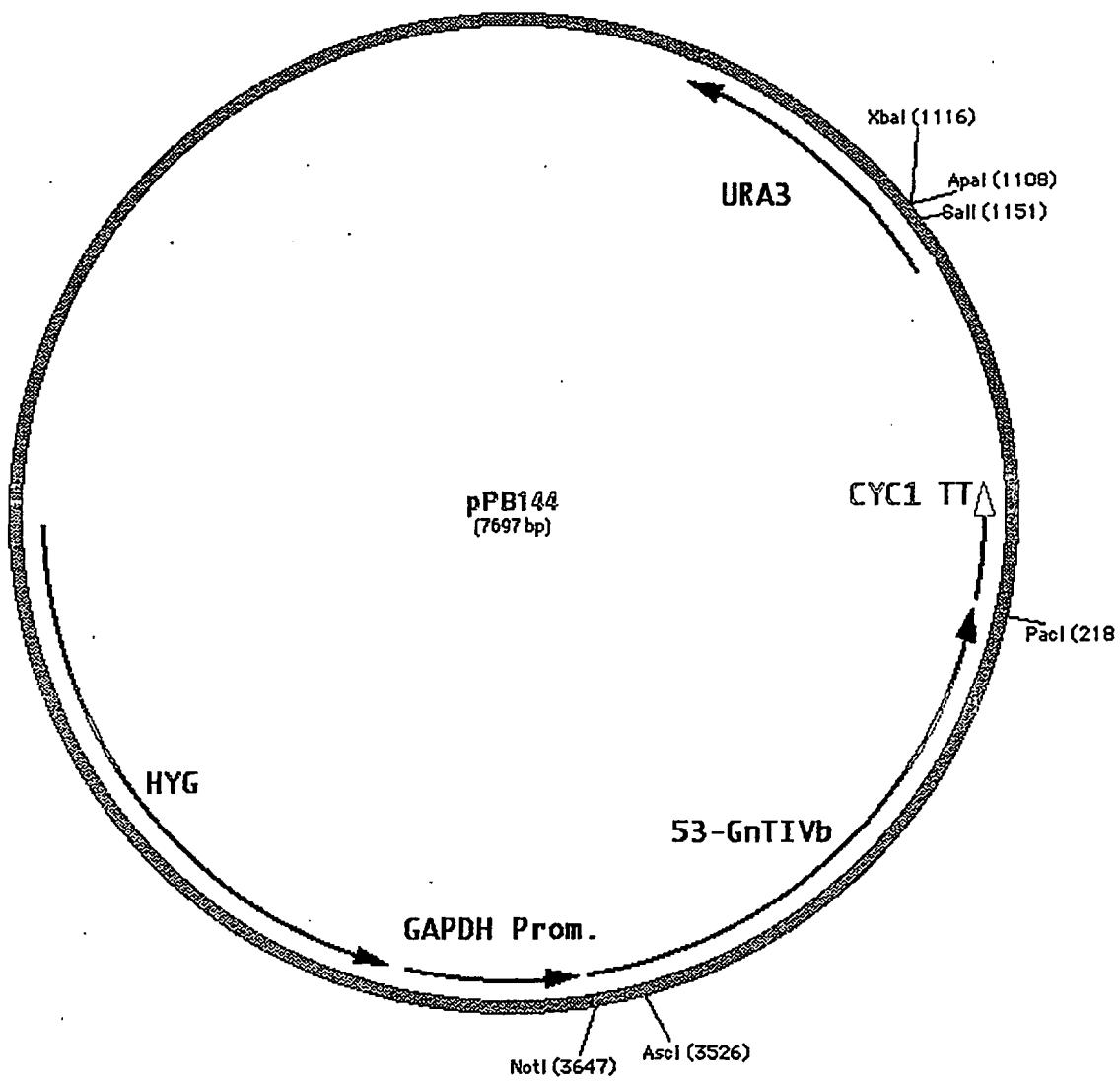
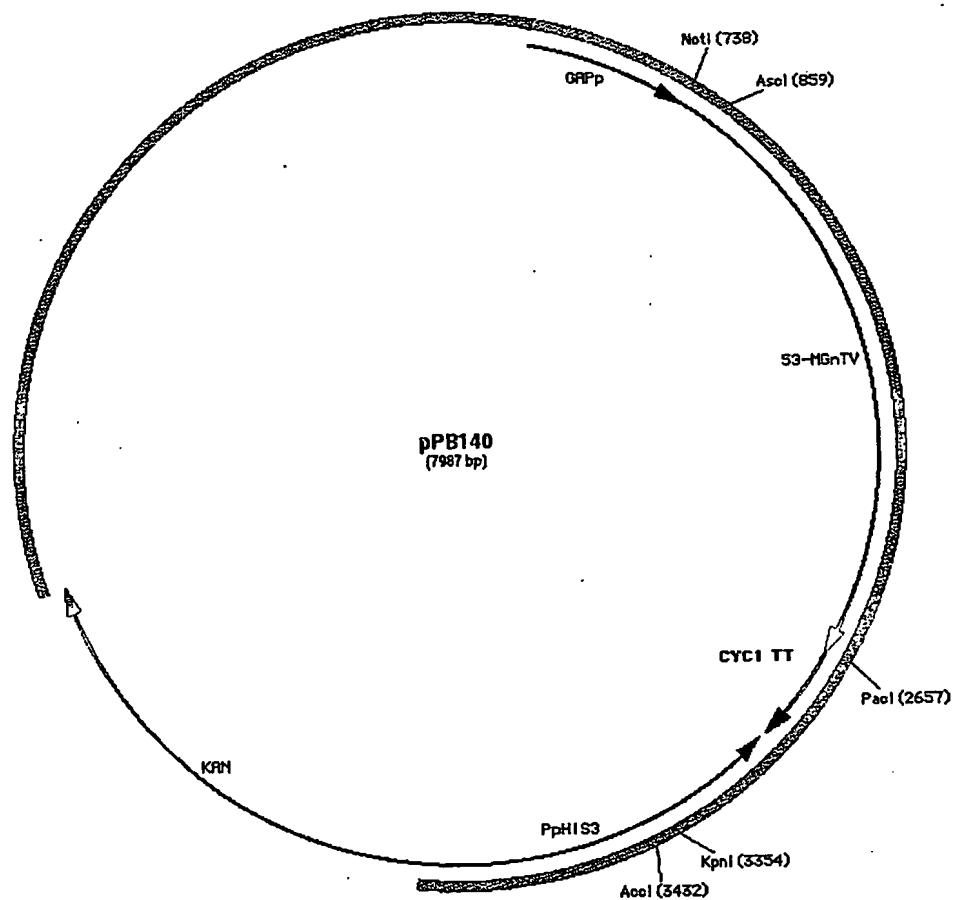
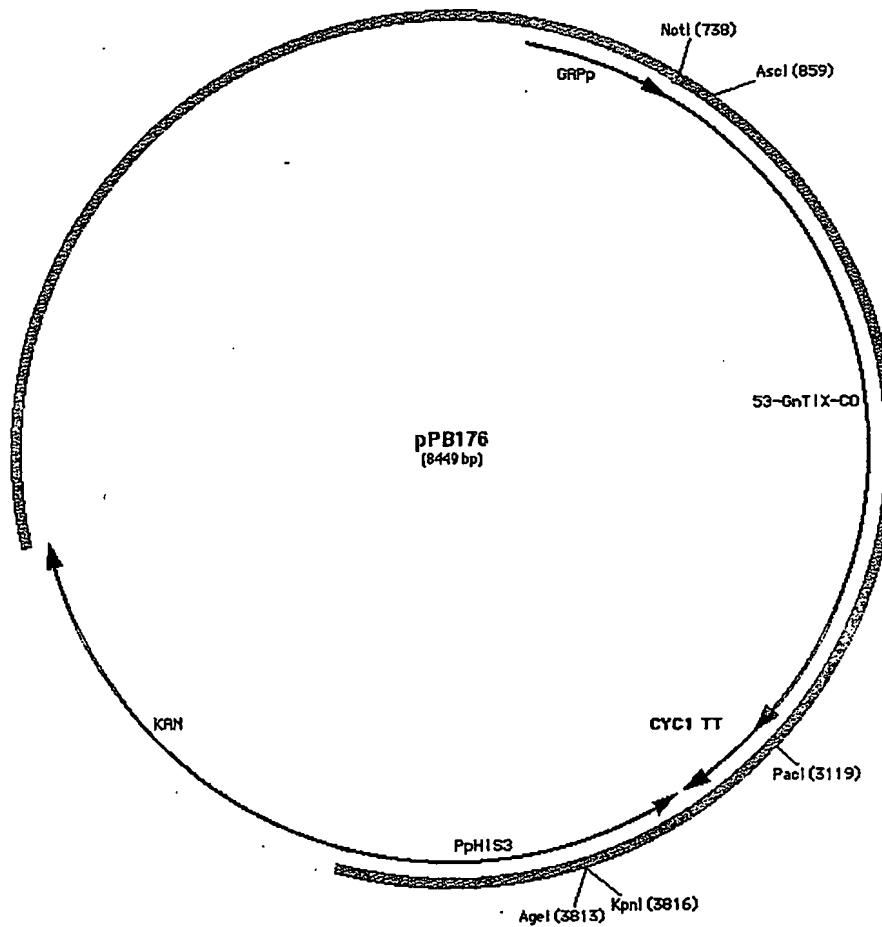


Fig. 40A

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**Fig. 40B**

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**Fig. 40C**

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Figure 41

Homo sapiens mannosyl (alpha-1,3)-glycoprotein beta-1,4-N-acetylglucosaminyltransferase, isoenzyme A (MGAT4A)
Accession number NM_012214.

MRLRNGTVATALAFITSFLTLSWYTTWQNGKEKLIAYQREFLALKERLRIAEHRIQSRS
 SELNTIVQQFKRVAETNGSKDALNKFDNTLKLLKELTSKKSLQVPSIYYHLPHLKKN
 EGSLQPAVQIGNGRTGVSIVMGIPTVKREVKSYLIETLHSЛИDNLYPEEKLDCVIVF
 GETDIDYVHGUVANLEKEFSKEISSGLVEISPPESYPPDLTNLKETFGDSKERVRWRT
 KQNLDYCFLMMYAQEKGIIYIQLLEDDIIVKQNYFNTIKNFALQLSSEEWILEFSQLGF
 IGKMFQAPDLTILIVEFIFMFYKEKPIDWLLDHILWVKVCNPEKDAKHCDRQKANLRIRF
 RPSLFQHVGLHSSLGKIQKLTDKDYMKPPLLKIHVNPPEVSTSLKVYQGHTLEKTYM
 GEDFFWAITPIAGDYILFKFDKPVNVESYLFHSGNQEHPGDILLNTTVEVLPFKSEGLE
 ISKETKDKRLEDGYFRIGKFENGVAEGMVDPDSLNPISAFRLSVIQNSAVWAILNEIHK
 KATN

Homo sapiens mannosyl (alpha-1,3)-glycoprotein beta-1,4-N-acetylglucosaminyltransferase, isoenzyme A (MGAT4A), mRNA

GAAATGAACCTCTCTTATTGATTTTATTGGCCTAGAGCCAGGAGTACTGCATTCAAGTT
 GACTTTCAAGGTAAAAGAAAACAGTCCTGGTGTGTCATCATAAACATATGGACCAAG
 TGTGATGGTGAATGAGATGAGGCTCGCAATGGAACGTAGCCACTGCTTAGCATT
 ATCACTCCTCCTTACTTTGTCTGGTATACATGGCAAATGGAAAGAAAAACT
 GATTGCTTATCAACGAGAATTCTTGCTTGAAAGAACGCTTCGAATAGCTGAACACA
 GAATCTCACAGCCTCTGAATTAAATACGATTGTGCAACAGTTCAAGCGTGTAGGA
 GCAGAAACAAATGGAAGTAAGGATGCGTTGAATAAGTTTCAGATAATACCTAAAGCT
 GTTAAAGGAGTTAACAGAAAAACTCTTCAAGTGCCAAGTATTATTATCATTGC
 CTCATTATTGAAAAATGAAGGAAGTCTCAACCTGCTGTACAGATTGGCAACGGAAGA
 ACAGGAGTTCAATAGTCATGGGCATTCCCACAGTGAAGAGAGAAAGTTAAACTTACCT
 CATAGAAACTCTTCAATTCCCTTATTGATAACCTGTATCCTGAAGAGAAAGTTGGACTGTG
 TTATAGTAGTCTTCAAGGAGACAGATATTGATTATGTACATGGTGTAGCCAAC
 CTGGAGAAAGAATTCTAAAGAAATCAGTTCTGGCTTGGTGGAAAGTCATATCACCCCC
 TGAAAGCTATTATCCTGACTTGACAAACCTAAAGGAGACATTGGAGACTCCAAGAAA
 GAGTAAGATGGAGAACAAAGCAAACCTAGATTACTGTTCTAAATGATGTATGCTCAA
 GAAAAGGGCATATTACATTACAGCTGAAGATGATATTATTGTCAAACAAAATTATT
 TAATACCATAAAAATTTCGACTTCAACTTTCTTGAGGAATGGATGATTCTAGAGT
 TTTCCCAGCTGGCTTCATTGGTAAATGTTCAAGCGCCGGATCTTACTCTGATTGTA
 GAATTCAATTATGTTACAAGGAGAACCCATTGATTGGCTCTGGACCATATT
 TCTGGGTGAAAGTCTGCAACCTGAAAAAGATGCAAAACATTGTGATAGACAGAAAGCA
 AATCTGCGAATTCCCTCAGACCTTCCCTTCAACATGTTGGCTGCACTCATCACT
 ATCAGGAAAAACTCAGGATAAAAGATTATATGAAACCATTACTCTTAA
 TCCATGTAACCCACCTGCGGAGGTACTACTTCTGAAAGGTCTACCAAGGGATACG
 CTGGAGAAAACCTACATGGGAGAGGATTCTCTGGCTATCACACCGATAGCTGGAGA
 CTACATCTGTTAAATTGATAAACACAGTCAATGTAGAAAGTTATTGTCAG
 GCAACCAAGAACATCCTGGAGATATTCTGCTAAACACAACACTGTGGAAGTTGCCTT
 AAGAGTGAAGGTTGGAAATAAGCAAAGAAACAAAGACAAACGATTAGAAGATGGCTA
 TTTCAGAATAGGAAAATTGAGAATGGTGTGAGAAGGAATGGTGGATCCAAGTCTCA

Figure 41 (Cont.)**54/68**

ATCCCATTCA~~G~~C~~T~~TCAGTT~~C~~AGTT~~A~~TCAGAATTCTGCTGTTGGGCCATTCTT
AATGAGATT~~C~~AT~~A~~TTAAAAAGCCACCAACTGATCATCTGAGAAACCAACACATT~~T~~TTT
CCTGTGA~~A~~TTTGT~~T~~TAATTAAAGATA~~G~~T~~A~~AGCATGTATCTTTTTTATTCTACTTGA
ACACTAC~~C~~CTTGTGAAGT~~C~~ACTGTAGATAAGACGATTGT~~C~~ATTCCACTTGGAAAGT
GAATCTCCATAATAATTGTATTGTTGAAACTAAGCTGCCTCAGATTTA~~A~~CTTGA
CTCAAACATTTCAATTATGACAGCCTGTTAATATGACTTG~~T~~ACTATT~~T~~GGTATTAT
ACTAATACATAAGAGTTGTACATATTGTTACATTCTTAAATTGAGAAAAACTAATGT
TACATACATTTATGAAGGGGGTACTTTGAGGTTCACTTATT~~T~~ACTATT

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Figure 42

Homo sapiens mannosyl (alpha-1,3)-glycoprotein beta-1,4-N-acetylglucosaminyltransferase, isoenzyme B (MGAT4B)
Accession number NM_014275.

MRLRNGTFLTLLLFCFLCAFLSLSWYAALSGQKGDVVDVYQREFLALRDRLHAAEQESLK
RSKELNLVLDEIKRAVSERQALRGDGNRWTGRLTEDPRLKPWNGSHRHVLHLPTVFHH
LPHLLAKESSLQPAVRVGQGRGVSVVMGIPSVRREVHSYLTDTLHSLISELSPQEKE
SVIVVLIAETDSQYTSAVTENIKALFPTEIHSGLLEVISPSPHFYPDFSRLRESFGDPK
ERVRWRTKQNLDYCFLMMYAQSKGIYYVQLEDDIVAKPNYLSTMKNFALQQPSEDWMIL
EFSQLGFIGKMFKSLDLSLIVEFILMFYRDKPIDWLDDHILWVKVCNPEKDAKHCDRQK
ANLRIRFKPSLFQHVGTHTSSLAGKIQKLKDKGKQALRKEHVNPPEAVSTSLKTYQHF
TLEKAYLREDFFWAFTPAAAGDFIRFRFFQPLRLERFFFSGNIEHPEDKLFNTSVEVLP
FDNPQSDKEALQEGRATLRYPRSPDGYLQIGSFYKGVAEGEVDPAGPLEALRLSIQT
DSPVWVILSEIFLKKAD

Homo sapiens mannosyl (alpha-1,3)-glycoprotein beta-1,4-N-acetylglucosaminyltransferase, isoenzyme B (MGAT4B), mRNA.

TGCAGCCTCGGCCCGGGCGCCGCCGCGCACCCGAGGAGATGAGG
CTCCGCAATGGCACCTTCCTGACGCTGCTCTCTGCCTGTGCGCCTT
CCTCTCGCTGTCTGGTACGGCACTCAGCGGCCAGAAAGGCGACGTT
GTGGACGTTTACCAAGCGGGAGTTCTCTGGCGCTGCGCAGTCGGTTGCACG
CAGCTGAGCAGGAGAGCCTCAAGCGCTCCAAGGAGCTCAACCTGGTGCT
GGACGAGATCAAGAGGGCCGTGTCAAGAAAGGCAGGCCCTGCCAG
AGACGGAATCGCACCTGGGCCCTAACAGAGGACCCCCGATTGAAG
CCGTGGAACGGCTCACACCGGACGTGCTGCACCTGCCACCGTCTTCC
ATCACCTGCCACACCTGCTGGCCAAGGAGAGCAGTCTGCAGCCCAG
GTGGGCCAGGGCCGCACCGGAGTGTGGTGTGGTATGGGCATCCCGAGCGTGC
GGTGCACTCGTACCTGACTGACACTCTGCACTCGCTCATCTCGAGCTGAG
AGAAGGAGGACTCGGTACGTGGTGTGATGCCAGACTGACTCACAGTAC
GCAGTGACAGAGAACATCAAGGCCTGTTCCCCACGGAGATCCATT
GGTCATCTCACCCCTCCCCCCTACCTCTACCCCTGACTTCTCCCGCT
GGGACCCCCAAGGAGAGACTCAGGTGGAGGACAAACAGAACCTCG
ATGATGTACGCGCAGTCAAAGGCATCTACTACGTGCACTGGAGGAT
CAAGCCCAACTACCTGAGCACCATGAAGAACTTGCAC
GGATGATCCTGGAGTTCTCCAGCTGGGCTTCATTGGTAAGATGTT
CTGAGCCTGATTGTAGAGTTCATCTCATGTTCTACCGGGACA
CCTGGACCATATTCTGTGGGTAAAGTCTGCAACCCGAGAAGGAT
ACCGGCAGAAAGCCAACCTGCGGATCC
GCTTCAAACCGTCCCTCTTCCAGCACGTGGCACTCACTCCTCG
CAGAAACTGAAGGACAAAGACTTGGAAAGCAGGCGCTGCC
GCCAGCAGAGGTGAGCACGAGCCTGAAGACATACC
ACCTGCGCAGGACTTCTTCTGGCCTTCACCCCTGCCGGGGACT
ACCTCGCTTC

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Figure 42 (Cont.)

CGCTTCTTCCAACCTCTAACAGACTGGAGCGGTTCTCTTCCGCAGTGGGAACATCGAGCA
CCC GGAGGACAAGCTCTTCAACACGTCTGTGGAGGTGCTGCCCTTCGACAACCCTCAGT
CAGACAAGGAGGCCCTGCAGGAGGGCCGCACC GCCACCCCTCCGGTACCCCTCGGAGGCC
GACGGCTACCTCCAGATCGGCTCCTCTACAAGGGAGTGGCAGAGGGAGAGGTGGACCC
AGCCTTCGGCCCTCTGAAGACTGCGCCTCTCGATCCAGACGGACTCCCCGTGTGGGG
TGATTCTGAGCGAGATCTTCCTGAAAAAGGCCGACTAAGCTGCGGGCTCTGAGGGTAC
CCTGTGGCCAGCCCTGAAGCCCACATTCTGGGGGTGTCGTCACTGCCGTCCCCGGAGG
GCCAGATA CGGCCCCGCCAAAGGGTTCTGCCTGGCGT CGGGCTTGGGCCGGCTGGGG
TCCGCCGCTGGCCGGAGGCCCTAGGAGCTGGTGCTGCCCGGCCGGCGCGGA
GGAGGCAGGC GGCCCCCACACTGTGCCTGAGGCCGGAACCGTTCGCACCCGGCCTGCC
CCAGTCAGGCCGTTTAGAAGAGCTTTACTTGGGCGCCCGCGTCTGGCGCGAAC
CTGGAATGCATATACTACTTTATGTGCTGTGTTTTATTCTGGATAACATTGATT
TTCACGTAAGTCCACATATACTTCTATAAGAGCGTGACTTGTAAATAAGGGTTAATGAA
G

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Figure 43

Mus musculus N-acetylglucosaminyltransferase V (Mgat5)

Accession number AF474154.

MAFFSPWKLSSQKLGFFLVTFGFIWGMMLLHFTIQQRTPESSSMLREQL
 LDLSKRYIKALAEENRDVVDGPyAGVMTAYDLKKTLAVLLDNILQRIGKL
 ESKVDNLVNGTGANSTNSTTAVPSLVSLEKINVADIINGVQEKCVLPPMD
 GYPHCEGKIKWMKDMWRSDPCYADYGVDTSCSFFIYLSEVENWCPLPWRANKPYEE
 ADHNSLAEIRTDNFNLYGMKKHEEFRWMRLRIRR MADAWIQAISLAEKQNLEKRKRK
 KILVHLGLLTKESGFKIAETAFSGGPLGELVQWSDLITSLYLLGHDIRISASLAELKEI
 MKKVVGNRSGCPTVGDRIVELYIDIVGLAQFKKTLPSPWVHYQCLMRVLDGFTEPEF
 NHASYAQSKGHKTPWGKWNLNPQQFYTMFPHTPDNSFLGFVVEQHLNSSDIHHINEIKR
 QNQSLVYGVDSFWKNKKIYLDI IHTYMEVHATVYGSSTKNIPSYVKNHGILSGRDLQF
 LLRETKLFVGLGFPYEGPAPLEAIANGCAFLNPKFNPPKSSKNTDFFIGKPTLRELTSQ
 HPYAEVFIGRPHVWTVDLNREEVEDAVKAILNQKIEPYMPYEFTECGMLQRINAFIEK
 QDFCHGQVMWPPLSALQVKLAEPGOSCKQVCQESQLICEPSFFQHLNKEKDLLKYKVTC
 QSSELYKDILVPSFYPKSKHCVFQGDLLLFSAGAHPTHQRICPCRDFIKGQVALCKDC
 L

Mus musculus N-acetylglucosaminyltransferase V (Mgat5) mRNA

ATTGCTAGAGAGAGATGGCTTCTTTCTCCCTGGAAGTTGTCCCTCTCAG
 AAGCTGGGCTTTCTGGTGACTTTCTGGCTTCATCTGGGGCATGATGCT
 TCTGCACCTCACCATCCAGCAGCGGACTCAGCCCCAGAGAGCAGCTCCATG
 TTACGGGAGCAGATCCTGACCTCAGCAAGAGGTACATTAAGGCAGTGG
 CAGAGGAGAACAGGGACGTGGTGGATGGCCCTACGCTGGTGTACAGCAGCCTATG
 ATCTGAAGAAAACGCTGCCGTCTGGATAACATCCTGCAGCGCATTGCAAGCTC
 GAGTCAAAGGTGGACAATCTGGTCAACGGCACAGGAGCGAACTCCACCAACTCCACCAC
 GGCTGTCCCCAGCTTGGTGTGCTTGAGAAAATTAAATGTGGCAGATATCATTAAATGGAG
 TTCAGGAAAAATGTGTATTGCCTCCTATGGATGGCTACCCCCACTGCGAGGGAAAATC
 AAGTGGATGAAGGACATGTGGCGCTGGACCCCTGCTACGAGACTATGGAGTGGACGG
 GACCTCCTGCTCCTTTTATTACCTCAGTGAGGGTGGAAAATTGGTGTCCCTCGTTAC
 CTTGGAGAGCAAAAATCCCTATGAAGAAGCTGATCATAACTCATTGGCGGAAATCCGT
 ACGGATTAAACATTCTCACGGCATGATGAAGAAGCAGCAGGAGTCCCGTGGATGAG
 GCTTCGGATCCGGGAATGGCTGACGCGTGGATCCAAGCTATCAAGTCTCTGGCGGAGA
 AACAAAACCTTGAGAAGAGGAAACGGAAGAAAATCCTGTTCACCTGGGCTCCTGACC
 AAGGAATCGGGCTCAAGATTGGAGACAGCATTAGCGGTGGCCCTCTGGTGAAC
 CGTTCAGTGGAGTGACTTAATCACATCTGTACCTGCTGGGCCATGACATCCGGATCT
 CGGCCTCACTGGCTGAGCTAAGGAGATAATGAAGAAGGTTGGAAACCGGTCTGGC
 TGTCCAACGTAGGAGACAGAATCGTGAGCTGATTATATCGATATTGTGGGACTTGC
 TCAATTAAAGAAAACACTAGGGCCATCCTGGGTTCATTA
 CCAGTGCATGCTCCGGTGTAGACTCCTTGGAACAGAACCTGAGTTCAATCATGCGA
 GCTATGCCAGTCAAAGGCCACAAGACCCCCCTGGGAAAGTGGAACTGAACCCGCA
 CAGTTTACACCATGTTCCCTCATACCCCAGACAACAGCTTCTGGCTCGTGGTGG
 GCAGCACCTGAACTCCAGCGACATTACCCACATCAACGAGATCAAAGGCAGAACCA
 CCTTGTGTATGGAAAGTGGATAGTTCTGGAAGAATAAGAAAATCTACCTGGATATC

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Figure 43 (Cont.)

ATTCACACGTACATGGAAGTGCACGCCACTGTTATGGCTCCAGTACCAAGAACATTCC
CAGTTACGTGAAAACCATGGCATTCTCAGTGGACGTGACCTGCAGTTCTTCTCCGGG
AAACCAAGCTGTTGGCTCGGATCCCTTATGAAGGCCAGCTCCCTGGAGGCC
ATCGCGAATGGATGTGCTTCCCTGAACCCAAGTTCAACCCCTCCAAAAGCAGCAAAA
CACAGACTCTTCAATTGGCAAGCCAACACTGAGAGAGCTGACATCCCAGCATCCTTACG
CAGAAGTCTTCAATCGGCCGGCCACACGTCTGGACTGTGGATCTCAATAACCAGAGGAA
GTTAGAAGATGCAGTAAAGCCATCTTAAACCAGAAGAGATTGAGCCGTATATGCCATATGA
GTTACATGTGAAGGCATGCTGCAGAGAAATCAACGCTTCATTGAAAAACAGGACTTCT
GCCATGGCCAAGTGTGATGTGGCCGCCCTCAGGCCCTGCAGGTTAACGCTGGCTGAGCCA
GGGCAGTCTGCAAACAGGTGTGCCAGGAGAGCAGCTATCTGCGAGCCATCCTTCTT
TCAACACCTCAACAAGGAAAGGACCTGCTGAAGTATAAGGTGACCTGCCAAAGCTCAG
AACTGTACAAGGACATCCTGGTGCCTCTACCCCAAGAGCAAGCAACTGTGTGTT
CAAGGGGACCTCCTGCTCTCAGTTGTGCCGGAGCCCACACACCAGGGATCTG
CCCCTGCCGGACTTCATCAAGGGCCAAGTGGCCCTCTGCAAAGACTGCCTATAGCAT
CGCTGCCCTGAATTAACACTCAGACGGGAAAGACGTGGCTCCACTGGGCAGGGCCAAGGGG
CACAAAGACATTAGGGACTCTGACAGAGCCTGAGATCTTGGTCCAGGGCTTGAGTT
TAGTACCGCTCCAGGCCACAGCCAGTGCATCCCAGTTACACAAAACCACAAGGAAACA
GGTTAGAACAGGAACCTGGTTCTCCTCAGTGTAAAGGAATGTCCCTCTGCTGGGAGA
TCGAGCGACTGTAGGGAAAGGATCCAGGCAGTTGCTCCGGGAATTTTTTTTTT
TTTTAAAGAAGGGATAAAAGTCCGGAGACTCATTCAAACGTAAAACAAAACAGGAAGA
GGGAATTGAGCCAATTGGGAAGGACTTTGGGCCGATCCTAAACCAATTAAATTATTTA
TTTGGGAGGATGGGGCGGGCTCGGGAGGGAGAGGGGTTGAACAGTTCTTTGT
TCCTCACTGTTAATTGCCACCTTCGGGCCCTCTGTCAGCGCCAAGCAGGGT
GCAGAGGGCTGTGGCTTGAGGGCCACTGTGGGCTTCACTCCTGGTCACAGGT
GGCAGCAGAGAAAAGAGATGTCTATAAGCAGGGGATGTAGCTCAGTTGAGAATGCT
TGCATAGCATAAATGAAGTCTGGTTCCATCCCCAGCACCACATAAAATGCAGGTAAGA
AACAGAGTCAGGAGGACCAAGCATTCTCCTGGTACATAACAAAAGCAAGGCCTTGT
CCCCATGTCCTGGTACAAGAGACCTATCTCAGAAAATTGTGGGGGGAGGGGGGGGG
AAATGGCCTTGAAAACACAGCCAGTCAGTCACTGTCAGTGCATTGCCAGAACTGGTGGATCCC
AGGTGTGCTTGGCAGATAACAGCTAAAGGCACATAACCTGGTGGGAAATAATGCC
TGTGGTGTCTGAGGGCCCCACCAAGTTCCAAAAAAAAAAAAAA

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Figure 44

Gallus gallus N-acetylglucosaminyltransferase VI, GnT-VI; Accession number AB040608

MRCSPKRSLTAVIAASFLLLLLHLRGSWQDPQEVRDLPDAVLKILKQGSLHIL
 QDTDNLCALHNISYHLLAGSPLPHKKFLAVGLSSVRRPRGYLPDTLQSLFKQSSEEEEL
 QEMVVVHLADADPIWNAQVAADISHRFAHHILLGRLVLIHTPHEFYPTLEGLKRNYND
 PEERVKFRSKQNVDYAFLLFTFAANLSSYYLMIEDDVWSAKSFTAIRKAVASQEGSNWA
 TLEFSKLGIGKLYIGRSSDLPLRARFLLFYQEMPCDWLLTHFRLLLTKQDKVIRFKPSLF
 QHMGlySSFQGTVNRLLEDDEFQADAMDLPDNPPAALFTNMVFENYEPSKAYSTARGYF
 WGKNPAVGSIIFSIVFHQPARVTRVRVQTGSSERPGDFLHAGVLELGRGRRADGRDCSVY
 TTVGTFEKGNNLEWRGLEKGMPNPVECVRIRVTQSQSEWLI IQSIGIWTAGT

Gallus gallus GnT-VI mRNA for *N*-acetylglucosaminyltransferase VI, complete cds

GGCACGAGGCAGAGCCCCAGCAGGGACGCCAGGGCCCAACGCCAACGCCACAG
 CTGCCACGCACCCGGCAGACCCAGTTGCATTATAATGAAAAATCCCAGGTAAACTC
 AGGAGCCTTGAGCCTCATGCCATGCCATGCCCTGCTGCGAGGAGAAACGTCTCAAGGGCACT
 AATGTCTCCATGAAGGGCTCTCAGCAGGCCGCTGGTGTATGCCGTGCTCCCCGAAA
 CGCTCCCTCACGGCTGTGATTGCGAGCCTCCTCCTCCTCCTCCTCCTCCTCCTCT
 GCACAGGGCAGCTGGCAGGACCCCCCAGGAGGTGCAGTTAGGGATCTACCTTCAGATG
 CGGTCTGAAGATACTGAAGCAAGGATCCCTGCACATCTCCAGGACACGGACAACCTC
 TGTGCACTCCACAACATCTCCTACCACCTCCTGCTGGTCCCCATTACCCACAAAAAA
 GTTCTTGGCGGTGGGCTGTCATCAGTGCAGACGACCACGTGGATAAACCTCCAGACA
 CGCTGCAGTCCTCTTCAGCAGTCATCAGAGGAGGAGCTGCAGGAGATGGTGGTGGTG
 GTGCACCTGGCAGATGCAGACCCCCATCTGAATGCCAGGTGGCCGCGACATCAGCCA
 TAGGTTGCTCACCAACATCTCCTGGCCGGCTCGTGTATCCATACTCCCCATGAGT
 TTTACCCAACCCCTGGAAGGCCTCAAGAGAAACTACAACGACCCAGAGGAGGGGTGAAG
 TTCAGGTCCAAGCAGAACGTGGATTACGCCCTCCTCACCTTGCTGCCAACCTTC
 CTCCCTACTACTTGATGATTGAGGATGACGTGTGGTCTGCCAAGTCCTCTTCACTGCCA
 TCCGCAAAGCTGTGGCCTCCAGGAAGGCTCCAACGGCCACCCCTGAGTTCTCCAAG
 CTGGGCTACATCGGTAAGCTTACCGCTCCAGTGACCTTCCTCGCTGGCTCGCTTCCT
 CCTCCTCTTCTACCAGGAGATGCCCTGTGACTGGCTGCTGACCCACTCCGCCCTCTGC
 TCACCCAGAAGGATGTGATCCGCTTCAGCCTCCCTCTCCAGCACATGGCCTCTA
 CTCCTCCCTCCAAGGTACCGCTCAACCGACTGGAGGATGACGAGTTTCAGGCTGATGCCA
 TGGACCTTCCAGACAACCCGCCAGCAGCCCTGTCACCAACATGGTTGTCTTGAGAAC
 TATGAGCCCTCCAAGGCTTACAGCACAGCAAGGGGTATTCTGGGGAAAAACCCAGC
 AGTTGGCAGCATTCTCCATTGTCTCCACCAACCAGGCCGTGTCACCCGCGTCCGGG
 TGCAGACGGGATCCAGTGAGGCCCTGGGACTTCTGCTGATGCCGAGACTGCTCTGTGACCC
 GGCGGGGGCGGCGGGCTGATGCCGAGACTGCTCTGTGACCCACTGTGGCACCTT
 TGAGAAAGGAACTTAGAGTGGCGGGGCTGGAGAAGGAAATGCCAACCCCTGTGGAGT
 GCGTGAGGATCCGGGTGACCCAGAGCCAGAGTGAGTGGCTCATCATCCAGAGCATTGGT
 ATTTGGACTGCAGGCACCTGACCAGGGCTGTGATGGGTGACCAACTGTGGCACCTT
 CTCTAAGAAGAGCTTATTTCTCAGTCCCTTTTCAGTGGGAATTAAATTATTCA
 GTCAAACGGTCCCTGCTGTAACGTAGAGGGGTGGCAGGGCAGCTGCGGGGTCTGCT
 TCCTGCACGGAGGTGGACGGGTTGGCTGTAGGGCCACTGTGCTGCACCAACTGGGG
 GATGCTGCAGAAAGCAGTGCCAGCCCCAGGCTGCAGCCTACGCCCATCAGTATGGG

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Figure 44 (Cont.)

GAAAGTGTGGACAGGCAGCTCTGCATACGCTTGTGTCTGATGGAGTGCCAGTTTC
GTGCTCCAAGCAGAGTCCTGCTCCTTGTACCCCAGTGCCCTTGTGATGCTTCCTTA
TGCCCTGACTCAGCTAATTAAATTAAAAACGGTGAGTCAATT

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Figure 45

Homo sapiens GnT-IX N-Acetylglucosaminyltransferase IX (GnT IX)

MITVNPDGKIMVRRCLVTLRPFRLFVLGIGFTILCFLMTSLGGQFSARRLGDSPFTIRT
 EVMGGPESRGVLRKMSDLLELMVKRMDALARLENSSELHRAGGDLHFPADRMPGAGLM
 ERIQAIAQNVDIAVKVDQIILRHSSLHSKVSEGRDQCEAPSDPKFPDCSGKVEWMRA
 RWTSDPCYAFFGVGDTECSFLIYLSEVEWFCPPLPWRNQTAQRAPKPLPKVQAVFRSN
 LSHLLDLMGSKESLI FMKKRTKRLTAQWALAAQRLAQKLGATQRDQKQILVHIGFLTE
 ESGDVFSPRVLKGGLGEMVQWADILTYVLGHGLRVTVSLKELQSNLGVPPGRGSCP
 LTMPPLPDFDLYTDYHGLQQMKRHMGLSFKKYCRIRVIDFGTEPAYNHEEYATLHGYR
 TNWGYWNLNPKQFMTPHTPDNSFMGFVSEELNETEKRLIKGGKASNMNAVYVGKEASI
 WKLQGKEKFLGILNKYMEIHGTVYYESQRPPPEVPAFKVNHGLLPQPEFQQLLRKAKLF
 GFGFPYEGPAPLEAIANGCIFLQSRFSPPHSSLNHEFFRKPKTSREVFSQHPYAENFIG
 KPHVWTVDYNNSEEFEAAIKAIMRTQVDPYLPYEYTCEGMLERIHAYIQHQDFCRAPDP
 ALPEAHAPQSPFVLAPEAHLNTHLEWARNTSLAPGAWPPAHALRAWLAVPGRACTDTCLDHG
 LICEPSFFPFLNSQDAFLKLQVPCDSTESEMNHLYPAFAQPGQECYLQKEPLLFCAGS
 NTKYRRLCPCRDFRKGQVALCQGCL

Homo sapiens GnT-IX mRNA for N-Acetylglucosaminyltransferase IX, complete cds; accession number AB109185.1

CTGCTCGCACCAACAAGTTGAACAATGATCACCGTCAACCCCGATGGGAAG
 ATAATGGTCAGAACAGATGCCCTGGCACCCCTGAGACCCCTTCGGTTTTGTCT
 GGGCATCGGCTTCTCACTCTGCTTCCTGATGACGTCTGGAGGGCCAGT
 TCTCGGCCCGGCCCTGGGGACTCGCCATTACCATCCGCACAGAAGTGAT
 GGGGGGCCCGAGTCCCCGGCGCTCTGCGCAAGATGAGCAGCTGCTGGAG
 CTGATGGTGAAGCGCATGGACGCACTGGCCAGGCTGGAGAACAGCAGTGAG
 CTGCACCAGGGCCGGCGGACCTGCACCTTCCCAGACAGGATGCCCTGACAT
 GGGCGGCCTCATGGAGCGGATCCAGGCTATTGCCAGAACGTCTCCGACAT
 CGCTGTGAAGGTGGACCAGATCCTGCGCACAGTCTGCTCTGCACAGCAAG
 GTGTCAGAACGGCCGGCGGGACCAAGTGTGAGGCACCCAGTGACCCCAAGTCC
 CTGACTGCTCAGGGAAAGGTGGAGTGGATGCGTGCCGCTGGACCTCTGACCC
 CTGCTACGCCCTTGGGGTGGACGGCACCGAGTGCTCCCTCATCTAACC
 TCAGTGAGGTGAGTGGTCTGCCCTGGAGGAACCAGACGGC
 TGCCCAGAGGGACCCAAGCCCCCTCCCCAAAGTCCAGGCAGTTCCGAAGC
 AACCTGTCCCACCTCTGGACCTGATGGGCAGCGGGAAAGG
 AGTCCCTGATCTTCAATGAAGAACGGGACCAAGAGGGCTCACAGCCCAGTGGCGCT
 GGCTGCCAGCGCCTGGCACAGAACGCTGGGGCACCCAGAGGGACCAGAACAG
 ATCCTGGTCCACATCGGCTTCCCTGACGGAGGAGTCCGGGACGTGTTCAGCCCTC
 GGGTCCTGAAGGGCGGGCCCTAGGGGAGATGGTGCAGTGGCGGACATTCTGAC
 TGCACTCTATGTCCTGGGCCATGGCCTGCGGGTCACAGTCTCCCTGAAGGAGCTG
 CAGAGTAACCTAGGGTACCGCCAGGCCGGGAAGCTGCCCTGAGCAGATGAAGCGGCA
 TGCCCTTCGACCTCATCTACACCGACTACCACGCCCTGAGCAGATGAAGCGGCA
 CATGGGACTCTCCTCAAGAAGTACCGGTGCCGAATCAGGGTCATGACACCTTC

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Figure 45 (Cont.)

GGGACGGAACTCGGTACAACCACGAGGAGTACGCCACGCTGCACGGCTACCGGA
CCAACTGGGCTACTGGAACCTCAACCCAAAGCAGTCATGACCATGTTCTCA
TACCCCCACAACCTCTCATGGGCTTGTCCGAGGAGCTAACGAGACGGAG
AAGCGGCTCATCAAAGCGGCAAGGCCAGAACATGCCGTGGTACGGCAAGG
AGGCGAGCATCTGGAAGCTCCAGGGAAAGGAGAAGTCTGGCATCCTGAACAA
ATACATGGAGATCCATGGCACCGTGTACTACGAGAGCCAGCGCCCCCGAGGTG
CCAGCCTTGTGAAGAACCAACGGCCTTACCGCAGCCTGAGTTTACAGCAGCTGC
TGCAGGCAAGGCCAAACTCTCATGGGTTGGCTTCCCCTACGAGGGCCCCGCCCC
CCTGGAGGCCATGCCAATGGTGATCTCCTGCAGTCCCCTCAGCCCGCCC
CACAGCTCCCTAACACGAGTTCTCCGAGGCAAGCCCACCTCCAGAGAGGTG
TCTCCCAGCATCCCTACGCCAGGAGAACTTCATGCCAGGCCCCACGTGTGGACAGT
CGACTACAACAACCTCAGAGGAGTTGAAGCAGCCATCAAGGCCATTATGAGAACT
CAGGTAGACCCCTACCTACCCCTATGAGTACACCTGCAGGGATGCTGGAGCGGA
TCCACGCCATCCAGCACCAAGGACTTCTGCAGAGCTCCAGACCCCTGCCCTACC
AGAGGCCACGCCCGCAGAGCCCTTGTCTGGCCCCAATGCCACCCACCTC
GAGTGGGCTCGAACACCAGCTGGCTCTGGGCTGGCCCCCGCGACGCC
TGCAGGCTGGCTGGCGCTGGCAGGAGGCTGCACCGACACCTGCCTGGACCA
CGGGCTAATCTGTGAGCCCTCTTCTTCCCCTGTGAACAGCCAGGACGCC
CTCAAGCTGCAGGTGCCCTGTGACAGCACCGAGTCGGAGATGAACCACCTGTACC
CGGCCTCGCCAGCCTGGCCAGGAGTGCTACCTGCAGAAGGAGCCTTGCTCTT
CAGCTGCCGGCTCCAACACCAAGTACGCCGGCTCTGCCCTGCCGCGACTTC
CGCAAGGGCCAGGTGGCCTGTGCCAGGGCTGTCTGTGAATCCGCTCTGCCGCC
CTGCCTGGCACCCACGCTGGCTCTCCCTGCCGGAGAAAGCACCAGCAGGTT
C

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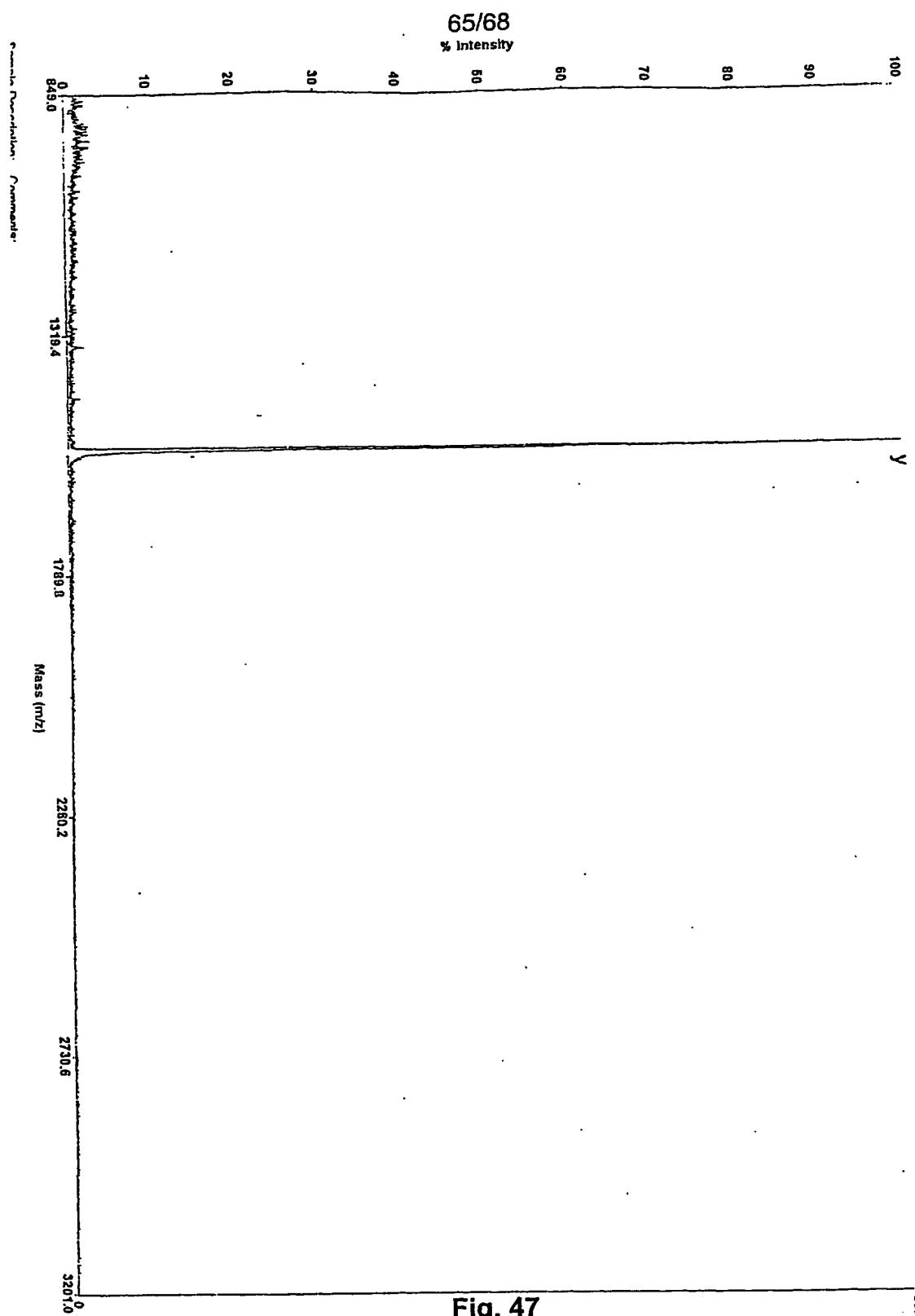
Figure 46

ggcgccgc CAGTTCTCTGCTAGAAGATTGGGTGACTCTCATTCACTATTAGAACCGAGGTATGGGTGGA
C
CAGAGTCC
AGAGGTGTC TTGAGAAAAGATGTCTGACTTGTTGGAGCTGATGGTTAAGAGAAATGGACGCTTGGCTAGATT
G
GAGAACTC
CTCTGAGTTGCACAGAGCTGGTGGTACTTACACTTCCCTGCTGACAGAAATGCCACCAGGTGCAGGTTGA
T
GGAGAGAA
TCCAAGCTATCGCACAGAACGTTCTGACATTGCTGTTAAGGTTGACCAAATCTGAGACACTCTTGCTG
T
TGCAC TCT
AAGGTTCTGAGGGTAGAAGAGATCAATGTGAGGCTCCTCTGATCAAAGTTCCCTGACTGTTCTGGTAA
G
GTTGAATG
GATGAGAGCAAGATGGACTTCTGACCCATGTTACGCTTCTCGGTGGACGGTACTGAGTGGTCTTCT
T
GATTTACT
TGTCTGAAGTTGAGTGGTTCTGTCCACCTTGCCTGGAGAAACCAAACCGCTGCTCAAAGAGCACCAAAG
C
CTTGCCA
AAGGTTCAAGCTGTCTTCAGATCCAATTGTCACTTGCTGGACTTGATGGTTCTGGTAAGGAGTCTT
G
ATCTTCAT
GAAGAAGAGAACTAAGAGATTGACTGCTCAATGGGCTTGGCTGCACAGAGATTGGCACAGAACGTTGGTG
C
TACTCAAA
GAGACCAGAAGCAAATCTGGTTCACATCGTTCTGACTGAGGAGTCTGGTACGTCTGGTACCGTCTCTCCAAGA
G
TCTTGAAG
GGTGGTCCATTGGGAGAGATGGTCCAATGGGCTGACATCTGACTGCTTGTACGTCTGGTACGGTT
G
AGAGTTAC
TGTTTCTTGAAGGAGTTGCAATCTAACATTGGGTGTTCCCTCCAGGTAGAGGTTCTGTCCACTTACTATGC
C
TTTGCAT
TCGACTTGATTACACTGATTACACGGTTGCAACAGATGAAGAGACACATGGTTATCTTCAAGAAC
T
ACAGATGT
AGAATTAGAGTCATTGACACTTCCGGTACTGAA CCTGCTTACAACCACGAGGAGTACGCTACTTGCACGG
T
TACAGAAC
TAACTGGGATACTGGAACCTGAAACCCAAAGCAATTGACTATGTTCCCTCACACCCCCAGACAAC
T
CATGGGTT
TCGTTTCTGAGGAGTTGAA CGAGACTGAGAAGAGATTGATTAAGGGTGGAAAGGCTTCTAACATGGCTGTT
G
TCTACGGT
AAGGAGGCTTCTATTGGAAAGTTGCAAGGTAAGGAGAAGTTCTGGAAATCTGAAACAAGTACATGGAGAT
T
CACGGTAC
TGTTTACTACGAGTCTCAAAGACCACCTGAAAGTTCCAGCATTGTCAAGAACCAACGGTTGGCCACAAC
C

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Figure 46 (Cont.)

TGAGTTCC
AACAGTTGTTAAGAAAGGCTAAGTTGTTCATCGGTTTGGTTCCCATACGAGGGACCTGCTCCCTTGAG
G
CTATTGCT
AATGGTTGTATCTTCITGCAGTCCAGATTCTCTCCACCTCACTCTTGAACCACGAGTTCTTCAGAGG
T
AAGCCAAC
TTCCAGAGAGGTCTCTCAACACCCATACGCTGAGAACATTGTAAGCCACACGTTGGACTGTTG
A
CTACAACA
ACTCTGAGGAGGTCGAAGCTGCTATTAAGGCTATTATGAGAACACTCAAGTTGACCCATACTTGCCCTACGAG
T
ACACTTGT
GAGGGTATGTTGGAGAGAATCCACGCATACATCAACACCAAGACTTCTGTAGAGCACCAGACCCCTGCATT
G
CCAGAGGC
TCACGCTCCACAATCTCATTGCTTGGCTCTTAATGCAACTCACCTGGAGTGGCTAGAAAACACTTCTT
T
GGCTCCTG
GTGCATGGCCACCTGCTACGCTTGAGAGCTTGGTTGGCTGTTGGTAGAGCTTGTACTGACACTTGT
T
TGGACCAC
GGTTGATTGTGAGCCATTTCTTCCCATTCTGAACCTCCAAGACGCTTCTTGAAGTTGCAAGTTCC
A
TGTGACTC
TACTGAGTCTGAGATGAACCACTTGTACCCAGCTTGCACAAACCTGGTCAAGAGTGTACTTGCAGAAGG
A
GCCTTTGT
TGTTCTTGTGCTGGTTCTAACACCAAGTACAGAAGATTGTGTCCATGTAGAGACTTCAGAAAGGGTCAA
G
TCGCTTTG
TGTCAAGGTTGTTGTAAgcttaattaa



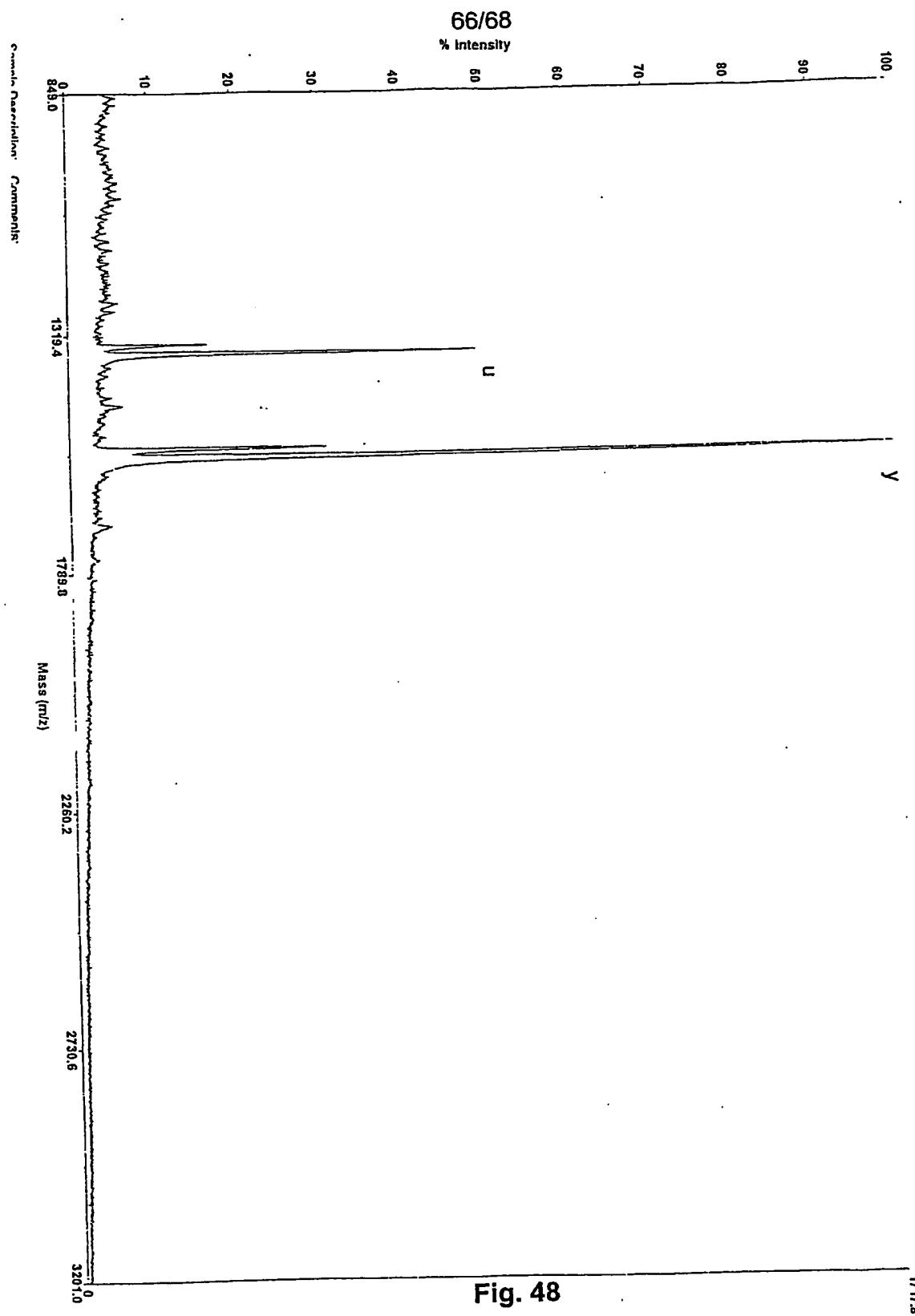


Fig. 48

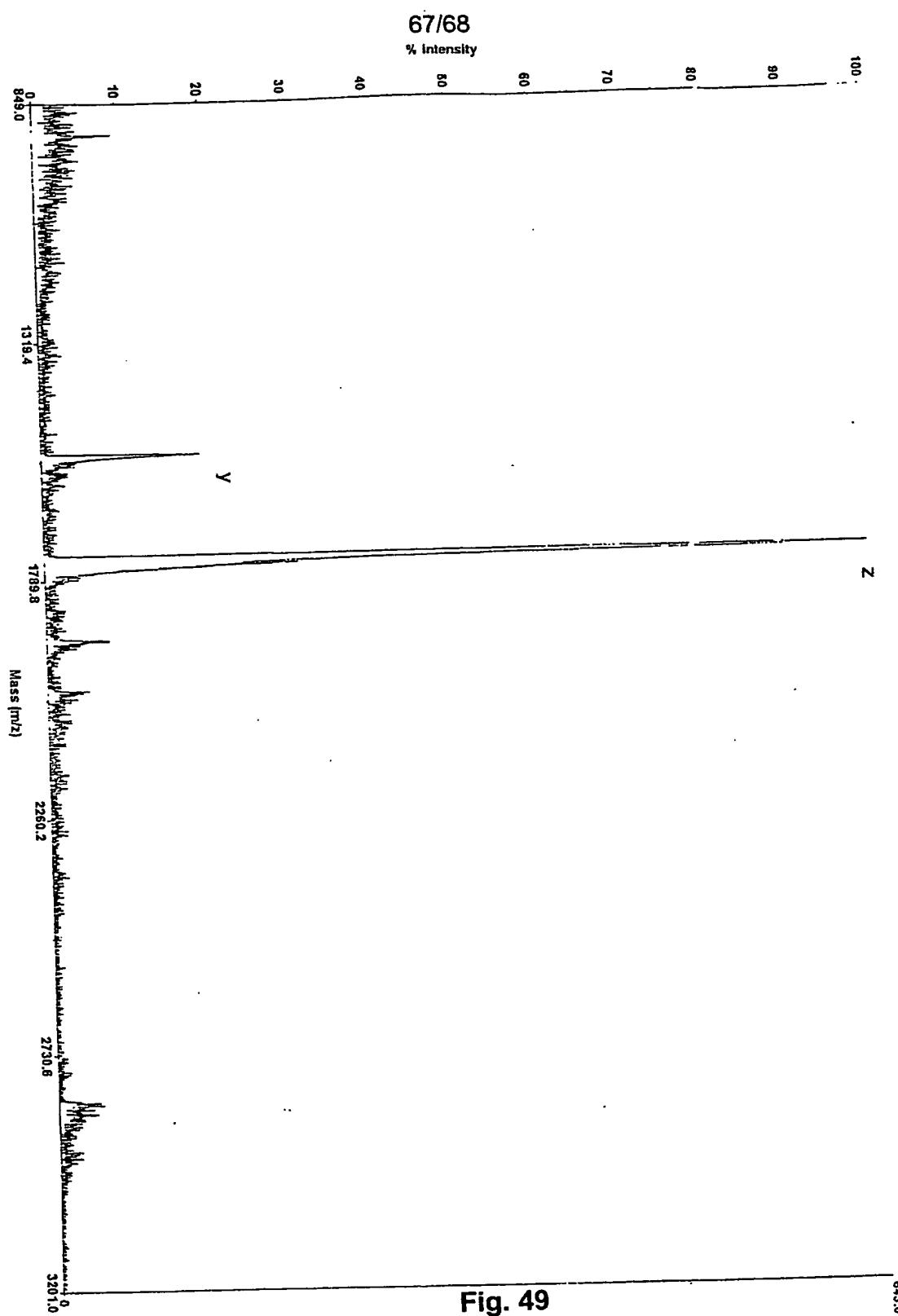


Fig. 49

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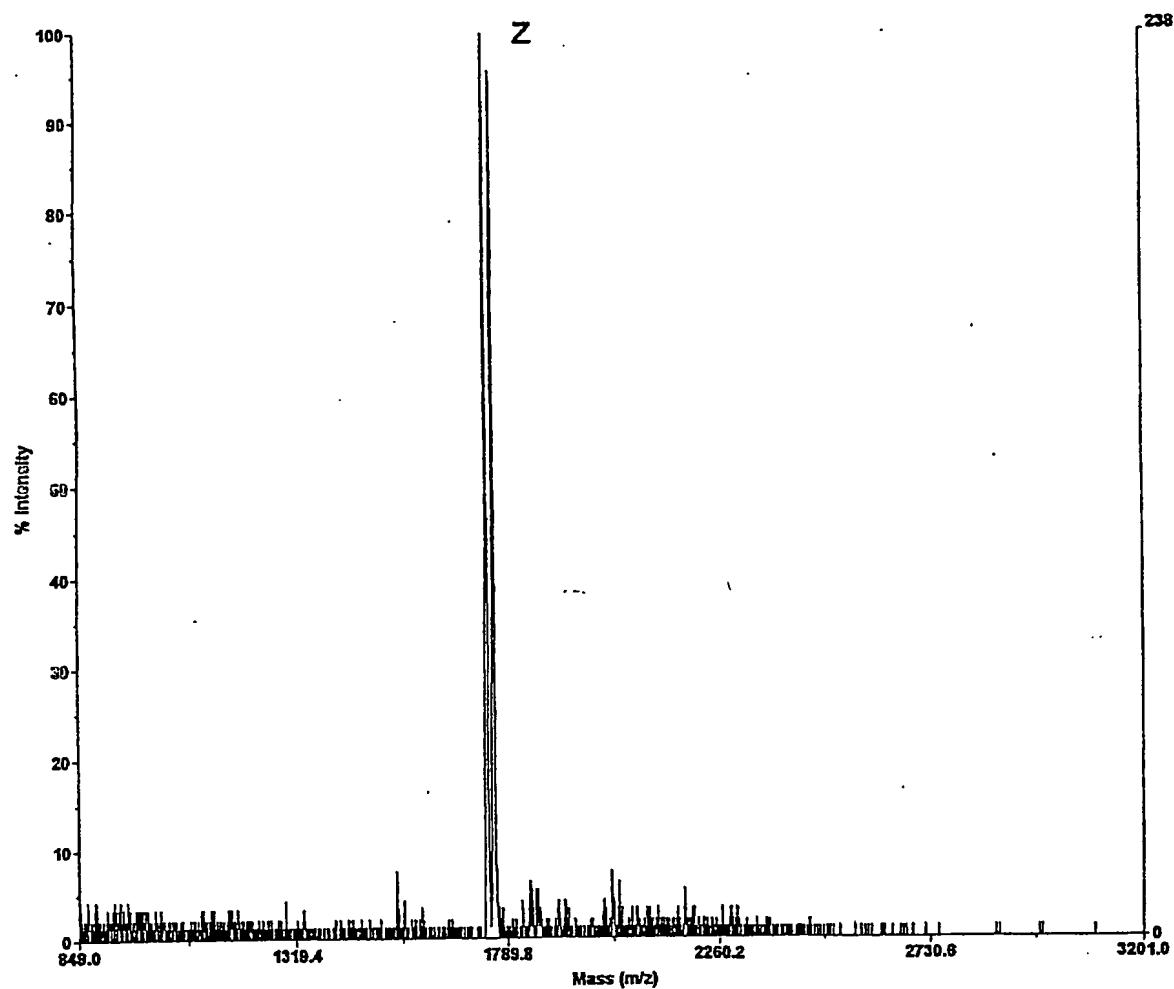


Fig. 50